

**INTERNATIONAL JOURNAL OF UNIVERSAL  
PHARMACY AND BIO SCIENCES****IMPACT FACTOR 4.018\*\*\*****ICV 6.16\*\*\*****Pharmaceutical Sciences****Review Article.....!!!****AN OVERVIEW: MEDICINAL PLANTS****DR. S. SENTHILKUMAR**

KARUR, TAMILNADU, INDIA.

**KEYWORDS:**

Phytochemicals, Flavonoid,  
Drug, Herbal medicine,  
Pharmacology.

**FOR CORRESPONDENCE:****DR. S. SENTHILKUMAR\*****ADDRESS:**

KARUR, TAMILNADU, INDIA.

**ABSTRACT**

India has a long history and strong base of Ayurveda, which is the traditional herbal medical system. Herbal plants play an important role in preventing and treating of human disease. People have been using plant as a traditional medicine for thousand years ago. Plants have been associated with the development of human civilization around the whole world. However, plants are considered as rich sources of phytochemical ingredients which enable to have medicinal value. Medicinal plants are a potential source for the development of new herbal drugs. In the 21th century, the pharmacological effects of medicinal plants have been considered as promising future drug/medicine for the management of health care. In recent years, there has been a resurgence of interest to rediscover medicinal plants as a source of potential drug candidate. Primitive man in search of food and to cope successfully with human sufferings began to distinguish those plants suitable for medicinal purpose from others with definitive pharmacological action. This relationship between plants and man has grown and many plants came to be used as medicines.

**INTRODUCTION:**

Nature is always a golden sign to show the prominent phenomena of coexistence. Natural products from plants, animals and minerals are the basis for treating human disease. Medicinal plants are presently in demand and their acceptance is increasing progressively. Undoubtedly, plants play an important role by providing essential services in ecosystems. Without plants, humans and other living organisms cannot live in a way living should be. Anyway, herbals especially medicinal herbs have constantly acted as an overall indicator of ecosystem health. Medicinal plants have undoubtedly been considered by human beings since ancient times. It can be said that before the history and since the early humans recognized and exploited the plants around them for use as fuel, clothing, shelter and food, they became aware of their properties more or less. Medicinal plants have been transformed into one of the oldest sciences in countries such as China, Greece, Egypt, and India. In ancient Persia, plants were commonly used as a drug and disinfectant and aromatic agent. In fact, the use of medicinal plants for the treatment of diseases dates back to the history of human life, that is, since human beings have sought a tool in their environment to recover from a disease, the use of plants was their only choice of treatment. More than a tenth of the plant species (over 50,000 species) are used in pharmaceutical and cosmetic products. However, the distribution of medicinal plants across the world is not uniform, and medicinal herbs are mainly collected from the wildlife population. Indeed, the demand for wildlife sources has increased by 8%-15% per year in Europe, North America and Asia in recent decades. The term *Medicinal plant* refers to a variety of plants that have medicinal properties. These plants are a rich source of compounds that can be used to develop drug synthesis. The parts of medicinal plants that may be used are different types of seeds, root, leaf, fruit, skin, flowers or even the whole plant. The active compounds in most parts of the medicinal plants have direct or indirect therapeutic effects and are used as medicinal agents. In the body of these plants, certain materials are produced and stored that are referred to as active compounds (Substance), which have physiological effects on the living organisms. Human is mainly dependent on raw plant materials in order to meet medical needs to maintain health and cure diseases. Medicinal plants are used for treatment because they have certain properties, including synergistic actions. The constituents of the plant may interact with each other, and this interaction can be beneficial for both or adverse to either of them or eliminate the harmful effects of both. Plant-derived compounds can dramatically improve hard-to-treat illnesses, such as cancer. Plant components are also characterized by their ability to prevent the development of certain diseases. The toxicity and adverse effects of conventional and allopathic medicines have

also been important factors in the sudden increase in population demands and increase in the number of herbal drug manufactures as well as a reduction in the use of chemical drugs.

Knowing the history of any science is effective in understanding and using that science. Hence, the historical significance of the past and present and future of medicinal herbs will continue to be addressed. In this perspective review, we have highlighted and discussed the history, current challenges, development and future outlook of using medicinal plants and their active compounds.

### **PHYTOCHEMICALS:**

Phytochemicals (from the Greek word phyto, meaning plant) are biologically active, naturally occurring chemical compounds found in plants, which provide health benefits for humans further than those attributed to macronutrients and micronutrients. They protect plants from environmental hazards such as pollution, stress, drought, UV exposure and pathogenic attack are called as phytochemicals. Recently, it is clearly known that they have roles in the protection of human health, when their dietary intake is significant. More than 4,000 phytochemicals have been catalogued and are classified by protective function, physical characteristics and chemical characteristics and about 150 phytochemicals have been studied in detail.

In wide-ranging dietary phytochemicals are found in fruits, vegetables, legumes, whole grains, nuts, seeds, fungi, herbs and spices. Broccoli, cabbage, carrots, onions, garlic, whole wheat bread, tomatoes, grapes, cherries, strawberries, raspberries, beans, legumes, and soy foods are common sources. Phytochemicals accumulate in different parts of the plants, such as in the roots, stems, leaves, flowers, fruits or seeds. Many phytochemicals, particularly the pigment molecules, are often concentrated in the outer layers of the various plant tissues. Levels vary from plant to plant depending upon the variety, processing, cooking and growing conditions. Phytochemicals are also available in supplementary forms, but evidence is lacking that they provide the same health benefits as dietary phytochemicals.

These compounds are known as secondary plant metabolites and have biological properties such as antioxidant activity, antimicrobial effects, modulation of detoxification enzymes, stimulation of the immune system, decrease of platelet aggregation and modulation of hormone metabolism and anticancer property. There are more than thousand known and many unknown phytochemicals. It is well-known that plants produce these chemicals to protect themselves, but recent researches demonstrate that many phytochemicals can also protect human against diseases.

Phytochemicals are not essential nutrients and are not required by the human body for sustaining life, but have important properties to prevent or to fight some common diseases. Many

of these benefits suggest a possible role for phytochemicals in the prevention and treatment of disease, because of this property; many researchers have been performed to reveal the beneficial health effects of phytochemicals. The purpose of the present review is to provide an overview of the extremely diverse phytochemicals presents in medicinal plants.

#### **CLASSIFICATION OF PHYTOCHEMICALS:**

The exact classification of phytochemicals could have been performed so far, because of the wide variety of them. In recent year Phytochemicals are classified as primary or secondary constituents, depending on their role in plant metabolism. Primary constituents include the common sugars, amino acids, proteins, purines and pyrimidines of nucleic acids, chlorophyll's etc. Secondary constituents are the remaining plant chemicals such as alkaloids, terpenes, flavonoids, lignans, plant steroids, saponins, phenolics, flavonoids and glucosides.

#### **BIOLOGICAL ACTIVITIES OF PHYTOCHEMICALS:**

The phytochemicals present in plants are responsible for preventing disease and promoting health have been studied extensively to establish their efficacy and to understand the underlying mechanism of their action. Such studies have included identification and isolation of the chemical components, establishment of their biological potency both by *in vitro* and *in vivo* studies in experimental animals and through epidemiological and clinical-case control studies in man. Study findings suggest that phytochemicals may reduce the risk of coronary heart disease by preventing the oxidation of low-density lipoprotein (LDL) cholesterol, reducing the synthesis or absorption of cholesterol, normalizing blood pressure and clotting, and improving arterial elasticity. Phytochemicals may detoxify substances that cause cancer. They appear to neutralize free radicals, inhibit enzymes that activate carcinogens, and activate enzymes that detoxify carcinogens. For example, according to data summarized by Meagher and Thomson, genistein prevents the formation of new capillaries that are needed for tumor growth and metastasis. The phytochemicals are well understood and more many research has focused on their possible role in preventing or treating cancer and heart disease. Phytochemicals have also been promoted for the prevention and treatment of diabetes, high blood pressure, and macular degeneration. While phytochemicals are classified by function, an individual compound may have more than one biological function serving as both an antioxidant and antibacterial agent.

#### **PHARMACOLOGICAL IMPORTANCE OF MEDICINAL PLANTS:**

Plants use as food and in traditional medicine are more likely to yield pharmacologically active compounds. The medicinal properties of plants have been investigated in the recent scientific

developments throughout the world, due to their potent therapeutic efficacy and antioxidant activities, no side effects and economic viability. Medicinal plants are serving as raw material for drugs which are effective and reasonable health care for people. However, all plants synthesize phytochemicals, which are beneficial for our health as they cannot be synthesized in the human body. Plants are also rich dietary sources of biomolecules, vitamins and minerals which are crucial for maintaining the healthy body.

It has been observed that numerous plants have pharmacological effects due to the presence of metabolites. Plant metabolites are organic compounds include glucose, starch, polysaccharide, protein, lipids and nucleic acid which are beneficial for growth and development of the human body. Plants synthesize secondary metabolites which include alkaloids, flavonoids, saponins, terpenoids, steroids, glycosides, tannins, volatile oils etc., The therapeutic efficacy of plants is because of these secondary metabolites for curing many diseases. Phytochemicals are pharmacologically active compounds. These include alkaloids have an antispasmodic, antimalarial, analgesic, diuretic activities; Terpenoids are known for their antiviral, anthelmintic, antibacterial, anticancer, antimalarial, anti-inflammatory properties; Glycosides are reported for antifungal and antibacterial properties; Phenols and flavonoids have an antioxidant, antiallergic, antibacterial properties etc. and Saponins are reported, to have anti-inflammatory, antiviral, defence activities.

A Chinese Pharmacologist, Youyou Tu, discovered and developed a new herbal antimalarial drug "Artemisinin" from *Artemisia annua* (a sweet Warmwood plant in China). Researchers identified several chemical compounds used in modern medicine, which were derived from plant sources include quinine, digoxin, Aspirin, atropine, and colchicines.

People have been using plants as a medicine without scientific knowledge and proper guidance for thousand years ago. Using plants as medicines it is considered as a natural healing Medical System. It has been scientifically established that every part of plants has medicinal properties include flower, root and stem, leaves, fruits, seed and whole plants. However, it has been observed that some plants are not safe for health because they contain some toxic compounds which show adverse effects in the body.

Herbal medicine is widely practiced in worldwide. For centuries, people have turned to natural remedies to cure common ailments such as colds, allergy, upset stomachs and toothaches and the trend is constantly increasing. Thus, there has been a shift in universal trend from synthetic to herbal medicines, which we can say "Return to Nature" for the prevention of diseases and ailments. Nature has been a source of medicinal plants/ the World Health Organization (WHO) reported that

4 billion people (80% of the world's population) use herbal medicines for some aspect of primary healthcare. Herbal medicine has been recognized by WHO as essential components for primary health care and about 11% of the 252 drugs are derived from plants.

Since time immemorial, human civilization has been used several plants as food, medicine, clothing and shelter. Vegetarian foods contain high amounts of various "Super-nutrients", such as protective antioxidants, phytochemicals, micronutrients, which promote health and protect from disease. Plants have several pharmacological roles such as antioxidant, antiviral, anticancer, antimicrobial, antifungal and antiparasitic. Plants have free radical scavenging molecules, including flavonoids, phenolics, anthocyanins and vitamins, which show antioxidant like activity. It has been reported that the antioxidant property of phytochemicals may be mitigated the oxidative stress in the biological system. Phytochemicals have been reduced the risk of many human diseases include cardiovascular disease, hepato-renal diseases, diabetes, cancers and neurodegenerative disorders. However, several herbal medicines are being derived directly or indirectly from plants that are considered as an important medicine currently in use for curing various human diseases.

The development of plant drug started when development of chemistry, isolation, purification, characterization of plant active compounds. Herbal medicine is effective, lesser side effect, and affordable than the medicines bought from an allopathic medicine. Herbal medicines include herbs, herbal materials herba preparations, and herbal products that contain different parts of plants or other plant materials as active ingredients. It has been well documented that herbal plants and their derivatives play critical role in modern drug development. Medicinal plants are the natural resources in developing of new drugs.

#### **HEAVY METALS IN MEDICINAL PLANTS:**

Medicinal plants are worldwide used in various disciplines of health care systems such as herbal, Ayurvedic, Unani and homeopathic system of medicines in the form of herbs and standardized extracts. According to the advance researches it has been suggested that plants contain secondary metabolites which are not only contain toxic substances (alkaloids), but they are also contaminated with environmental pollutants specially heavy metals, which are very dangerous to all living organisms upon long term exposures. Heavy metals are classified in two main categories i.e. essential and non essential heavy metals. Essential heavy metals (Cu, Cr, Zn, Fe, and Co), are present in very little concentration in the body for the proper functioning of enzyme systems, vitamin synthesis and haemoglobin formation in men and also required for the growth,

development and photosynthesis in plants. On the other hand toxic metals (Pb, As, Cd and Hg) are not needed to perform specific function in the body and they have deleterious effects even at very low concentrations. According to the advanced research which has shown that crude extract of herbal origin can prove fatal for the health because these drugs may contain toxic metabolites or some trace elements which cause damages to health.

Some heavy metals are needed by the body in very low concentration for proper functioning or synthesis of various biomolecules inside the body. These trace elements are called essential heavy metals. Heavy metals are present in the environment which is subjected to bioaccumulation in food chains. Anthropogenic processes, involving the organic manure, synthetic fertilizers, industrial residues and lime upon exposure contribute various amounts of heavy metals to the ecosystem. Heavy metals cannot be decomposed by the microorganisms as they are resistant to biodegradation. During cultivation process addition of synthetic fertilizers or lime is one cause which leads to enhance the concentration of heavy metals to the soil form where they are absorbed by the plants. Industrial exhaust, vehicle exhaust and careless handling of wastes are also the main cause of producing heavy metals which may pollute the environment. This contamination of heavy metals in medicinal plants remains continuous during storage and transportation process of plants where these are exposed to environmental pollution and heavy metals. The major source of accumulation of heavy metals in human organs is because these medicinal plants are not only used as folk medicines and food supplements, but many of them are utilized as condiments in daily routine. WHO/FAO has put forward this critical issue and analyzed heavy metals in the herbal medicines along with other necessary chemical, biological, and environmental analysis in their guidelines and also suggested the absorption, elimination, and toxic profiles of heavy metals. Although herbal drugs are utilized on a large scale by the people but these may be contaminated with heavy metals beyond their recommended level. The biological effects caused by these heavy metals necessitate the need to assess the heavy metal level of medicinal plants before they are utilized or taken for medicinal purposes.

#### **EXAMPLE:**

#### **MEDICINAL PLANTS HAVING ANTICANCER POTENTIAL:**

##### **1. *ADIANTUM VENUSUTUM*:**

*Adiantum venusutum* (Adiantaceae) is traditionally very useful in treating tumour. The phytochemicals, terpenoids, phytosterols, flavanoids and saponins are obtained from the petroleum ether and ethanolic extract of leaves and stem of *Adiantum Venusutum*: and screened

for the anticancer activity on Ehrlich Ascites Carcinoma in animals by using dose of 150-250mg/kg. A triterpenic ether, lanost-20(22)-en-3, 19-ether, named adiantulanostene ether was isolated from *A. Venustum*.

## 2. *ABELMOSCHUS MOSCHATUS*

The antiproliferative activities of ethanolic and aqueous extracts of *Abelmoschus moschatus* (*Malvaceae*) seed (AMS) and *Abelmoschus moschatus* leaf (AML) against two human cell lines- Colorectal adenocarcinoma (COLO-205) and retinoblastoma (Y79) were investigated. Flavanoids are responsible for the antiproliferative activity of the extract.

## 3. *ASPIDOSPERMA TOMENTOSUM*:

The antiproliferative activity of terpenoids and alkaloids obtained from crude dichloromethane (CHD) and crude hydroalcoholic extract (CHE) of *Aspidosperma tomentosum* (*Apocynaceae*) twigs and aerial part was tested against five human cell lines: K562 (leukemia), MCF7 (breast), NCIADR (BREAST expressing the multidrug resistance phenotype), NCI460 (lung) and UACC62 (melanoma), in a concentration-dependent way.

## 4. *ANEMOPSIS CALIFORNICA*:

Three different extract conditions (aqueous EtOH and EtOAc) of four different parts (Bracts, leaves, roots and stems) of *Anemopsis californica* (*Saururaceae*) were evaluated for their effect on the growth and migration of human colon cancer cells, GCT-8, and the breast cancer cell lines Hs 578T and MCF-7/AZ.

## 5. *ALANGIUM SALVIIFOLIUM*:

The phytoconstituents like sterols, glycosides, saponins, carbohydrates, alkaloids, flavanoids, tannins, proteins and triterpenoids are identified in the ethanolic, chloroform, alcohol and distilled water extract of *Alangium salviifolium* (AS) [*Cornaceae* (*Alangiaceae*)] seeds, flowers, roots and leaves showed significant antitumor activity against Ehrlich Ascites Carcinoma (ECA) in mice at the doses of 10mg/kg body weight intraperitoneally.

## 6. *SVOTUD VLSMUD*:

Several essential oils and bioactive compounds like  $\beta$ -asarone (46.78%), linalool (0.41%), farnesol (11.09%), methyleugenol (6.10%),  $\alpha$ - and  $\beta$ -pinene (both 0.06%), [E]-caryophyllene (0.11%),  $\beta$ -elemene (0.39%), ocimene (0.7%), aromadendrene (0.26%), camphor (0.03%), from *Acorus calamus* (*Araceae*) were identified for the antitumor activity and assayed in MDA\_MB-435S and Hep3B cell lines. The plant possesses anti-

tumor properties at the dose of 30 µg/ml. Sesquiterpenes, phenylpropanoid etc. are isolated from of the ethanolic extract of *A. Calamus* rhizomes and were evaluated for anticancer activity.

7. **ANTIARIS AFRICANA:**

The methanol extract from the stem bark of *Antiaris Africana* (Moraceae) as well as compounds isolated and identified as betulinic acid, 3β-acetoxy-1β,11 α-dihydroxy-olean-12-ene, ursolic acid, oleanolic acid, strophanthidol, periplogenin, convallaroxin, strophanthidinic acid, methyl strophanthinate, and 3, 39-dimethoxy-49-O-β-d-xylopyronosyl ellagic acid, were tested for their anticancer activities against DU-145 and Hep G2 cells.

8. **AMOORA ROHITUKA:**

Amooranin (AMR), a triterpene acid isolated from the petroleum ether, dichloromethane, and ethanol fraction of stem bark of *Amooea tohituka* (Meliaceae). The mechanism of cell death associated with AMR cytotoxicity in human mammary carcinoma MCF-7, multidrug resistant breast carcinoma MCF-7/TH and breast epithelial MCF-10A cell lines. AMR IC<sub>50</sub> values ranged between 3.8-6.9 µg/ml among MCF-7, MCF-7/TH and MCF-10A cells. The induction of apoptosis in AMR treated cells was accompanied by the elevation of total caspase and caspase-8 activities.

9. **AMEBIA NOBILIS:**

Beta-do, etju; acru; sjolpmom from the root of *Arnebia nobilis* (Boraginaceae) possess anti-cancer activity by blocking of cell cycle progression in G1 phase, decreased expression of Cyclin D, CDK 4 and PCNA, inhibition of bcl2 expression at transcriptional level and induction of caspase-3 activity. Arnebin isolated from the roots of *A. Nobilis*, inhibits rat Walker carcinoma, but activity was not found in the leaves and the stem.<sup>29</sup>

10. **AESCULUS HIPPOCASTANUM:**

Recent studies *in vivo* and *In vitro* indicate that aescin (β-escin) has significant antitumor activities. β-escin from *A hippocastanum* (Sapindaceae) inhibited chemically induced colon carcinogenesis in rats, and *in vitro* exhibited cytotoxicity at 30 µmol/l. Or above concentrations in colon cancer cell lines.

11. **AEGLE MARMELOS:**

The hydroalcoholic leaf extract of *Agele marmelos* (AME) (Rutaceae) was studied in the Ehrlich ascites carcinoma bearing Swiss albino mice and anticancer activity is due to

presence of skimmianine in the extract. Butylp-tolyl sulfide, 6-methyl-4-chromanone and 5-methoxypsoralen were isolated from the extracts of *Aegle marmolos* is able to inhibit the *in vitro* Proliferation of human tumor cell lines, including the leukemic K562, T-lymphoid Jurkat, B-lymphoid Raji, erythroleukemic HEL, melanoma Colo38, and breast cancer MCF7 and MDA-MB-231 cell lines.

#### 12. **ALLIUM SATIVUM:**

The anti-cancer activity of garlic-derived organosulfur compounds (OSCs) origination from aqueous extract (GAE) of aerial part and bulbs of *Allium sativum* (*Alliaceae*) cause cell cycle arrest, inhibit cancer (HeLa) cell line and generate reactive oxygen species (ROS). In addition inhibition of lymphocyte proliferation was observed in the presence of higher concentrations of GAE. The garlic derivative *S*-allylmercaptocysteine (SAMC) inhibits growth, arrests cells in G<sub>2</sub>-M, and induces apoptosis in human colon cancer cells by thus arrestin cells in mitosis and triggering JNK1 and caspase-3 signaling pathways that lead to apoptosis. Allicin, a major component of garlic has antitumoral activity in L5178Y lymphoma bearing mice.

#### 13. **BIOHYTUM SENSITIVUM:**

An alcoholic extract to *Biophytum sensitivum* (*Oxalidaceae*) leaves was also found to be cytotoxic towards L929 cells in culture at a concentration of 0.1 mg/ml. The extract was 100% toxic at a concentration of 0.5 mg/ml to Dalton's lymphoma ascites (DLA) and Ehrlich ascites carcinoma (EAC) cells.

#### 14. **BETULA UTILIS:**

Betulinic acid (3 $\beta$ -Hydroxy-lup-20(29)-en-28-oic acid), pentacyclic lupine-type triterpene is obtained from the chloroform bark extract of *Betula utilis* (*Betulaceae*). It exhibits selective cytotoxicity against several melanoma-derived cell lines by means of inducing apoptosis in cells irrespective of their p<sup>53</sup> status.

#### 15. **CUSCUTA REFLIXA:**

The antitumor activity of the chloroform and ethanol extracts of whole plant of *Cuscuta reflexa* (*Convolvulaceae*) was evaluated against Ehrlich ascites carcinoma (EAC) tumor in mice at doses of 200 and 400mg/kg body weight orally, respectively. Anticancer activity of the water extract of *C. reflexa* was analysed on Hep3B cells.

**16. CAESALPINIA BONDNCELLA:**

The methanol extract of *Caesalpinia bonducella* (*Caesalpiaceae*) Leaces (MECB) were evaluated for antitumor activity against Ehrlich ascites carcinoma (EAC)-bearing Swiss albino mice. The extract was administered at the doses of 50, 100, and 200mg/kg body weight per day for 14 days after 24h of tumor inoculation. After the last dose and 18 h fasting, the mice were sacrificed. Life span of EAC-bearing hosts, hemarological profile, and biochemical parameters such as lipid peroxidation (LPO), glutathione content (GSH), superoxide dismutase (SOD), and catalase (CAT) activities.

**17. CASSIA FISTULA:**

Effects of methanolic extract (ME) of *Cassia fistula* (*Fabaceae*) seed on the growth of Ehrlich ascites carcinoma (EAC) and on the life span of tumor bearing mice were studied. ME treatment showed an increase of life span, and a decrease in the tumor volume and viable tumor cell count in the EAC tumor hosts<sup>48</sup>.

**18. CASSIA TORA:**

Methanolic leaf extract of *Cassia tora* (*Fabaceae*) (CTME) was evaluated for antiproliferative activity of with Cisplatin using human cervical cancer cells (HeLa). The plant extract induced a marked concentration dependent inhibition on proliferation, reduced DNA content and apoptosis in HELa. Phenolic compounds are responsible for the antiproliferative activity.

**19. CLEOME GYNANDRA:**

The anticancer activity of the methanol extract of whole plant of *Cleome gynandra* (*Capparidaceae*) (MECG) was evaluated in Swiss albino mice against Ehrlich Ascites Carcinoma (EAC) cell line at the doses of 200 and 400 mg/kg body weight intrperitoneally.

**20. CENTELLA ASIATICA:**

The crude extract (CE) of *Centella asiatica* (*Apiaceae*) inhibited the proliferation of hte transformed cell lines significantly. 50% effective doses was foun don 17 and 22 µg/ml for ehrlich ascites tumour cells (EAC) and dalton's lymphoma ascites tumour cells (DLA), respectively. Oral administration CE retarded the development of solid and ascited tumours and increased the life span of these tumour bearing mice. Asiatic acid (AA) is a pentacyclic triterpene found in *Centella asiatica* decreased viability and induced apoptosis inhuman melanoma SK-MEL-2 cells in time-and dose-dependent manner.

**21. COLA NITIDA:**

The potential anticarcinogenic effect of cola nut methanol extract on human breast cancer cell lines. MCF-7 was investigated. MCF7 cells treated with 80 µg/ml cola nut extract showed an increase of 8.29% in population of apoptotic cells with a concomitant decrease in the percentage of cells in the S and G2/M phase of cell cycle compared to DMSO-treated control cells.

**22. CIRSIUM JAPONICUM:**

Potential antioxidant activities of methanol and water extracts of *Cirsium japonicum* (*Asteraceae*) (CI) leaves and roots showed a concentration-dependent reducing power, ranging from 0.228 to 1.072 (0.1-0.5 mg/mL), as well as a high DPPH free radical-scavenging activity ( $EC_{50}=40.25$  µg/mL). Anticancer activity of *C. Japonicum* In the S180 and H22 mice was separated and purified with several chromatographic techniques and two flavone compounds, pectolinarin and 5,7-dihydroxy-6,4'-dimethoxyflavone, were isolated and greatly inhibited cancer cell growth.

**23. CEPHALOTAXUS HARRINGTONIA:**

Several phytochemicals obtained from the chloroform extract of leaves and stems of *Cephalotaxus harringtonii* (*Cephalotaxaceae*) are cephalotaxine and its antitumor esters (harringtonine, isoharringtonine and honoharringtonine) in both callus and medium. It shows significant activity against experimental P388 leukemia and L-1210 leukemia in mice. A new alkaloid deoxyharringtonine (80%), and isoharringtonine, with significant antileukemic activity has been isolated from *C. harringtonia*.

**24. CROCUS SATIVUS:**

Crocin, crocetin, picrocrocin and safranal are isolated from 80% ethanol extracts of the whole plant of *Crocus sativus* (*Iridaceae*) have been reported to inhibit cell growth of human tumor cells, P38B, S-180, EAC and DLA tumor cells *In vitro*. Saffron can prevent chemically induced skin carcinogenesis in Swiss Albino mice. Crocin inhibits proliferation, nucleic acid synthesis and induces apoptosis in the human tongue squamous cell Carcinoma Cell Line, Tea 8113.

**25. CINNAMOMUM ZELLYANICUM**

The anti-neoplastic activity in the aqueous extract of cinnamon bark (ACE-C) in cervical cancer cell line was reported through increase in intracellular calcium signaling as well as loss of mitochondrial membrane potential, SiHa due to downregulation of MMP-2

expression. In addition, *C. zeylanicum* (Lauraceae) aqueous extract significantly ( $P < 0.01$ ) induced 20 and 37% thymic cells lymphoproliferation at 62.5 and 125  $\mu\text{g/ml}$  respectively.

#### 26. *CONYZA CANASENSIS*:

Methanol extract of the roots of *Conyza Canadensis* (Asteraceae) led to the isolation of two new dihydropyranones named conyzapyranone A and B and the known 4Z, 8Z-matricaria- $\gamma$ -lactone, 9,12,13,-trihydroxy-10 (E)-octadecenoic acid, epifriedelanol, friendeline, taraxerol, simiarenol, spinasterol, stigmasterol,  $\beta$ -sitosterol, and apigenin. The isolated compounds were demonstrated to exert considerable cell growth-inhibitory activity against human cervix adenocarcinoma (HeLa), skin carcinoma (A431), and breast adenocarcinoma (MCF-7) cells. The antitumor activities of novel antitumor, duocarmycins (DUMs), A, B1, B2 C1, and C2 obtained from the extract of *Conyza Canadensis* were examined against human and murine tumor cells, also inhibited the growth of adriamycin (ADM)-resistant lines of human nasopharynx carcinoma KB cells and breast carcinoma MCF-7 cells as well as their sensitive lines.

#### 27. *CLAUSENA LANSIUM*:

An investigation of *C. lansium* seed extract led to the isolation and identification of two new amides, clasue-nalansamide A and clausenalansamide B along with three known human cancer cell lines, KB, MCF7 and NCIH187. The phenolic and flavonoids content of the extract is responsible for antiproliferative activity.

#### 28. *CROTON MACROBOTRYS*:

n-hexane, dichloromethane and methanol extracts of the whole plant of *Croton macrobotrys* (Euphorbiaceae) were evaluated for their *In vitro* antiproliferative activity on cell lines, 786-0 (kidney), HT-29 (colon), K562 (leukemia), NCI-ADR/RES (drug resistant ovary), NCI-H460 (lung), MCF-7 (mammary), PC-3 (Prostate), OVCAR-3 (ovary), U251 (glioma) and UACC-62 (melanoma).

#### 29. *DRACOCEPHALUM TANGUTICUM*:

A chloroform extract of the whole plant of *Dracocephalum tanguticum* (CEDT) (*Lamiaceae*) was found to contain high content of saponin or saponin (53.7%). 90  $\mu\text{g/ml}$  dose of CEDT was effective in anticancer activity on T98G glioblastomas cells by induction of p21.

**30. DIOSCOREA COLLETTII:**

Methyl protodioscin (MPD) extracted from *Dioscorea collettii* (*Dioscoreaceae*) has anti-proliferative effect on the HepG2 cells and cytotoxicity against most cell lines from solid tumors. It results in G2/M arrest and apoptosis in HepG2 cells. Three effects were attributed to down-regulation of Cyclin B1 and the signaling pathways leading to up-regulation of Bax and down-regulation of BCL2, suggesting that methyl protodioscin may be a novel anti-mitotic agent. MPD, ranging from 2.4 to 20 mM, significantly inhibited growth of HepG2 cells in a time- and dose-dependence manner.

**31. EMILIA SONCHIFOLI:**

The anticancer efficacy of n-hexane extract of whole plant of *Emiliasonchifolia* (*Asteraceae*) was determined in mice using Dalton's lymphoma ascetic (DLA) cells. The hexane extract was found to be most active in a concentration- and time-dependent manner; it induced membrane blebbing, nuclear condensation, DNA ladder formation, and formation of apoptotic bodies which are characteristic to apoptotic cell death.

**32. EUPHORBIA HIRTA:**

The antiproliferative activity of the methanol extract of leaves of *Euphorbia hirta* (*Euphorbiaceae*) on HepG2 cells from human epithelioma of larynx was investigated. The dose, 625 µg/ml of methanol extract showed the antiproliferative activity. The acetone root extracts of *E. Hylonoma* were investigated for 1,1-diphenyl-2-picrylhydrazyl (DPPH) free radical, superoxide anion and hydroxyl radical scavenging activity and cytotoxic activity against the human hepatoma cell line SMMC-7721, human cervix epitheloid carcinoma cell line HeLa and the human gastric cancer cell line SGC-7901.

**33. EMBLICA OFFICINALIS:**

The efficacy of methanolic extract of *Embllica officinalis* (*Phyllanthaceae*) fruits polyphenol fraction (EOP) on the induction of apoptosis in mouse and human carcinoma cell lines (Dalton's Lymphoma Ascites (DNA) and CeHa cell) at 200 µg/ml dose and its modulatory effect on N-nitrosodiethylamine (NDEA) induced liver tumors at 60 mg/kg. In rats were determined. Aqueous extract of *E. Officinalis* was found to be cytotoxic to L929 cells, ascites and solid tumours in mice induced by DLA cells in a dose dependent manner.

**34. GMELINA ASIATICA**

The effect of lignans and flavonoids obtained from the ethyl acetate extract of *Gmelina asiatica* (*Lamiaceae*) roots (EGAR) on estrogen receptor-positive (MCF-7) and negative (MDA-MB-231) human breast cancer cell lines were studied.

**35. HERACLEUM PERSICUM:**

Antitumor activity was found in the methanol and petroleum ether extract from the root and fruits of *Heracleum persicum* (*Apiaceae*) and had inhibition effects on *Agrobacterium tumefaciens* which induced crown gall tumor on potato disk.

**36. HIBISCUS MUTABILIS:**

A hexameric 150-kDa lectin was isolated from dried *Hibiscus mutabilis* (*Malvaceae*) seeds using a chromatographic protocol that involved ion exchange chromatography on SP-Sepharose, and gel filtration on Superdex 75 and Superdex 200. The hemagglutinating activity of the lectin, which was stable at pH 4-7 and up to 50 degrees C, could be inhibited by 25 mM galactonic acid. This is the first report of a galactonic acid-binding lectin. It potently inhibited HIV-1 reverse transcriptase with an IC<sub>50</sub> of 0.2 microM.

**37. HEDYOTIS DIFFUSA:**

The cellular effects of the ethanol extract of whole plant of *Hedyotis Diffusa* (EEHDW) (*Rubiaceae*) in the HT-29 human colon carcinoma cell line inhibited the growth of HT-29 cells, demonstrating EEHDW-induced cell morphological changes, collapse of mitochondrial membrane potential, activation of caspase-9 and caspase-3, and increase of the ratio of pro-apoptotic Bax to anti-apoptotic Bcl-2, suggesting that the HT-29 cell growth inhibit activity of EEHDW. The isolated splenocytes from *H. Diffusa* extract-treated leukemin mice demonstrated an increase of T- and B-cell proliferation *in vivo*.

**38. INDIGOFERA LINNAEI:**

Methanol extract of *indigofera linnaei* (MEIL) of family *fabaceae* was investigated for antitumor activity against transplantable tumors and human cancer cell lines HeLa, Hep-2, HepG-2, MCF-7, HT-29, Vero and NIH 3T3 cells. At the dose of 200 and 400 mg/kg, significantly increase the mean survival time, exerted a protective effect on the hemopoietic system and significantly reduce solid tumor volume. The extract of *I. linnaei* is rich in flavonoids and saponins. Flavonoids have been found to possess antimutagenic and antimalignant effect.

**39. LEEA INDICA:**

The crude methanolic extract of the leaves of *Leea indica* (*Vitaceae*) was examined for their anti tumor activity against Ehrlich Ascites Carcinoma (EAC) cells in Swiss albino mice. The compound at the dose of 40mg/kg/day significantly decreases tumor weight. Glycosides, mollic acid arabinoside (MAA) and mollic acid xyloside the growth of Ca<sup>2+</sup> Ski cervical cancer cells with IC<sub>50</sub> of 19.21 μM. Remarkable cytotoxic activity shown by *L. indica* extracts can be attributed mainly to phenol, flavonoids and gallic acid. Gallic acid is isolated from the leaves of *L.indica* and active against several cancer cell lines.

**40. LIRIDENDRON TULIPIFERA:**

The methanolic extract of the stem, leaves and roots bark of *Liriodendron tulipifera* (*Magnoliaceae*) showed significant activity against the KB cell culture and inhibitory activity towards farnesyl protein transferase (FPTase). 977). Three active constituents, costunolide, tulipinolide, liriodenine and a new germacranolide, were isolated which are having antiproliferative activity.

**41. LITCHI CHINENSIS:**

The ethyl acetate extract of *Litchi Chinensis* (*Sapindaceae*) fruit pericarp (LFP) displays inhibitory effects on human breast cancer. The major flavonols in the LEP are reported to be procyanidin B4, procyanidin B2 and epicatechin, while cyanindin-3-rutinside, cyaniding-3-glycoside, quercetin-3-rutinosde and quercetin-3-glucoside are identified as the important anthocyanins.

**42. MORINGA OLERFERA:**

The chemopreventive efficacy of the hydroalcoholic extract of *Moringa oleifera* (*Moringaceae*) was evaluated in a two stage model of 7, 12-dimethyl benz (a)-anthracene induced skin papillomagenesis. Topical application of the extract at a dose of 5mg/ kg body weight inhibited the tumor multiplicity. The ethanol extract of the seeds of *M. Oleifera* were examined and the new O-ethyl-4-(α-1-rhamnosyloxy) nebzyl carbamate together with seven known compounds, 4 (α-1-rthamnosyloxy)-benzyl isothiocyanate, niaximicin, niazirin, β-sitosterol, glycerol-1-(9-octadecanoate), 3-O-(6'-O-oleoyl-β-d-glucopyranosyl)-β-sitosterol and β-sitosterol-3-O-β-d-glucopyranoside were isolated.

**43. MAGNOLIA SIEBOLII:**

An aqueous extract of *M. Officinalis* (*Magnoliaceae*) inhibited cell viability and DNA sunthesis in cultured human urinary bladder cancer 5638 cells. Inhibition of proliferation

was the result of apoptotic induction, because FACS analyses of 5638 cells treated with *M. Officinalis* showed a sub-G1 phase accumulation. The extract also increased cytoplasmic DNA-histone complex dose-dependently.

44. **MATRICARIA CHAMOMILLA:**

The anticancer properties of aqueous and methanolic extracts of chamomile (*Marticanria chamomilla*, *Asteraceae*) flower against various human cancer cell lines causes minimal growth inhibitory responses in normal cells, whereas a significant decrease in cell viability was observed in various human cancer cell lines. Chamomile extract confirmed apigenin 7-O-glucoside as the major constituent of chamomile.

45. **MALLOTUS PHILIPPINENSIS:**

95% ethanolic, 50% ethanolic and aqueous extract of glands/hairs obtained from the fruits of *Mallotus philippinensis* (*Euphorbiaceae*) at the concentration of 100 µg/ml were studied for *in vitro* cytotoxic activity against 14 human cancer cell lines from nine different origins. Results revealed that the 95% ethanolic extract showed highest *in vitro* cytotoxic effect against all the 14 human cancer cell lines.

46. **NELUMBO NUCIFERA:**

(S)-armepavine (C19H23O3N; MW313) from ethanolic leaves extract of *Nelumno nucifera* (*Nelumbonaceae*) inhibits the proliferation of human PBMCs activated with PHA. The main polyphenols identified in the above extract were gallic acid, rutin, and quercetin. Cell-cycle arrest of MCF-7 cells treated with NLE at the G0/G1 phase. Its flower also has antitumor activity.

47. **OPERCULINA TURPETHUM:**

A significant increase in lipid peroxidation levels were observed in tested samples of cancer induced rats while the activities of enzymatic antioxidants such as superoxide dismutase, catalase, glutathione peroxidase and nonenzymatic antioxidants like glutathione, ascorbic acid and alpha tocopherol were decreased in cancer bearing animals when compared to controlled animals. Significant increase in breast tumor weight was observed in DMBA group while breast tumor weight decreased significantly in combination of DMBA and *O. Turpethum* extract group.

48. **OLDENLANDIA DIFFUSA:**

Leaf methanol and hexane extract of *Oldenlandia diffusa* (*Rubiaceae*) has been identified for antitumor properties through *Agrobacterium tumefaciens* infection.

Significant tumor inhibition was observed at 100ppm and 1000ppm of the extract. Maximum tumor inhibition of 40.98, 41.93 and 41.89% were observed at 1000 ppm for the accessions of the *Agrobacterium tumefaciens* AtTs0112, AtAc0114 and AtS10105 respectively.

**49. OPHIORRHIZA MUNGOS:**

The anticarcinogenic potential of the phytochemical Luteolin-7-O-glucoside (LUT7G) and camptothecin, isolated from the methanolic extract of leaves and roots of *Ophiorrhiza mungos* (Rubiaceae) was studied against 4 different cancer cell lines.

**50. OCIMUM SANCTUM:**

An aqueous and ethanolic extract of *Ocimum sanctum* (Lamiaceae) leaves has been investigated against human fibrosarcoma cells (HFS cells) and a significant reduction in tumor volume of mice bearing Sarcoma -180 solid tumors. Ethanol extracts of *Ocimum sanctum* (EEOS) has antitumor mechanism in A549 cells and the Lewis lung carcinoma (LLC) animal model.

**51. PHASEOLUS VULGARIS:**

Twenty-one-day-old female Sprague Dawley rats were given an intraperitoneal injection of 1-methyl-1-nitrosourea and 7 d after carcinogen injection were randomized to 1 of 5 groups fed a modification of the AIN-93G diet formulation containing 0, 6.5, 15, 30, or 60% (wt:wt) small red dry bean (*Phaseolus vulgaris*, L.) (*Fabaceae*) incorporated as cooked, freeze-dried, and milled powder. All experimental diets had the same macronutrient content based on proximate analysis.

**52. PERISTROPHE BICALYCOLATA:**

The oil of *P. Bicalycolata* (*Acnathaceae*) displayed *in vitro* cytotoxicity (2.5-22.3 µg/ml) to MCF-7 (human breast tumor) and MDA-MB-468 (human breast tumor) cells. With respect to the oil of *P. Bicalycolata*, beta-caryophyllene (33.9%), alpha-zingiberene (10.4%), germacene D and globulol (5.0%) were the compounds occurring in abundance.

**53. PHYLLANTHUS RHEEDII:**

The alcohol (95%) extract of the whole plant of *Phyllanthus rheedii* (*Euphorbiaceae*) has antitumoral activity on lung carcinoma cell lines (A549), colon carcinoma cell lines (HCT-116), liver carcinoma cell lines (HEPG-2) and cervical carcinoma cell lines (HELA). The HELA cells were treated with 5 mg/ml and the remarkable inhibition of cells was found. The water extract of *P. Urinaria* did not exert any cytotoxic effect on normal cells

such as endothelial cells and liver cells. The anti-cancer activity of *P. Urinaria* extract was due to the apoptosis induced in Lewis lung carcinoma cells (LLC), which was demonstrated by DNA fragmentation analysis and increased caspase-3 activity.

**54. PARIS POLYPHYLLA:**

Eight compounds were obtained from *Paris polyphalla* (*Trilliaceae*) and identified as Falcarindiol,  $\beta$ -ecdysterone and six known saponins. The dry rhizomes (5kg) of *P. Polyphylla* were refluxed three times with 70% EtOH extract of *P. polyphylla*, And then tested for their antitumor against HepG2 cells, with IC<sub>50</sub> values of 13.5  $\mu$ M and 11.6  $\mu$ M respectively. The antitumor activity steroid saponins of *P. Polyphylla* against lung adenocarcinoma cells was found.

**55. PLATYCODON GRANDIFLORUM:**

Petroleum ether extract from *Platycodon grandiflorum* (*Campanulaceae*) was evaluated using human cancer cell lines (HT-29, HRT-18 and HepG2). *P. Grandiflorum* can reduce the extent of a lung metastasis of B16-F10 melanoma cells by inhibiting the adhesion of tumor cells to the basement membrane possibly and activating NK cells. The inhibitory effects of CKS on MMP-2 and MMP-9 activation, relation of tumor invasion and migration in vitro possibly involve mechanisms related to its ability to suppress PMA-enhanced NF-kB activation through ROS signaling pathway. Aqueous extract from the root of *Platycodon grandiflorum* (AEPG) was studied for the cell growth and apoptosis in human lung carcinoma cell line A549.

**56. PLUMBAGO ZEYLANICA:**

Plumbagin (5-hydroxy-2-methyl-1, 4-naphthoquinone) from *Plumbago zeylannica* (*Plumbaginaceae*) may have potential as an antitumor agent by releasing of mitochondria cytochrome c and  $\gamma$ -independent pathways, as shown by the plumbagin-mediated activation of caspase-3 and -9. Plumbagin exhibited cell proliferation inhibition by inducing cells to undergo G<sub>2</sub> -M arrest and autophagic cell death. Blockade of the cell cycle was associated with increased p21/WAF1 expression and Chk2 activation, and reduced amounts of cyclin B1, cyclin A, Cdc2, and Cdc25C. plumbagin also reduced Cdc2 function by increasing the association of p21/EAF1/Cdc2 complex. The reducing effect plumbagin treatment on viability of human prostate cancer cells (PC-3, LNCaP, and C4-2 ) with apoptosis induction, which was accompanied by ROS generation and depletion of intracellular GSH levels. Plumbagin inhibited in a dependent manner.

**57. PIPER NIGRUM:**

Six alkaloids Piperine, piperidine, piperettine, piperanine, piperlongumine and piperlonuminine extracted from *Piper nigrum* (*Piperaceae*) sample showed antiproliferative activity against several human cancer cell lines. The incubation of tumor cell lines with 5-FU in the presence of piperidine or piperine produced an increase in tumor growth inhibition and significantly lowered in lung-cancer bearing mice and B16F-10 melanoma cells. The alkaloid piperidine was purified by refluxion method to check the antitumor activity which shows 51.38% of inhibition at 5µg/ml concentration.

**58. PERILLA FRUTESCENS:**

The methanolic extracts from different parts of *perilla frutescens* (*Lamiaceae*) were evaluated for their antiproliferative activities by using human non-small cell lung A549 cancer cells. The methanolic extract of stalk exhibited moderate antiproliferative activity. Stalk of *P. Frutescens* might be used as a potential source of natural antioxidants and as anti-tumor agent.

**59. RHIZOPHORA APICULATA:**

*Rhizophora apiculata* (*Rhizophoraceae*) extract of whole plants was found to protect mice from cyclophosphamide (CTX) induced leukopenia. The leukocyte counts in the *R. Apiculata* extract treated animals was significantly increased ( $10425 \pm 163 \text{ mm}^3$ ) whereas the control group shows no significant increase ( $7855 \pm 282 \text{ mm}^3$ ).

**60. RUBIA CORDIFOLI:**

The powdered roots of *Rubia cordifolia* (*Rubiaceae*) was percolated with 80% methanol to obtain the methanol extract. Several secondary plant metabolites and four new naphthohydroquinones and two naphthohydroquinone dimers were isolated from roots of *R. Cordifolia* was investigated for the Cox-2 inhibitory activity which may serve as lead molecules for cancer chemoprevention studies.

**61. RADIX SOPHORAE:**

Leachianone A, isolated from Radix Sophorae (*Luguminosae*), possesses cytotoxic activity against human hepatoma cell line HepG2 *in vitro* via induction of apoptosis involved both extrinsic and intrinsic pathways, with an IC<sub>50</sub> value of 3.4 µg/ml 48h post treatment.

**62. SAUSSUREA LAPPA:**

The dried roots of *Saussurea lappa* (*Asteraceae*) were investigated for the anticancer constituents from the hexane extract of this plant, a new sesquiterpene was isolated along with the known compounds costunolide,  $\beta$ -cyclocostunolide, dihydro costunolide and dehydro costuslactone having anticancer active isolates and its derivatives were studied. Three components cynaropicrim reynosin, and santamarien, were isolated from the methanol extract of *S. lappa* radix stem bark showed potent inhibitory effect on the production of tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), a proinflammatory cytokine, in murine macrophage-like cell (RAW246.7 cells).

**63. TYLOPHORA INDICA:**

Tylophorine was extracted from the leaves of regenerated plants using organic solvents such as like hexane, chloroform and dichloromethane and separated on high performance thin layer chromatography (HPTLC) using toluene: chloroform: ethanol: ammonia (4:3.5:1.5:drop) as mobile phase. Amount of tylophorine obtained was 80 and 71  $\mu\text{g/ml}$  from callus raised and directly cultured *in vitro* plants respectively.

**64. TARAXACUM OFFICINLE:**

The ethylacetate, butanol, methanol and cold water extracts of the plant had significantly higher radical scavenging (%) and total phenolic contents than the hot water, and hexane extracts. Cytotoxic effects of all the extracts evaluated using the SRB assay on HeLa, HT-29, HepG2 and KB cell lines demonstrated that the ethylacetate extract with an  $\text{IC}_{50}$  of 15.5  $\mu\text{g/ml}$  was potentially very toxic against human mouth epidermal carcinoma (KB) than all other extracts.

**65. TAXUS YUNNANENSIS:**

The extract of roots of *T. Yunnanensis* (*Taxaceae*) showed that they were rich in taxane diterpenoids, including paclitaxel and dihalocephalomannine with anticancer Property. Two new taxane diterpenes, dantaxusin C and dantaxusin D were isolated from an ethanol extract of the aerial parts of *Taxus yunnanensis* along with 14 known taxoids.

**66. TABEBUIA AVELLANEDAE:**

Callus and cell suspension cultures of *Tabebuia acellanadae* (*Bignoniaceae*) produced promising antitumour-promoting furananaphthoquinones, 5-hydroxy-2-(1-hydroxyethyl)naphtha [2,3-b] furan-4,9-dione in high yields. A series of naphthoquinones based on the naphtha [2,3-b] furan-4,9-dione skeleton such as (-)-5-hydroxy-2-(1'-hydroxyethyl)naphtha

[2,3-b]furan-4,9-dione) and its positional isomer, (-)-8-hydroxy-2-(1'-hydroxyethyl) naphtha [2,3-b] furan-4,9-dione, which are secondary metabolites found in the inner bark of *Tabebuia avellanadae*, were synthesized and their biological activities such as antiproliferative and cancer chemopreventive activity were examined.

#### 67. **VCCINIUM MACROCARPON:**

Triterpene cinnamates 3-O-p-hydroxycinnamoyl ursolic acid purified from the ethyl acetate extracts of *Vaccinium macrosporan* (Ericaceae) showed slightly greater activity of compound in most tumor cell lines, with GI<sub>50</sub> values of approximately 20 µM in MCF-7 breast, ME180 cervical and PC3 prostate tumor cell lines. Antiproliferative activities of isolated compounds from cranberries extracts against HepG2 human liver cancer and MCF-7 human breast cancer cells were evaluated.

### **MEDICINAL PLANTS OF INDIA:**

#### 1) **PLANTAGO OVATA:**

This belongs to family Plantaginaceae is an annual herb grown during the rainy season. The swelling property of the husk after absorption of water is the cause of its use as famous medicine against constipation and gastrointestinal irritations. It is also used for the treatment of number of other stomach related disorders. Various polysaccharides and amino acids are reported from its seeds. In addition, it is used in food industries for the preparation of ice creams, candy etc. the raw drug is collected solely from cultivation.

#### 2) **CASSIA ANGUSTIFOLIA:**

The herb is a member of family, Fabaceae. It contains sennosides, which is responsible for the purgative action. The drug is used as a cathartic in giving relief to constipation.

#### 3) **ALOE BARBADENSIS:**

Aloe-a member of the family Liliaceae. The plant is perennial herb with fleshy leaves and condensed stem. Flowering occurs in winter and the inflorescence stalk is about 90-150 cm long with orange coloured flowers. Leaves contain gel (polysaccharides) and leaf exudates contain aloins, which are commercially useful. Gel has a cooling and moisturizing action and hence used in cosmetic industries and the leaf exudates contains aloins and aloe emodine, which are used as pain killer and purgative.

**4) WITHANIA SOMNIFERA:**

*Withania somnifera*, a member of family Solanaceae and is distributed throughout India. Root is the major medicinally important part in addition to leaves and seeds. Alkaloids (withanoloids and withanins) present in the roots are believed to be effective in treatments of stress induced disorders, fatigue, dropsy, male impotency, neurosis, etc. It is commonly used as a general tonic.

**5) CHLOROPHYTUM BORIVILIANUM:**

The plant, a member of family Liliaceae is a perennial herb with condensed stem disc and a whirl of sessile leaves. Fasciculated roots contain saponins and are medicinally important. It is used as a general tonic and is a well-known aphrodisiac. Raw drug is collected both from wild as well as from cultivation. The plant is propagated by the stem disc with the attached fleshy roots as well as by seeds. Unorganized collection of the species from the natural habitat has caused endangered species status.

**6) COMMIPHORA WIGHTII:**

The important medicinal plant belongs to family Burseraceae. The oleo-gum resin of guggal having chief constituents like myrcene and dimyrcene is known to be highly effective in the treatment of obesity, arthritis and several other diseases in Indian system of medicine. The species is included in the Red data book (IUCN) as over exploited species in the country.

**7) TINOSPORA CORDIFOLIA:**

It is a deciduous perennial climber belongs to the family Menispermaceae. The stem and leaves are medicinally used as raw drug. *Tinospora* stem is a common constituent of a number of ayurvedic vital tonics for the treatment of general debility, dyspepsia, fevers and urinary diseases. Starch called as "Sat Giloe" present in the stem along with alkaloids is the active principle of the species. Major constituents are tinosporon, tinosporic acid, tinosporoside, tinosporin and cordifolide. Leaf also contains a number of alkaloids. Leaf is used for the treatment of gout, jaundice and rheumatism. Raw drug is mainly obtained from the wild habitats of species.

8) ***ASPARAGUS RACEMOSUS:***

Satavary is a perennial spiny woody climber belongs to the family Liliaceae. Fasciculated roots are medicinally important. It contains saponins and is used for the treatment of dysentery, tumours, rheumatism and kidney and liver disorders. Powdered roots are a common ingredient of a number of vital tonics, which are believed to cure sexual weaknesses, leucorrhoea and increase lactation in feeding mothers.

9) ***BACOPA MONNIERI:***

Member of Scrophylariaceae family is a creeping, branched succulent perennial herb distributed in wet and marshy lands through out India. The whole herb is the source of the source of the ayurvedic drug 'Brahmi'. It is used in improving memory and intelligence and also in the treatment of dermatosis, anaemia, deabetics and insanity. Bacoposide is considered as teh major active ingredient in this plant. Raw drug is mainly collected from the wild.

10) ***CENTELLA ASIATICA:***

A member of family Apiaceae is a prostrate slightly aromatic, perennial herb commoly found as a weed creeping on ground in crop fields. It is used for the treatment of leprosy, skin diseases and to improve memory. It is also against cholera, ulcers, bronchitis, leucorrhoea and kidney troubles. Asiaticosie, indocentelloside, thankuniside are the major glycosides responsible for the medicinal properties.

11) ***ANDROGRAPHIS PANICULATA:***

Belongs to family Acanthaceae is a brached annual undershrun of about 30-100cm tall. The whole herb is medicinally useful. Andrographolide is the active principle having the therapeutic considered as a blood purifier and used for the treatment of skin deseases.

12) ***SWERTIA CHIRAYITA:***

It is an erect annual herb belongs to family Gentianaceae. The bitter tonic made from the raw drug inproves bile secretion and used for the treatment of bronchial asthma, liver disorders, and anaemia. The juice of fresh plant or infusion of dry plants is prescribed as a "blood purifier" in skin diseases. The active ingredient of the raw drug includes ophelic acid, glucosides, etc.

13) ***MUCUNA PRURIENS:***

The species is a pubescent annual liana belongs to Fabaceae family. The active principle is L-Dopa. The seeds are used to treat, Parkinson disease, sexual disorders, cholera, urinary troubles and liver and gall bladder diseases. L-dopa present in the seeds is the active principle responsible for therapeutic action.

14) ***GENTIANA KURROO:***

The plant is a small perennial herb belongs to family Gentianaceae. Gentiopicrotin and tannins are present in the raw drug. Rhizome is used as a gastric stimulant and improves appetite. The raw drug is mainly collected from the wild. It is also used for the treatment of fever, abdominal pains, and for the purification.

15) ***VALERIANA WALLICHII:***

Indian valerian belongs to family Valerianaceae is a perennial herb of about 45 cm height and rootstock is thick, nodular and aromatic. Roots of the species are useful in diseases related to eye, blood, liver and spleen. Leaves are used for the treatment of headache. Valeportialtes are obtained from roots and rhizomes. Roots are also used in aromatic industry. Raw drug is collected mainly from the wild since cultivation is not yet popularized.

16) ***PODOPHYLLUM HEXANDRM:***

The species belongs to family Berberidaceae is an erect, succulent herb with creeping perennial rhizome bearing numerous roots. The rhizome and the root constitute the raw drug source of resin, podophyllin or podophyllin resin podophyllotoxin is the active principle of podophyllin. The raw drug is used for the treatment of ulcers, cuts and wounds. It is a purgative and is used in curing skin diseases and tumor growth. Podophyllin has acquired special attention during recent time due to its action against cancers. The raw drug is collected mainly from the wild since the cultivation is not gained momentum.

17) ***GYMNEMA SYLVESTIS:***

It is a woody climber of family Asclepiadaceae. The leaves when chewed temporarily cease one's ability to sense sweet taste. The species gained importance, since it is used to cure diabetics. Leaves are used for the treatment of diabetics. Gymnemic acid present in the leaves is believed to reduce blood glucose level.

Leaves and roots are also used to treat headache, polyuria, leprosy, wounds and pruritis. The raw drug is mainly collected from the wild.

18) ***ACONITUM HETERPHYLLUM:***

It is a biennial herb with fleshy roots belongs to family Ranunculaceae. Roots are used medicinal purposes. It is used in diarrhoea. Dysentery and gastric pain. It is used as a bitter tonic to combat debility after malaria and other fevers. It is also used against hysteria, dyspepsia, vomiting and cough. Wild habitats are the sole source of the raw drug since the plant is not in cultivation.

19) ***TERMINALIA BELLERICA:***

Is a deciduous tree of the family Combretaceae that grows up to 40m heights. The fruit of the species is commercially known as myrobalan or belliric myrobalan and is medicinally important. It is anticanerous, and used against heart against heart diseases, anaemia and rheumatism. Beta sitosterol, gallic acid and bellericanin are the major active ingredients of the raw drug. Raw drug is mainly collected from the forests.

20) ***TERMINALIS CHEBULA:***

Harde is a tree of about 15-24 m tall belongs to the family Combretaceae. In combination with Emblic myrobalam (*Emblica officianalis*) and belliric myrobalan (*Terminalia bellirica*), under the formulation 'triphala' (thre fruit), *T.chebula* is used extensively in a number of ailments. The main purgative ingredient of thriphala is harde and the purgative principle is in the pericarp of the fruit, which is a glycoside similar to sennoside A of Senna (*Cassia angusrifolia*). Chebulin, chebulinic acid, tannic acid and behinic acid are the major active ingredient of the raw drug. Forests are the main sourcues of raw drug collection.

21) ***EMBLICA OFFICINALIS:***

The plant is a small to medium sized deciduous tree belongs to family Euphorbiaceae. It is one of the richest sources of vitamin C. the fruits, leaves and bak are rich in tannins, root contains ellagic acid and lupeol. The fruits are used for the treatment of vomiting, biliousness, urinary discharges, constipation, leprosy, piles and diseases related to eyes. Triphala, an ayurvedic formulation includes amla as one of the three constituents.

**22) PIPER LONGUM:**

Long pepper is a slender aromatic perennial undershrub belonging to the family Piperaceae. Ripened green fruits and roots are used as the raw drug. India imports a large quantity of raw drug from Malaysia and Singapore. The fruits are used as spice also. It has a pepper like taste. Commercially, the raw drug of long pepper is mixed with material from *P. Retrofractum* or *P. Peepuloides* ., and it is very difficult to distinguish the raw drug from these different species. Piperine and pipartine are the two important alkaloids responsible for the therapeutic action. In addition also the raw drug contains a number of essential oils. The fruits as well as the roots are used for the treatment of diseases related respiratory tracts viz., bronchitis, asthma, cough, etc. it is also used in muscular pains, inflammations, drowsiness, epilepsy, leprosy, dysentery and ailments related gall bladder and bile duct.

**23) SIDA CORDIFOLIA:**

An annular herb and member of Malvaceae family. *Sida cordifolia* is considered as the source of raw drug bala in North India while in South India *Sida rhombifolia* is accepted as the source of the raw drug. The root of the species is used as the raw drug for the treatment rheumatism. It imparts strength to the body and is useful in the treatment of facial paralysis, general debility, sciatica, headache, uterine disorders, etc.

**24) ADHATODA VASICA:**

It is an evergreen, perennial shrub belongs to family Acanthaceae. The species is a source of a well-known raw drug, 'vasaka' which is mainly used for the treatment of bronchitis, asthma, cold, cough and whooping cough. Fresh or dried leaves are used mainly as the raw drug. It is used in the form of fresh juice, decoction, infusion or powder. The leaf juice is also used to cure dysentery, diarrhoea and glandular tumour. A number of alkaloids are present in the raw drug of which vascine and vasicinone are important.

**25) ACORUS CALAMUS:**

The species is a native of Eupore belongs to family Araceae. The dried rhizomes constitute the commercial raw drug of 'Calamus'.  $\alpha$  and  $\beta$  asarone, acoric acid and choline are the major active principle of the raw drug. It is believed to

improve memory power and intellect. It is also useful in the treatment of diarrhoea, dysentery, abdominal obstructions and colic. Anticarcinogenic property of the species is also reported recently.

26) ***SARACA ASOCA***:

It is a medium size, evergreen tree with black belongs to family Fabaceae. The activity of the drug is due to the presence of steroidal component and calcium salt. Bark also contains tannins. It is propagated by seeds and is also cultivated as an ornamental plant. The increased demand of the raw drug in recent years has caused overexploitation of the species in wild habitats. It one of the flagship species targeted for wide scale cultivation in south India.

27) ***RAUVOLFIA SERPENTINA***:

Sarpahandha a member of family Apocynaceae is a perennial under shrub with irregular tubular roots distributed through our India. Roots contain alkaloids (reserpine, deserpidine and reseinamine) which a sedative and used to control high blood pressure. It is also used for the treatment of insomnia, asthma and acute stomachache. Ruthless collection of the species from its wild habitats and the Government of India has prohibited its collection from the wild. The crop is under cultivation and propagated by seeds.

28) ***CATHARANTHUS ROSEUS***:

Periwinkle belongs to the family Apocynaceae is an erect annual or perennial herb with white or pink flowers. All parts of the plant, especially roots contain alkaloids. Leaves are used to treat menorrhagia and the plants are also used to treat diabetics in Ayurveda. However, the alkaloids vincristine and vinblastine present mainly in the roots of the plant in used to treat a various types of cancers including leukaemia in modern medical systems.

29) ***PHYLLANTHUS AMARUS***:

It is a small herb of about 60cm height and belongs to the family Euphorbiaceae. The whole herb is used for the medicinal purpose. It is bitter in taste and is used mainly for the treatment of jaundice. It is also used in dyspepsia, diarrhoea and dysentery. The herbage portion contains the bitter principle phyllanthin which is responsible for the therapeutic action.

**30) *OCIMUM SANCTUM*:**

It is an erect highly branched aromatic perennial herb belongs to the family Lamiaceae. Leaves, flowers and occasionally the whole plant are medicinally used to treat heart diseases, leucoderma, asthma, bronchitis and fever. The leaves and tender parts of the shootsm are economically important and it yields essential oils. The essential oils obtained have immense value in aroma industry. The chemical constituents of the esessential oils are monterpenes, sesquiterpines, sesquiterpines and phenols with their alcohols, esters, aldehydes, etc.

**31) *ATROPA BELLADONNA*:**

A perennial branching herb growing to 5 feet tall, with 8 inch long ovate leaves, belongs to family Solanaceae. The medicinal peoperties of Belladonna depend on the presence of Hyoscyamine and Atropine. Belladonna is a most valuable plant in the treatment of eye disease, Atropine, obtained during extraction, being its most important constituent on account of its power of dilating the pupil.

**32) *CINFHONA OFFICINALIS*:**

Cinchona officinalis is a tree native to Amazon Rainforest vegetation, belongs to family Rubiaceae. This plant is used for the production of quinine, which is an anti-fever agent especially useful in the prevention and treatment of malaria. There are a number of various other chemicals which are made from this tree, and they include cinchonine, cinchonidine and quinidine.

**33) *PAPAVER SOMNIFERUM*:**

The plant is an erect herb characterized by a drooping bud and grey latex, from the family Papaveraceae. In midicine, opium as such, the capsule after the recovery of opium adn teh seeds are used. It is a well-know sedative, having a constipating effect, so used in diarrhoea and pains in the body; the dry capsule may be used where mild action id required. The major alkaloids are morphine, thebaine, noscapine and papacerine.

**34) *DIGITALIS PURPUREA*:**

It belongs to family Scrophulariceae. Due to the presencne of the cardiac glycoside digitoxin, the leaves, flowers and seeds of this plant are all poisonous to

humans and some animals and can be fatal if eaten. In allopathy, cardiac-glycoside digitoxin isolated from the leaves or seeds is used for various heart troubles.

## MAJOR AROMATIC MEDICINAL PLANTS:

### 1. *ABELMOSCHUS MOSCATUS*:

Muskdana belongs to family Malvaceae is an erect perennial herb with fruits similar to Lady's finger but smaller in size. Seeds are musk scented and are used to flavour food and as a substitute of musk. It is used to treat intestinal problems, stomatitis and heart diseases. In Unani, it is also used to treat dyspepsia, urinary discharges, gonorrhoea, leucoderma and itching.

### 2. *CYMBOPOGON FLEXUOSUS*:

Lemongrass belongs to family Poaceae and three species of *Cymbopogon* i.e., *C. flexuosus*, *C. citrates* and *C. pendulus*, commonly known as lemongrass. Citral extracted from the oil is of commercial importance. The oil is used in flavouring and medicinal industry and also for production of Vitamin A and synthetic violet. India is one of the major suppliers of lemongrass oil in the world market. A large number of high yielding varieties are available for cultivation.

### 3. *CYMBOPOGON MARTINI*:

Palmarosa or Rosha grass is a tall perennial herb belongs to family Poaceae. The oil has high demand in perfumery, soap, cosmetic and blending tobacco products industries. Owing to its oil, hence referred to as East Indian Geranium oil in commerce. The species is under cultivation in Central, western and southern states of India.

### 4. *PELARGONIUM GRACEOLENS*:

The genus *Pelargonium* belonging to the family Geraniaceae. Rose scented essential oil, extracted from its leaves and tender shoots are commercially important. The crop is propagated by stem cuttings and root suckers. Geranium oil contains 60-70% alcohols like, citronellol, geraniol; 20-30% esters like, geranyl tosylate, geranyl acetate, citronellyl acetate and the rest aldehyde and ketones, etc.

### 5. *POGOSTEMIN PATCHOULI*:

The plant belongs to family Lamiaceae is a perennial erect or ascending herb. Oil distilled from shade dried leaves is used in perfumery to give a solid foundation and lasting character of other fragrances since it has a fixative property to prevent the rapid evaporation of perfumes. The patchouli oil is generally blended with other essential oils like geranium

oil or clove oil before use. The oil possesses antibacterial activity and is used as an ingredient in insect-repellent preparations. The oil is also used in major food products as a flavouring agent.

6. ***MENTHA ARVENSIS***:

Mentha or mints includes a number of aromatic perennial herbs belonging to family Lamiaceae. Peppermint oil is carminative, antiseptic and gastro-stimulant properties and used in confectionaries, alcoholic drinks, dental creams and mouthwashes. Spearmint oil is rich in carvone and has digestive and gastrostimulant properties. It is used in confectionary, chewing gum and tooth pastes. Nergamot mint oil is rich in linalyl acetate and linalool and mainly used in cosmetic industry. The herbage of the plants are economically useful which yields essential oils. The chief chemical constituents of the essential oils are menthol, carvone, linalyl acetate and linalool and they are highly demanded in pharmaceutical, food flavour, cosmetics, beverages and related industries.

7. ***ROSA DAMASCENA*** :

Rose belongs to family Rosaceae. Half opened flowers have more fragrance and oil yield than the fully opened flowers. In India about 1500 to 2000 hectares of land is under rose cultivation, which is mainly located in Uttar Pradesh for extraction of rose oil, rose water, absolute and rose concrete. Different perfumery products obtained from rose are rose attar, Gulkhand, Gul-roghan, Punkhuri and Otto rose.

8. ***JASMINUM SAMBAC***:

The species belongs to family Oleaceae. The essential oils obtained from the flowers are used in perfumery and are export oriented also.

9. ***POLIANTHUS TUBEROSA***:

It is a short – stemmed plant with underground bulbs belongs to family Amaryllidaceae. The fresh flowers yield about 0.08 to 0.11 percent oil. There are different varieties in tuberose based on the number of petals or petal type viz. Calcutta single, single Mexican, semi-double, Variegated, Pearl double, Rafat Rekha and Swarna Rekha. The plant is propagated mainly by bulbs.

10. ***VERICERIA ZIZANIODES***:

It is a tall perennial grass of about 1.5 to 2.0 m height belongs to family Poaceae. The aromatic roots are traditionally used for making mats, fans, door-screens which produce

fragrance imparting a cooling effect during extreme summer when sprinkled with water. The roots on distillation produce volatile oils. The oil finds extensive use in perfumery as a fixative and as an odour contributor in bases, and also as a raw material for the isolation of vetiverol and vetiverone. It is also medicinal and used against flatulence and colic pains.

#### 11. *CYMBOPOGON WINTERIANUS*:

It is a perennial herb from the family Poaceae; Citronella oil is obtained by steam distillation of partially wilted leaves. The oil whose ingredients are citronellal, citronellol and geraniol are employed in the manufacture of soap, pharmaceuticals, and perfumery, cosmetic and flavouring agents.

#### 12. *LAVANDULA ANGUSTIFOLIA*:

It is a perennial aromatic herb from the family Lamiaceae. Major components of the essential oil are linalool and linalyl acetate. The essential oil is antiseptic and antispasmodic.

**TABLE 1. SOME COMMON MEDICINAL PLANTS AND PHARMACOLOGICAL USES**

S.NO	BOTANICAL NAME	FAMILY NAME	PHARMACOLOGICAL USES
1.	<i>Acacia asak</i>	<i>Fabaceae</i>	Skin diseases and tooth ache
2.	<i>Acacia ehrenbergiana</i>	<i>Fabaceae</i>	Tooth ache and ear infection
3.	<i>Acacia etbaica</i>	<i>Fabaceae</i>	Constipation, and ringworm infection
4.	<i>Acacia laeta</i>	<i>Fabaceae</i>	Diabetes and antibiotics
5.	<i>Acacia lahai</i>	<i>Fabaceae</i>	Taeniocides and rheumatism
6.	<i>Acacia mellifera</i>	<i>Fabaceae</i>	Evil eyes and diabetes
7.	<i>Acacia nilotica</i>	<i>Fabaceae</i>	Abdominal pain and antiemetic
8.	<i>Acacia oerfota</i>	<i>Fabaceae</i>	Eye disease and evil eyes
9.	<i>Acacia Oregana</i>	<i>Fabaceae</i>	Easing labor and back pain
10.	<i>Acacia polyacantha</i>	<i>Fabaceae</i>	Rheumatism and sciatica
11.	<i>Acacia Senegal</i>	<i>Fabaceae</i>	Diabetes and Abdominal pain

12.	<i>Acacia Seyal</i>	<i>Fabaceae</i>	Diabetes and hypertension
13.	<i>Acacia Sieberiana</i>	<i>Fabaceae</i>	Abdominal helmentes and scabies
14.	<i>Acacia Tortilis</i>	<i>Fabaceae</i>	Snake and Scorpion bite and gonfi
15.	<i>Achyranthes aspara</i>	<i>Amaranthaceae</i>	Tonsillitis and Nails inflammation
16.	<i>Acokanthera schimperi</i>	<i>Apocynaceae</i>	Skin wound , eye infection
17.	<i>Adansonia digipata</i>	<i>Bombacaceae</i>	Anti abortion and eye infection
18.	<i>Aerva lanata</i>	<i>Amaranthaceae</i>	Gerefta , Febrile
19.	<i>Agave sisalana</i>	<i>Agavaceae</i>	Diarrhea and ear infection
20.	<i>Ajuga integrifolia</i>	<i>lamiaceae</i>	Malaria hebatis, dysenter and swelling
21.	<i>Albizia amara</i>	<i>Fabaceae</i>	Blood pressure and anti-lice
22.	<i>Albizia anthelmintice</i>	<i>Fabaceae</i>	Abdominal helminthes,Diarrhea and burns
23.	<i>Allium cepa</i>	<i>Alliaceae</i>	Tp hepatitis
24.	<i>Allium sativum</i>	<i>Alliaceae</i>	Hyper tension, malaria and asthma
25.	<i>Aloe camperi</i>	<i>Aloaceae</i>	Malaria , hepatomegaly and splenomegaly
26.	<i>Aloe elegance</i>	<i>Aloaceae</i>	Tiabetics and abdominal pain
27.	<i>Aloe macrocarpa</i>	<i>Aloaceae</i>	Impotency malaria and easing labor

28.	<i>Aloe percarsa</i>	<i>Aloaceae</i>	Anthrax, malaria
29.	<i>Amara</i>	<i>Amaranthaceae</i>	Back pain, Tooth pain and Anti-helminthes
30.	<i>Annona muricata</i>	<i>Annonaceae</i>	Kill lice, Bedbugs and Gastritis
31.	<i>Anogeissus leiocarpus</i>	<i>Combretaceae</i>	Anthrax, Hepatitis
32.	<i>Apium graceolnese</i>	<i>Apiaceae</i>	Theumatism, Asthma and Bladder infection
33.	<i>Argemone Mexicana</i>	<i>Papaveraceae</i>	Antibiotics and Cataracts
34.	<i>Artemisia annua</i>	<i>Astereaceae</i>	Bladder infection, Hepatitis and Malaria
35.	<i>Asparagus africanus</i>	<i>Asparagaceae</i>	Skin lesion and Heart diseases
36.	<i>Aspilia mosambicensis</i>	<i>Asteraceae</i>	Kidney diseases and Bleeding after delivery
37.	<i>Astragalus atropilosulus</i>	<i>Fabaceae</i>	Hemorrhoids, latching and diarrhea
38.	<i>Anicenna marina</i>	<i>Verbeaceae</i>	Ulcers, Diabetics, Asthma and Cancer
39.	<i>Azadirachta indica</i>	<i>Meliaceae</i>	Haemorrhoids, Fungal and insects
40.	<i>Balanites aegyptiaca</i>	<i>Balanitaceae</i>	Head ache, abdominal pain and Bhlharzias
41.	<i>Barbeya oleoides</i>	<i>Barbeyaceae</i>	Wound infection
42.	<i>Barleria eranthemoides</i>	<i>Acanthaceae</i>	Eye problem, Tonsillitis and Eczema
43.	<i>Becium grandiflorum</i>	<i>Lamiaceae</i>	Cramps, Anti-inflammatory and Malaria
44.	<i>Bersama abyssinica</i>	<i>Melanthaceae</i>	Heart disease and tape-worm
45.	<i>Bidens pilosa</i>	<i>Asteraceae</i>	Gerefta and gonfii
46.	<i>Boscia angustifoila</i>	<i>Capparidaceae</i>	Snake-bite, Gastritis and Gonfii
47.	<i>Boscia salicifolia</i>	<i>Capparidaceae</i>	Scabies, Bloating and Choilynichites
48.	<i>Noscia senegalensis</i>	<i>Capparidaceae</i>	Snake and Scorpion venom
49.	<i>Boswellia papyrifera</i>	<i>Burserceae</i>	Fever, Tranquilizer and Evil Spirit
50.	<i>Brassica olearacea</i>	<i>Brassicaceae</i>	Cough and Fungus
51.	<i>Brassica</i>	<i>Brassicaceae</i>	Gastritis, Rhenmatism and

	<i>olearacea</i>		Asthma
52.	<i>Brucea antidysesteica</i>	<i>Simaroubaceae</i>	Diarrhea, Evil eyes and Rabies
53.	<i>Buddleia polystachya</i>	<i>Loganiaceae</i>	Evil eye, Segri and insects
54.	<i>Cadaba farinose</i>	<i>Capparidaceae</i>	Ophthalmia, insect repellent and body pains
55.	<i>Cadalia purpurea</i>	<i>Fabacase</i>	Wound infection and nail inflammation
56.	<i>Colotropis procera</i>	<i>Asciepiadaceae</i>	Homorrhoids, Wound and leprosy
57.	<i>Calpurnia aurea</i>	<i>Fabaceae</i>	Snake and Scorpion venom and leprosy
58.	<i>Capparis decidua</i>	<i>Capparidaceae</i>	Chest pains, Jaundice and malaria
59.	<i>Capparis tomentosa</i>	<i>Capparidaceae</i>	Cold, Wound infection and purgative
60.	<i>Carica papaya</i>	<i>Caricaceae</i>	Diabetes, Amoeba and Typhoid fever
61.	<i>Caralluma speciosa</i>	<i>Asclepediaceae</i>	Burns and Wound infection
62.	<i>Carissa spinarum</i>	<i>Apocynaceae</i>	Malaria, Splenomegaly and Hepatomegaly
63.	<i>Casimiroa edulis</i>	<i>Rutaceae</i>	Menstrual disorder and Constipation
64.	<i>Capcicum annum</i>	<i>Solanaceae</i>	Arm-stream, Alopecia and Hepatomegaly
65.	<i>Celtis africana</i>	<i>Ulmaceae</i>	Rheumatism and Lung disease
66.	<i>Chenopodium album</i>	<i>Chenopodiaceae</i>	Choilynichitis, Burns and Wounds
67.	<i>Cicer arietinum</i>	<i>Fabaceae</i>	Malaria and Diarrhea
68.	<i>Cicorium endive</i>	<i>Asteraceae</i>	Diabetes and wound infection
69.	<i>Cinnamomum</i>	<i>Lauraceae</i>	Rheumatism, Diabetes and Blood circulation
70.	<i>Cissus quadrangularis</i>	<i>Vitaceae</i>	Anti-fungal and insects
71.	<i>Ciddus rotundifolia</i>	<i>Vitaceae</i>	Antibiotic, Gonfi and Antifungal
72.	<i>Citrus limon</i>	<i>Rutaceae</i>	Gastric and Digestion problem
73.	<i>Citrus sinensis</i>	<i>Turaceae</i>	Cough and Antibiotic
74.	<i>Citrus reticulate</i>	<i>Rutaceae</i>	Diarrhea, Helminthic infection
75.	<i>Clematis simensis</i>	<i>Ranunculaceae</i>	Leishmaniasis and Eczema

76.	<i>Cherodendrum myricoides</i>	<i>Verbenacea</i>	Dysentery and Abdominal helmets
77.	<i>Clutia lanceolata</i>	<i>Euphorbiaceae</i>	Gastritis, Diabets and Rheumatism
78.	<i>Coffea Arabica</i>	<i>Rubiaceae</i>	Eye diseases, Snake poison and Burns
79.	<i>Combretum aculeatum</i>	<i>Combretaceae</i>	Conjunctivitis and Ear infection
80.	<i>Combretum fragrans</i>	<i>Combretaceae</i>	Lung diseases and constipation
81.	<i>Combretum molle</i>	<i>Combretaceae</i>	Jaundice and Epidermal wounds
82.	<i>Conetes abyssinica</i>	<i>Carryphyllaceae</i>	Sun strike, insects and Viral infection
83.	<i>Commelina Africana</i>	<i>Commelinaceae</i>	Skin diseases and Hypertension
84.	<i>Commicarpus pedunculatus</i>	<i>Nyctaginaceae</i>	Scorpion and Snake bite
85.	<i>Commiphora Africana</i>	<i>Burseraceae</i>	Expel the placenta after birth and gastric
86.	<i>Cannabis sativa</i>	<i>Cannabidaceae</i>	Bronical asthma and Segri
87.	<i>Cordial Africana</i>	<i>Boraginaceae</i>	Abdominal pain and Wound infection
88.	<i>Cordial monoica</i>	<i>Moraginaceae</i>	Wound infection and Sun strikes
89.	<i>Coriandrum sativum</i>	<i>Apiaceae</i>	Diuritic and Fever, Hypertension
90.	<i>Croton macrostychus</i>	<i>Euphorbiaceae</i>	Head ache, Eye diseases, Ascariasis
91.	<i>Cucumis dipsaceus</i>	<i>Cucurbitaceae</i>	Abdominal helminthes and Diarrhea
92.	<i>Cucurbita pepo</i>	<i>Cucurbitaceae</i>	Intestinal warms, Diuretics and Lactation
93.	<i>Cymbopogon citrates</i>	<i>Poaceae</i>	Fever of domestic animals and Ascariasis
94.	<i>Cynodon dactylon</i>	<i>Poaceae</i>	Snake poison and Skin crack
95.	<i>Cynoglossum lanceolatum</i>	<i>Boraginaceae</i>	Abdominal pain and Ear infection
96.	<i>Cyphostemma adenocoule</i>	<i>Vitaceae</i>	Snake poison and Amoeba
97.	<i>Dalbergia melanoxyton</i>	<i>Fabaceae</i>	Snake poison and insect repellants
98.	<i>Datura carota</i>	<i>Solanaceae</i>	Rabies, Dandruff and Tooth ache
99.	<i>Daucus carota</i>	<i>Solanaceae</i>	Diabets, Gastritis and Eye clearness
100.	<i>Delonix elata</i>	<i>Fabaceae</i>	Laxative and purgative
101.	<i>Dichrostachys cinerea</i>	<i>Fabaceae</i>	Impotency
102.	<i>Diospyros abyssinica</i>	<i>Ebenaceae</i>	Fungus and Termite resistant

103.	<i>Diospyros mespiliformis</i>	<i>Ebenaceae</i>	Fungus and Termite resistant
104.	<i>Dobera glabra</i>	<i>Salcadoraceae</i>	Intestinal parasites and Rheumatism
105.	<i>Dodonaea angustifolia</i>	<i>Sapindaceae</i>	Abdominal Helminthes and Dandruff
106.	<i>Dombeya torrid</i>	<i>Sterfuliaceae</i>	Asthma, Cough and Skin wound
107.	<i>Dracaena schizanta</i>	<i>Lilaceae</i>	Ear ache and Chronic ulcer
108.	<i>Embelia schimperi</i>	<i>Myrsinaceae</i>	Taeniaceae and Insecticides
109.	<i>Eragrosis teff</i>	<i>Poaceae</i>	Anemia and Constipation
110.	<i>Erythrina abyssinica</i>	<i>Fabaceae</i>	Tooth pain, Fever and Gastrointestinal
111.	<i>Eucalyptus camaldulensis</i>	<i>Myrtaceae</i>	Cough, Antibiotic and insecticide
112.	<i>Eucalyptus globules</i>	<i>Myrtaceae</i>	Diabetes, Cough, Rheumatism and Gonfii
113.	<i>Euclea racemosa</i>	<i>Ebenaceae</i>	Malaria, Hepatitis and purgative
114.	<i>Eugenia carophyllata</i>	<i>Myrtaceae</i>	Emetic, Tooth Ache, TB, and Menstruation
115.	<i>Euphorbia abyssinica</i>	<i>Euphorbiaceae</i>	Abdominal pain, hepatitis and antibiotic
116.	<i>Eupharbia candelabrum</i>	<i>Euphorbiaceae</i>	Wound haling and Malaria
117.	<i>Euphorbia nubica</i>	<i>Euphorbiaceae</i>	Kidney problem and insecticides
118.	<i>Euphorbia plicanatha</i>	<i>Euphorbiaceae</i>	Asthama, leshmaniasis
119.	<i>Euphorbia prostrate</i>	<i>Euphorbiaceae</i>	Abdominal pain, Hepatitis and Antibiotic
120.	<i>Euphorbia tirucalli</i>	<i>Euphorbiaceae</i>	Abdominal pain, skin diseases and warts
121.	<i>Ficus carica</i>	<i>Moraceae</i>	Eczema, alopecia, insecticides and malaria
122.	<i>Ficus glumosa</i>	<i>Moraceae</i>	Itching and Rabies
123.	<i>Ficus sucomorus</i>	<i>Moraceae</i>	Internal swell of abdomen
124.	<i>Ficus thonigii</i>	<i>Moraceae</i>	Insecticides and Cough
125.	<i>Ficus vasta</i>	<i>Moraceae</i>	Cough, Abdominal pain and Obesity
126.	<i>Flaceria trinervia</i>	<i>Asteraceae</i>	Malaria, Pneumonia and Diarrhea
127.	<i>Foeniculum vulgare</i>	<i>Apiaceae</i>	Hemorrhage and Endoparasites
128.	<i>Flueggia virosa</i>	<i>Euphorbiaceae</i>	Intestinal worms
129.	<i>Grewia ferrugenia</i>	<i>Tiliaceae</i>	Burns nad Evil eyes
130.	<i>Grewia flavescens</i>	<i>Tiliaceae</i>	Delivery of the placenta

131.	<i>Grewia mollis</i>	<i>Tiliaceae</i>	Burns and sun strike
132.	<i>Grewia tenax</i>	<i>Tiliaceae</i>	Throat sore, Bone fracture and Myalgia
133.	<i>Grewia villosa</i>	<i>Tiliaceae</i>	Relieve headache and breathlessness.
134.	<i>Gulzotia abyssinica</i>	<i>Asteraceae</i>	Tonsillitis, Dandruff and Ring worm
135.	<i>Heteromorpha arborescens</i>	<i>Apiaceae</i>	Asthama and Tooth ache
136.	<i>Heliotropium cinerascens</i>	<i>Boraginaceae</i>	Asthama and Tooth ache
137.	<i>Hibiscus eriospermus</i>	<i>Malvaceae</i>	Tape worm and Gastritis
138.	<i>Hibiscus macranthus</i>	<i>Malvaceae</i>	Digestion problems and bone setting
139.	<i>Hibiscus macranthus</i>	<i>Malvaceae</i>	Gastritis, diabetes and Anemia
140.	<i>Hardeum vulgare</i>	<i>Poaceae</i>	Sun strike, generfta and Dyspepsia
141.	<i>Hyphaene thebaica</i>	<i>Arecaceae</i>	Evil eyes and diarrhea
142.	<i>Hypoestes forskalil</i>	<i>Acanthaceae</i>	Increases lactation and tape worm
143.	<i>Indigofera arrecta</i>	<i>Fabaceae</i>	Ringworm, laxative and stomach worms
144.	<i>Jacaranda mimosifolia</i>	<i>Bignoniaceae</i>	Warts and Bloating
145.	<i>Jasminum gradiflorum</i>	<i>Oleaceae</i>	Over-sweating, fever and madness
146.	<i>Juniperus procera</i>		Cold, intestinal parasites and Burns
147.	<i>Justicia cordata</i>	<i>Acanthaceae</i>	Purgative, dysentery and spider disease
148.	<i>Justicia schimperiana</i>	<i>Acanthaceae</i>	Antibiotics, Evil eyes and abscess
149.	<i>Kalanchoe marmorata</i>	<i>Crassulaceae</i>	Sun-strike, evil-eye and Retain placenta
150.	<i>Kigelia Africana</i>	<i>Bignoniaceae</i>	Purgative , tonic and stimulant
151.	<i>Kniphofia isoetifolia</i>	<i>Asphodolaceae</i>	Arthritis, Choilynichitis and Abscess
152.	<i>Lannea fructicosa</i>	<i>Anacardiaceae</i>	Intestinal worms, Bloating and Menstruation
153.	<i>Lantana camara</i>	<i>Verbenaceae</i>	TB, Hepatitis, Amoeba and taenia capitis
154.	<i>Lawsonia inermis</i>	<i>Lythraceae</i>	Endo parasites and cold
155.	<i>Leonotis ocimifolia</i>	<i>Lamiaceae</i>	Cough, gastritis and Arthritis
156.	<i>Lepidium sativum</i>	<i>Lepidium sativum</i>	Limb inflammation and abdominal pain
157.	<i>Leucas abussinica</i>	<i>Lamiaceae</i>	Inflammation of nails and gastritis
158.	<i>Leucas martinensis</i>	<i>Lamiaceae</i>	Constipation and digestive problem

159.	<i>Linum usitatissimum</i>	<i>Linaceae</i>	Bronchitis, diabetes and fever
160.	<i>Lycopersicum esculentum</i>	<i>Solanaceae</i>	Could, skin inflammation and haemorrhoids
161.	<i>Malus domestica</i>	<i>Rosaceae</i>	Eye problems and head ache
162.	<i>Mangifera indica</i>	<i>Anacardiaceae</i>	Arthritis and Eye problems
163.	<i>Matricoria chamomile</i>	<i>Asteraceae</i>	Fungal diseases and haemorrhoids
164.	<i>Maytenus arbutifolia</i>	<i>Celesteraceae</i>	Hypertension and dandruff
165.	<i>Maytenus senegalensis</i>	<i>Celesteraceae</i>	Hypertension, diabetes and emetics
166.	<i>Melia azadirachta</i>	<i>meliaceae</i>	Asthma, cough and bladder infection
167.	<i>Menthe poperita</i>	<i>Lamiaceae</i>	Gerefta and evil eyes
168.	<i>Meriandra dianthera</i>	<i>Lamiaceae</i>	Anemia, diarrhea and rheumatism
169.	<i>Mimusops kummel</i>	<i>Sapotaceae</i>	Constipation and digestion disorder
170.	<i>Mimusops schimperi</i>	<i>Sapotaceae</i>	Head ache and sun strike
171.	<i>Moringa oleifera</i>	<i>Moringaceae</i>	Tooth-ache and cough
172.	<i>Musa sapientum</i>	<i>Musaceae</i>	Antibiotic and gerefta
173.	<i>Nepeta azurea</i>	<i>Lamiaceae</i>	Tooth-ache, insects and snake bite
174.	<i>Nicandra physaloides</i>	<i>Solanaceae</i>	Digestion prob and bronchila prob
175.	<i>Nicotiana glauca</i>	<i>Solanaceae</i>	Abscess and myalgia
176.	<i>Nicotiana tabacum</i>	<i>Solanaceae</i>	Head ache, hypertension and vomiting
177.	<i>Nigella sativa</i>	<i>Ranunculaceae</i>	Cough malaria and hear disease
178.	<i>Nuxia cogesta</i>	<i>Loganiaceae</i>	Ring warm, dandruff
179.	<i>Ocimum basilicum</i>	<i>Lamiaceae</i>	Diuretic, abscess and wounds
180.	<i>Ocimum forskolei</i>	<i>Lamiaceae</i>	Anti-inflammatory, wound infection
181.	<i>Olea eruopeana/ cuspidate</i>	<i>Oleaceae</i>	Gastritis and constipation
182.	<i>Opuntia ficus-indica</i>	<i>Cactaceae</i>	Tonsillitis, arthritis and endo parasites
183.	<i>Ormocarpum pubescens</i>	<i>Fabaceae</i>	Uvulitis, abdominal-pain and tonsillitis
184.	<i>Osyris quadrpartita</i>	<i>Santalaceae</i>	Insect and fungal repellent
185.	<i>Otostegia fruticosa</i>	<i>Lamiaceae</i>	Gynecological problem and arthritis
186.	<i>Otostegia intefrifolia</i>	<i>Lamiaceae</i>	Eczema and abdominal pain
187.	<i>Ozoroa insignis</i>	<i>Anacardiaceae</i>	Heae ache, asthma and rheumatism
188.	<i>Pappea capensis</i>	<i>Sapindaceae</i>	Cancer, evil spirit and diarrhea

189.	<i>Parkinsonia aculeata</i>	<i>Fabaceae</i>	Eye infection and ear infection
190.	<i>Passiflora molissima</i>	<i>Passifloraceae</i>	Diarrhea, constipation and burn
191.	<i>Pavetta gardenifolia</i>	<i>Rubiaceae</i>	Urination, gastritis and tb
192.	<i>Petrolobium stellatum</i>	<i>Fabaceae</i>	Infectious dermatitis
193.	<i>Phoenix dactylifera</i>	<i>Arecaceae</i>	Ring worm and antibiotics
194.	<i>Phytolacca dodecandra</i>	<i>Phytolaccaceae</i>	Rheumatism, evile eye and snake venom
195.	<i>Piliostingma thonningii</i>	<i>Fabaceae</i>	Asthma and respiratory problems
196.	<i>Plantoga zeylanica</i>	<i>Plantaginaceae</i>	Choilynichitis and tonsillitis
197.	<i>Plumbago zeylanica</i>	<i>Plumbaginaceae</i>	Snake bite, and eye disease
198.	<i>Podocarpes latifolius</i>	<i>Podocarpaceae</i>	Digestion problems
199.	<i>Pittosporium viridifolium</i>	<i>Pittosporaceae</i>	Constipation
200.	<i>Pollichia campestris</i>	<i>Fabaceae</i>	Amoeba
201.	<i>Prosopis juliflora</i>	<i>Fabaceae</i>	Dysentery
202.	<i>Prunus persica</i>	<i>Rosaceae</i>	Anti-insect
203.	<i>Psiadia ppunctulata</i>	<i>Asteraceae</i>	Dandruff
204.	<i>Psidium guajava</i>	<i>Myrtaceae</i>	Uvulitis
205.	<i>Psydrax schimperiana</i>	<i>Rubiaceae</i>	Ectoparasites
206.	<i>Rhamnus prinoides</i>	<i>Rhamnaceae</i>	Scabies
207.	<i>Rhamnus staddo</i>	<i>Rhamnaceae</i>	Vomiting
208.	<i>Rhus glutinosa</i>	<i>Anacardiaceae</i>	Diarrhea
209.	<i>Rhus retinorrhoea</i>	<i>Anacardiaceae</i>	Fungal problems
210.	<i>Rhus natalensis</i>	<i>Anacardiaceae</i>	Tape worm
211.	<i>Rhus retinorrhoea</i>	<i>Anacardiaceae</i>	Hypertension
212.	<i>Ricinus communis</i>	<i>Euphorbiaceae</i>	itching
213.	<i>Rosa abyssinica</i>	<i>Risaceae</i>	Anti-cancer
214.	<i>Rosmarinus officinalis</i>	<i>Lamiaceae</i>	Coughing
215.	<i>Rumex nervosus polygonaceae</i>	<i>Polygonaceae</i>	Antibiotics
216.	<i>Rumex abyssinica</i>	<i>Polygonaceae</i>	purgative
217.	<i>Ruta chalepnensis</i>	<i>Rutaceae</i>	Malaria
218.	<i>Sageretia thea</i>	<i>Thamnaceae</i>	Abdominal pain

219.	<i>Salvadora schimperii</i>	<i>Salvadoraceae</i>	Antibiotics
220.	<i>Salvia schimperii</i>	<i>Lamiaceae</i>	Hepatitis
221.	<i>Schinus molle</i>	<i>Anacardiaceae</i>	Loss of appetite
222.	<i>Sclercarya birrea</i>	<i>Anacardiaceae</i>	Urination
223.	<i>Senna alexandrina</i>	<i>Fabaceae</i>	Loss of hair
224.	<i>Senna singueana</i>	<i>Fabaceae</i>	Snake and cancer
225.	<i>Sida schimperiana</i>	<i>Malvaceae</i>	Scabies
226.	<i>Solanum incanum</i>	<i>Solanaceae</i>	Eye cleaning
227.	<i>Solanum nigrum</i>	<i>Solanaceae</i>	Nerve problems
228.	<i>Solanum shcimperianum</i>	<i>Solanaceae</i>	Gastritis
229.	<i>Solanum tuberosum</i>	<i>Solanaceae</i>	Stabbing pain
230.	<i>Sorghum bicolor</i>	<i>Poaceae</i>	Bronchitis
231.	<i>Steganotaenia araliaceae</i>	<i>Apiaceae</i>	insecticide
232.	<i>Stereospermum kunthianum</i>	<i>Bignoniaceae</i>	Emetic
233.	<i>Suzugium auineens</i>	<i>Myrtaceae</i>	Stomach disorders
234.	<i>Tagetes minuta</i>	<i>Asteraceae</i>	Cough
235.	<i>Tamarindus indica</i>	<i>Fabaceae</i>	Fingers inflammation
236.	<i>Tamarix aphylla</i>	<i>Tamaricaceae</i>	Insecticides
237.	<i>Teclea nobilis</i>	<i>Rutaceae</i>	Hepatitis
238.	<i>Tragia pungent</i>	<i>Euphorbiaceae</i>	Asthma
239.	<i>Trichilia emetic</i>	<i>Meliaceae</i>	Intestinal worm
240.	<i>Terminalia brownii</i>	<i>Combretaceae</i>	Sun strike
241.	<i>Trigonella foenum- graecum</i>	<i>Facaceae</i>	Allergies
242.	<i>Vangueria madagascariensis</i>	<i>Tubiaceae</i>	Sprain
243.	<i>Verbascum sinaiticum</i>	<i>Scrophulariaceae</i>	Skin burns
244.	<i>Vernonia amygdalina</i>	<i>Asteraceae</i>	Cold
245.	<i>Vernonia schimperi</i>	<i>Asteraceae</i>	Arthritis
246.	<i>Vicia faba</i>	<i>Fabaceae</i>	Abdominal pain
247.	<i>Vitis vinifera</i>	<i>Vitaceae</i>	Antibiotic
248.	<i>Withania samnifera</i>	<i>Solanaceae</i>	Arthritis

249.	<i>Xanthium strumarium</i>	<i>Asteraceae</i>	Abdominal helminthes
250.	<i>Ximenia Americana</i>	<i>Olacaceae</i>	Antibiotic
251.	<i>Zea mays</i>	<i>Poaceae</i>	Kidney problem
252.	<i>Zehneria scabra</i>	<i>Cucurbitaceae</i>	Tapeworm expectorant
253.	<i>Zingiber officinale</i>	<i>Zingiberaceae</i>	Menstruation
254.	<i>Ziziphus abyssinica</i>	<i>Rhamnaceae</i>	Cough
255.	<i>Ziziphus mucronata</i>	<i>Rhamnaceae</i>	Sauna
256.	<i>Ziziphus spina-christi</i>	<i>Rhamnaceae</i>	Fugal migraine

**TABLE-2. PLANTS USED IN TRADITIONAL MEDICINE AND THE DRUGS DERIVED FROM THEM**

PLANT	DRUG
<i>Adhatoda vasica</i>	Vasicine
<i>Ahoni vernalis</i>	Adoniside
<i>Aesculus hippocastanum</i>	Aescin
<i>Agrimonia eupatoria</i>	Agrimophol
<i>Ammi majus</i>	Xanthotoxin
<i>Ammi visnaga</i>	Khellin
<i>Anabasis aphylla</i>	Anabesine
<i>Ananas comosus</i>	Bromelain
<i>Anamirta cocculus</i>	Picrotoxin
<i>Andrographis paniculate</i>	Andrographolide Neoandrographolide
<i>Anisodus tanguticus</i>	Anisodamine Anisodine
<i>Areca catechu</i>	Arecoline
<i>Ardisia japonica</i>	Bergenin
<i>Artemisia maritime</i>	Santonin
<i>Atropa belladonna</i>	Adropine
<i>Berberis vulgaris</i>	Berberine
<i>Brassica nigra</i>	Allyl isothiocyanate
<i>Camellia sinensis</i>	Caffeine theophylline
<i>Cannabis sativa</i>	$\Delta^9$ - Tetrahydrocannabinol
<i>Carica papaya</i>	Chymopapain Papain
<i>Cassia acutifolia</i>	Senosiders A & B
<i>Cassia angustifolia</i>	Senosiders A & B

<i>Cassia species</i>	Danthron
<i>Catharanthus roseus</i>	Vinblastine Vincristine
<i>Centella asiatica</i>	Asiaticoside
<i>Cephaelis ipecacuanha</i>	Emetine
<i>Chondodendron tomentosum</i>	Tubocurarine
<i>Cinchona ledgeriana</i>	Quinidine Quinine
<i>Cinnamomum camphora</i>	Camphor
<i>Cissampelos pareira</i>	Cissampeline
<i>Citrus species</i>	Hesperidin Rutin
<i>Colchicum autumnale</i>	Colchiceine amide Colchicines Demecolcine
<i>Convallaria majalis</i>	Convallatoxin
<i>Coptis japonica</i>	Palamtine
<i>Corydalis ambigua</i>	(±)- Tetrahydropalmatine
<i>Crotalaria sessiliflora</i>	Monocrotaline
<i>Curcuma longa</i>	Curcumin
<i>Cynara scolymus</i>	Cynarin
<i>Cytisus scoparius</i>	Sparteine
<i>Daphe genkwa</i>	Yuanhuacine Yuanhuadine
<i>Datura metel</i>	Scopolamine
<i>Digenia simplex</i>	Kainic acid
<i>Digitalis lanata</i>	Acetyldigoxin Deslanoside Digoxin Lanatosides A, B, C
<i>Digitalis purpurea</i>	Digitalin Digitoxin Gitalin
<i>Ephedra sinica</i>	Ephedrine Pseudoephedrine Pseudoephedrine, nor-
<i>Erythroxylum coca</i>	Cocaine
<i>Fraxinus rhynchophylla</i>	Aesculetin
<i>Gaultheria procumbens</i>	Methyl salicylate
<i>Glaucium glabra</i>	Glycyrrhizin

<i>Gossypium species</i>	Gossypol
<i>Hemsleya amabilis</i>	Hemsleyadin
<i>Hydrangea macrophylla</i> Var. <i>Thunbergii</i>	Phyllodulcin
<i>Hydrastis Canadensis</i>	Hydrastine
<i>Hyoscyamus niger</i>	Hyoscyamine
<i>Larrea divaricata</i>	Nordihydroguaiaretic acid
<i>Lobelia inflata</i>	$\alpha$ – Loneline
<i>Lonchocarpus nicou</i>	Torenone
<i>Lycoris squamigera</i>	Galanthamine
<i>Menthe species</i>	Menthol
<i>Nicotiana tabacum</i>	Nicotine
<i>Ocotea glaziovii</i>	Glaziovine
<i>Papaver somniferum</i>	Codeine Morphine Noscapine Papaverine
<i>Pausinystalia yohimba</i>	Yohimbine
<i>Physostigma venenosum</i>	Physostigmine
<i>Pilocarpus jaborandi</i>	Pilocarpine
<i>Piper methsticum</i>	Kawain
<i>Podophyllum peltatum</i>	Etoposide <sup>b</sup> Podophyllotoxin Teniposide <sup>b</sup>
<i>Potentilla fragarioides</i>	(+) –Catechin
<i>Quisqualis indica</i>	Quisqualic acid
<i>Rauvolfia canescens</i>	Deserpidine
<i>Rauvolfia serpentine</i>	Ajmalicine Rescinnamine Reserpine
<i>Rhododendron molle</i>	Rhomitoxin
<i>Rorippa indica</i>	Rorifone
<i>Salix alba</i>	Salicin
<i>Sanguinaria Canadensis</i>	Sanguinarine
<i>Silybum marianum</i>	Silymarin
<i>Simarouba glauca</i>	Glaucarubin
<i>Sophora pachycarpa</i>	Pachycarpine
<i>Stephania sinica</i>	Rotundine

<i>Stephania tetrandra</i>	Tertandrine
<i>Stevia rebaudiana</i>	Stevioside
<i>Strophanthus gratus</i>	Ouabain
<i>Strychnos nux-vomica</i>	Strychnine
<i>Theobroma cacao</i>	Theobromine
<i>Thymus vulgaris</i>	Thymol
<i>Triachosanthes kirilowii</i>	Trichosanthin
<i>Urginea maritime</i>	Scillarin A
<i>Valeriana officinalis</i>	Valepotriates
<i>Veratrum album</i>	Protoveratrines A&B
<i>Vinca minor</i>	Vincamine
<i>Several plants</i>	Allantoin Benzyl benzoate Borneol Pinitol

**CONCLUSION:**

Scientists in developing countries are entering an era in which plants can be expected to occupy a prominent position in the list of national priorities. This type of drug research could lead to industrial development in the country where the discoveries are made. The source of starting materials is normally abundant and readily available since in most developing countries the flora remains virtually unexploited, and we believe that over the next two decades many useful drugs will be isolated from plants. The majority of these discoveries should and will be made by enthusiastic, energetic, and highly motivated scientists in developing countries.

**REFERENCES:**

1. Cotton CM (1996). Ethnobotany: Principles and applications. John Willy and Sons, Ltd. Chichister, England.
2. WHO (2003). African traditional Medicine: Our Culture, Our Future. African Health Monitor.
3. Richard MM(2006). Efficacy of Medicinal Plants used by communities around lake Victoria region and the Samburu against mycobacteria, selected bacteria and candida albicans.
4. Yemane B, Medhanie G (2016). Ethnobotanical study of medicinal plants in sub Zoba Debarwa, Zoba Debub, Eritrea. Eritrea journal of Science and engineering 2: 65-97.

5. Yemane B, Berhane Y, Surender Reddy K (2016). Ethnobotanical study of medicinal plants in Sub region Logo anseba, Region Gash Barka, Eritrea Journal of pharmacy and Biological Science IOSR-JPBS 11: 63-73.
6. Ghirmay S (2000). Traditional use of traditional medicinal plant is highland region of Eritrea, Agricultural University of Norway.
7. Tecleab G, Thomas K, Tesfalem R, Gebrehiwet M, Bereket T (2009). Antibacterial activity and phytochemical study of nine medicinal plants from Eritrea. Pharmacology online journal 3: 546-555.
8. Thomas K, Tecleab G, Gebrehiwet M, Bereker T (2007). Chemical composition and Antibacterial activity of Essential oils from salvia species of Eritea. Pharmacology online journal 3: 546-555.
9. Bein E, Habte B, Jaber A, Brine A, Tengnas B (1996). Useful trees and shrubs in Eritrea. Regional Soil Conservation unit. RSCU/Sida, Nairobi.
10. Amsalu n (2010). Ethno botanical study of medicinal plants used in Farta Wereda, Southern Gonder, Amhara region Ethiopia. University of Addis Ababa a Master's of Science thesis.
11. Kumar N, Wani ZA, Dhyani S (2015). Ethnobotanical study of the plants used by the local people of Gulmarg and its allied areas, Jammu & Kashmir, India. International Journal of Current Research in Bioscience and Plant biology 2 (9): 16-23.
12. Bernhofr A (2010). A brief review o bioactive compounds in plants, In: Bioactive compounds in plants- benefits and risks for man and animals, Oslo: The Norwegian Academy of Science and Letters, 11-17.
13. Sarker SD, Nahar L (2007). Chemistry for Pharmacy Student sGeneral, Organic and Narural Product Chemistry. England: John Wiley and Sons. 283-359.
14. Kar A. Pharmaocgnosy and pharmacobiotechnology (Revised- Expanded Second Edition).
15. New Age International Limited Publishers New Delhi; 2007, 332-600.
16. Firm R(2010). Nature's Chemicals. Oxford university Press, Oxford, 74-75.
17. Kar A (2010). Pharmaochnosy and Pharmacobiotechnology (Revised-Expanded Second Edition). New Age International Limited Publishers New Delhi, 332-600.
18. Augustin Scalbert, Ian T Johnson, Mike Saltmarsh (2005). Polyphenols: antioxidants and beyond 1-3. American journal of Clinicla Nutrition. 81: 215S-217S.

19. Martinez MJA, Lazaro RM, del Olmo LMB, Benito PB (2008). Anti-infectious activity in the anthemideae tribe. In: Attaur (Ed). Studies in Naturla Products Chemistry. 35: 45-516.
20. Maurya R, Singh G, Yadav PP (2008). Antiosteoporotic agents from natural sources. In: Atta-ur-Rahman (Ed). Studies in natural Products Chemistry. 35: 517-545.
21. Chopra A, Doiphode V (2002). Ayurvevdic medicine: Core concept, therapeutic principles and current relecance. Medical Clinics of North America. 86: 75-89.
22. Ram J, Motriaya P, Chanda S (2015). Phytochemical screening and reported biological activities of some medicinal plants of Gujarat region. Journal of Pharmacognosy and Phytochemistry. 4 (2)L 192-198.
23. Moteriya P, Satasiya R, Chanda S (2015). Screening of phytochemical constituents in some ornamental flowers of Saurashtra region. Journal of Pharacognosy and Phytoachemistry 3(5):112-120.
24. Wink M (2010). Introduction : Biochemistry, Physiology and Ecological Functions of Secondary Metabolites, Second Edition Annual Plant Reviews, 40.
25. Fabicant DS, Farnsworth NR (2001). The value of plants used in traditional medicine for drug discovery. Environ Health Perspective. 109: 69-75.
26. Taylor Leslie ND (2000). Plant Based Drugs and Medicines, Rain tree Nutrition.
27. Modak M, Dixit P, Londhe J, Ghaskadbi S, Paul T, Devasagayam A (2007). Indian Herbal Drugs used for the treatment of Diabetes. Journal of Clinicla Biochemistry & Nutrition. 40(3): 163-173.
28. Shakya AK, Shukla S (2011). Evaluationof hepatoprotective efficacy of Majoon-e-Dabeed-ul-ward against *acetaminophen* induced liver damage: A Unani herbal formulation. Drug development Research. 72(4): 346-352.
29. World Health Organisation Media Centre, Traditional Medicine.(2008).
30. Heinrich M (2000). Ethnobotany and its role in drug development, phytotherapy Res. 14(7): 479-488.
31. Shakya AK, Sharma N, Saxena M, watal G (2014). Preliminary Phytochemical screenignof six medicinal plants used in traditional medicine. International Journal of Pharmacy and Pharmaceutical Sciences. 6 (5): 539-542.
32. Yadav M, Chatterji S, Gupta SK, Watal G ( 2014). Preliminary phytochemical screening of six medicinal plants used in traditional medicine. International Journal of Pharmacy and Pharmaceutical Sciences. 6 (5): 435-542.

33. Patwardhan B, Vaidya ADB, Chorghade M (2004). Ayurveda and Natural Products Drugs Discovery. *Current Science/* 86(6): 789-799.
34. Dash B, Sharma BK. *Charak Samhita* (2001). 7<sup>th</sup> ed. Varanasi (India): Chaukhamba Sanskrit Series.
35. Clark AM (1996). Natural products as a source for New Drugs Pharmaceutical Research. *13* (8): 1133-1141.
36. Rates SMK (2001). Plants as source of drugs. *Toxicon*. 39: 603-613.
37. Priyadarshini K, Keerthi Aparajitha U (2012). Paclitaxel against *Cancer*: A short review. *Medicinal chemistry*. 2: 139-141.
38. Gajalakshmi S, Vijayalakshmi S, Devi Rajeswari V (2013). Pharmacological Activities of *Catharanthus roseus*: A Perspective review: *International Journal of Pharmaceutical and Biological Sciences*. 4 (2): 431-439.
39. Shiyou Li, Wanli Zhang (2014). Ethnobotany of *Camptotheca Decaisne*: New Discoveriies of Old Medicinal Uses. *Pharmaceutical Crops*. 5:140-145.
40. Vivek Sharma. (2013). Part Based HPLC-Pda Quantificatio of Podophyllotoxin in Populations of *Podophyllum hexandrum* Royle Indian Mayapple from Higher Altitude Himalayas. *Journal of Medicinal Plants Studies*. 1 (3): 176-183.
41. Akaram M, Shahab –uddin, Afzal Ahmed, Khan Usmanghani Abdul, Hannan Mohiuddin, Asif M (2010). *Curcuma Longa* and Curcumin: A review Article. *Romanian Journal of Biology- plant Biology*. 55 (2): 65-70.
42. Das SK, Mukherjee S, Vasudevan DM( 2008). Medicinal properties of milk thistle with special reference to silymarin: An Overview. *Natural Product radiance*. 7 (2): 182-192.
43. Jena J, Gupta AK (2012). *Ricinus Communis* Linn: A Phyto pharmacological review. *International Journal of Pharmacy and Pharmaceutical Sciences*. 4 (4): 25-29.
44. Rathinamoorthy R, Thilagavathi G (2014). *Terminalia Chebula*-review on Pharmacological and Biochemical Studies. *International Journal of Pharma Tech Research*. 6(1): 97-116.
45. Umadevi M, Rajeswari R, Sharmila Rahale C, Selvavenkadesh S, Pushpa R, Sampath KP *et al.* (2012). Traditional And Medicinal Uses of *Withania Somnifera*. *The Pharma Innovation*. 1 (9): 102-110.
46. Gupta SK, Sharama A (2014). Medicinal properties of *Zingiber officinale* Roscoe- A Review. *IOSR Journal Of Pharmacy and Biological Sciences*. 9(5): 124-129.

47. Kausik Biswas, Ishita Chattopadhyay, Ranaji K. Banerjee and Uday Bandyopadhyay (2002). Biological activities and medicinal properties of neem (*Azadirachta indica*). *Current Science*. 82 (11): 1336-1345.
48. Nisar Ahmad, Hina Fazal, Bilal Haider Abbasi, Shahid Farooq, Mohammad Ali, Mubarak Ali Khan (2012). Biological role of *Piper nigrum* L. (Black Pepper): A review. *Asian Pacific Journal of Tropical Biomedicine*. S1945-S1953.
49. Jitendra Mittal, Madan Mohan Sharma, Amla Batra (2014). *Tinospora cordifolia*: a multipurpose medicinal plant- A review. *Journal of Medicinal Plants Studies*. 2 (2): 32-47.
50. Pankaj K, Sahu Deen Dayal Giri, Ritu Singh, Priyanka Pandey, Sharmistha Gupta, Atul Kumar Shrivastava, *et al* (2013). Therapeutic and Medicinal Use of *Aloe vera*: A Review. *Pharmacology & Pharmacy* 4: 599-610.
51. Jain S, Mohan R, Rai R (2015). *Ocimum sanctum* as an Herbal Medicine: A Review. *International journal of Maxillofacial Research* 1(1): 1-12.
52. Seyyed MJ, Saied RF (2012). Therapeutic application of different parts *Berberis vulgaris*. *International Journal of Agriculture and Crop Sciences*. 4(7): 404-408.
53. Chauhan R, Km. Ruby, Dwivedi Jaya (2012). *Bergenia ciliata* Mine of Medicinal Properties: A Reviews. *International Journal of Pharmaceutical Sciences Review and Research* 15 (2): 20-23.
54. Negi JS, Bisht VK, Bhandari AK, Sundriyal RC (2012). Determination of mineral contents of *Digitalis purpurea* L. and *Digitalis lanata* Ehrh. *Journal of Soil Science and Plant Nutrition*. 12 (3): 463-469.
55. Parakh PM(2010). *Nigella sativa* Linn. A Comprehensive review. *I J of Natural Products and Resources*. 1 (4): 409-429.
56. Vincent PK, Titanji, Denis Zofou, Moses N Ngemenya (2008). The Antimalarial Potential of Medicinal Plants Used for the Treatment of Malaria in Cameroonian Folk Medicine. *African Journal of Tradition, Complementary and Alternative Medicine*. 5 (3): 302-321.
57. Sanjeev Krishnaa, Anne-Catrin Uhlemanna, Richard K. Haynesb (2005). Artemisinin: Mechanisms of action and potential for resistance. *Drug Resistance*. 233-244.
58. P. Joshi, V. Dhawan (2005). *Swertia chirayita* an overview. *Current Science*. 89(4): 635-640.
59. Singh VK, Singh DK (2008). Pharmacological Effects of Garlic (*Allium sativum* L.). *ARBS Annual Review of Biomedical Sciences*. 10:6-26.

60. Paarakh PM. Terminalia arjina (Roxb.) Wt. and Arn: A Review. Interanational Journal of Pharamcology. 6(5): 515-534.
61. Khan KH (2009). Roles of Emblica officinalis in Medicine- A Review/ Botany research International. 2 (4): 218-228.
62. Abu-darwish MS, Abu-Dieyeh ZH, Mufeed B, Al-Tawaha ARM, Al-Dalain SYA (2009). Trace element contents and essential oil yields from wild thyme plants (*Thymus serpyllum* L.) grown ar different natural variable environments. Jordan. J. Food Agr. Environ., 7(4), 920-924.
63. Feiberg L, Nordhergs GF and Vouk VB (1986). Hand book on the toxicology of metals 2<sup>nd</sup> ed. Vol.2, Amsterdam, Oxford:Elsevier Science Punllishers BV, New York. pp. 43.
64. Iqbal H, Khattak B, Ayaz S, Rehman A, Ishfaq M, Abbas MN, malik MS, Wahab A, Imran, Mehsud S (2013). Pollution based study of heavy metals in Medicinal Plants *Aloe vera* and *Tamarix aphylla*, J app Pharm. Sci, 3(4): 54-58.
65. Ishaq M, Rehman A, Adnan M, Ullah N, Ahmad I, Aamir M (2013). Comparative study of Heavy metals in Albizia lebbeck, collected from different environmental sites, int. J. Pharm. Sci. Rev. Res., 20(2), 5-9.
66. Islam E, Yang X, He Z and Mahamood Q (2007). Assessing potential dietary toxicity of Heavy metals in selected vegetables and food crops. J. Zhejiang Univ. Sci. 8(1): 1-13.
67. Itanna F (2002). Metals in leafy vegetables grown in addis ababa and toxicological implications. Ethiop. J. Health Dev., 16 (3): 295-302.
68. Khan SA, Khan L, Hussain I, Marwat KB and Akhtar N(2002). Profile of heavy metals in selected medicinal plants. Park J Weed Sci. Res., 14(1-2), 101-110.
69. Lasisi AA, Ejelonu BC, Nwosu FO, Olayiwola MA and Yusuff AA (2006). Heavy metals and macronutrients contents in selected herbal plants of south-western Nigeria. Hamdard Medicus. XLIX (4), 71-76.
70. Martins TCG, De Nadai Farnandas EA, Ferrari AA Tagliaferro FS and Bacchi MA (2008). Chemical characterization of agricultural supplies applied to organic tomato cultivation. J Radioanal Nucl Chem. 278(2): 517-520.
71. Obi E, Akunyili DN, Ekpo B and Orisakwe OE (2006). Heavy metal hazards of Nigerian herbal remedies. Sci. Total Environ. 369(1-3), 35-41.

72. Rehman A, Iqbal H, Rehman H, Iqbal T, Ullah W, Ranf MK, Jabbar A, Shagufta BI, Ullah S, Ahmad, I (2013). Study of heavy metals in medicinal plant *Solanum xanthocarpum*. International Journal of Science Innovations and Discoveries. 3(2): 254-260.
73. Rehman A, Ullah H, Khan RU, Ahmad I (2013). Population based study of heavy metals in medicinal plant, *Capparis decidua*, Int, J. Pharm. Pharm, Sci. 5(1): 108-113.
74. Singh V and Garg AN (2006). Availability of essential trace elements in Indian cereals, vegetables, and spices using INAA and the contribution of spices to daily dietary intake. Food Chemistry. 94 (1): 81-89.
75. WHO (1993). Evaluation of certain food additives and contaminants, forty-first Report of the Joint FAO/WHO Expert Committee on Food Additives. WHO Technical Report Series, No. 837, World Health Organization, Geneva. pp. 28-35.
76. WHO (2007). Health risks of heavy metals from long range transboundary air pollution. Joint WHO/Convention task Force on the Health Aspects of Air Pollution, SCherfigheg 8 DK-2100 Copenhagen, Denmark, World Health Organization. pp . 1-70.
77. American Cancer Society. Cancer facts & figures. Atlanta: American cancer society ; 2010
78. Boivin JF (1990). Second Cancers and other late Side Effects of Cancer Treatment. Cancer 65(S3): 770-5.
79. Yates JS, Mustian KM, Morrow GR, Gillies LJ, Padmanaban D, Atkins JN, Issell B, Kroshner KK, Colman LK (2005). Prevalence of complementary and alternative medicine use in cancer patients during treatment. Support Care Cancer. 13 (10): 806-11.
80. Sri PU, Sree NV, RevathiS, Kumar YVVA, Sri ND (2010). Role of herbal medicines in cancer. Int J Pharm Sci Res. 1(11): 7-21.
81. Devmurari VP, Pandey S, Goyani MB, Jivani NP, Matottrao S, Sivakumar P (2010). Evaluation of Anticancer Activity of *Adiantum venustum* (Don). Int J Pharmacog Phy Res. 2(1): 5-10.
82. Mubashir S, Shah WA (2011). Phytochemical and Pharmacological Review Profile of *Adiantum venustum*. Int J Pharm Tech Res. 3(2): 827-30.
83. Gul MZ, Bhakshu LM, Ahmad F, Kondapi AK, Qureshi IA, Ghazi IA (2011). Evaluation of *Abelmoschu moschtus* extracts for antioxidant , free radical scavenging, antimicrobial and antiproliferative activities using *In vitro* assays. BMC complement Altern Med. 11(1):64.

84. Kohn LK, Pizao PE, Foglio MA, Antonio MA, Amaral MCE, Bittric V *et al* (2006). Antiproliferative activity of ctude extract and fractions obtained from *Aspidospermatomentosum* Mart. Rev. Bras. PI Med Botucatu. 8(esp):110-5.
85. Andrea L, Holguin MF, Holguin O, Micheletto S, Goehle S, Julian A, Mary A, Connell O (2008). Chemotypic Variation of Essential Oils in the Medicinal Plant *Anemopsis californica* . Phytochemistry. 69(4): 919-27.
86. Catherine N, Seth KL, lowrey T, Guerra L, slambrouck SV, WIm FA (2010). *In vitro* anti-cancer activity of *Anemopsis californica* Oncol Lett. 1(4):711-15.
87. Childs RF, Cole JR (1965). Phytochemical an dpharmacological investigation of *Anemopsis californica* J. Pharm Sci. 54(5): 789-91.
88. Laizuman N, Ronok Z, Ashik M, Saiful I, Anamul H, Abul F, Mele J (2012). Antioxidant and antitumor activity of chloroform extract of of *Alangium salvifolium* flowers. Phytopharmacol. 2(1): 123-34.
89. Ronok Z, Badrul AM, saiful IM, Gopal SC, Nargis CS, salman HB, Mosaddik MA, Mele J Ekramul HM (2011). Anticancer avtivity of *Alangium salvifolium* flower in Ehrlich ascites carcinoma bearing mice. J Cancer Res. 7(3):254-62.
90. Shurma AK, Agarwal V, Kumar R, Balasubramaniam A, Mishra A, Gupta R (2011). Pharmaceutica Drug Res. 68(6): 897-904.
91. Prasad L, Khan TH, Tamanna J, Sarwat S (2006). *Acorus calamus* extracts and nickelchloride, prevention of oxidative damage and hyperproliferation response in rat kidney. Boil Trace Elem Res. 113(1):77-91.
92. Rajkumar V, Guha G, Kumar AR, Mathew L (2009). Evaluation of cytotoxic potential of *acours clalamus* thizome. Ethnobotanical leaflets. 13: 832-9.
93. Gyawali R, Kim K (2009). Volatile organic compounds of medicinal values from Nepalese *Acorus calamus*. J Sci Engg Tech 5(2): 51-65.
94. Palani S, Raja S, Kumar RP, Venkdesan D, Devi K, Sivaraj A, Senthil BK (2009). Therepeutic efficacy of antihepatotoxic and antioxidant activities of *Acorus calamus* on acetaminophen-induced toxicity in rats. Int J Integr Biol. 7(1): 39-44.
95. Singh R, Shrama, PK, Rishabja M (2011). Pharmacological properties and ayurvedic value of Indian buch plant (*Acorus calamus*): A short Review. Adv Biol Res. 5(3): 145-54.

96. Kuete V, Vouffo B, Mhaveng AT, Vouff EY, Siagat RM, Dongo E (2009). Evaluation of *Antiaris Africana* methanol extract and compounds for antioxidant and antitumor activities. *Pharm Biol.* 47(11): 1942-9.
97. Vouffo B, Etienne D, Petrea F, Andrea T, George S, Armin M, *et al* (2010). Antiaroyl cinnamate and africanoside, a cinamoyl triterpene and a hydroperoxy-cardenolide from the stem bark of *Antiaris Africana*. *Planta Medica.* 76(15): 1717-23.
98. Rabi T, Karunagaran D, Nair MK, Bhattathiri VN (2002). Cytotoxic activity of amooranin and its derivatives. *Phytother Res.* 16(S1):84-6.
99. Rabi T, liming W, Banerjee S (2007). Novel triterpenoid 25-hydroxy-3-oxoolean-12-en-28-oic acid induces growth arrest and apoptosis in breast cancer cells. *Breast Cancer Res Treat* 101(1):27-36.
100. Rabi T, Ramachandran C, Fonseca HB, Nair RPK, Alamo A, Melinck SJ, Escalon E(2003). Novel drug amooranin induces apoptosis through caspase activity in human breast carcinoma cell lines. *Breast Cancer Res Treat.* 80(3): 321-30.
101. Chan LL, Shrine G, Irfan A, Saujanya GL, Atiya A, Cunningham BT, Watikin KL (2011). Cytotoxicity effects of *Amoora tohituka* and *chttagonga* on Breast and Pancreatic Cancer Cells. *Evid Mased Complement ALternat Med Article ID 860605*, 8 pages.
102. Cheppail R, Thangaiyan R, Fonseca HB, Steven MJ, Enrique AE (2003). Novel plant triterpenoid drug ammoranin overcomes multidrug resistance in human leukemia and colon carcinoma cell lines. *Int J Cancer.* 105(6): 784-9.
103. Rabi T, Banerjee S (2009). Novel semisynthetic triterpenid AMR-Me inhibits telomerase activity in human leukemic CEM cells and exhibits in vivo antitumor activity against Dalton's lymphoma ascited tumor. *Cancwe Lett.* 278(2): 156-63.
104. Thangapazham RL, Singh AK, Seth P, Misra N, Mathad VT, Raj K, *et al* (2008). Shikonin analogue (SA) 93/637 induces apoptosis by activation of caspase-3 in U937 cells. *Front Biosci* 13: 561-8.
105. Bibi Y, Nisa S, Zia M, Waheed A, Ahmed S, Chaudhary MF (2012). Adenocarcinoma cell Line (MCF-7) and phytochemical analysis. *Pak J Pharm Sci* 25(1): 183-7.
106. Zhang Z, Li S, Lian XY (2010). An overview of genus *Aesculus* L.: Ethnobotany, Phytochemistry, and Pharmacological activities. *Pharm Crops.* 1:24-51.

107. Jagetia GC, Venkatesh P, Baliga MS (2005). *Aegle marmelos* (L). correa inhibits the Proliferation of Transplanted Ehrlich Ascites Carcinoma in Mice. *Biol Pharm Bull* 28(1): 58-64.
108. Lampronti ID, Martello N, Bianchi M, BOrgatti E, Lambertini R, Piva S, *et al* (2003). *In vitro* antiproliferative effects on human tumor cells lines of extracts from the Bangladeshi medicinal Plant *Aegle marmelos* Correa. *Phytomedicine*. 10(4):300-8.
109. Gangadevi V, Muthumary J, Taxol (2008). An anticancer drug produced by an endophtic fungus *Bartalinia tobillardoides* Tassi, isolated from a medicinal plant, *Aegle marmelos* Correa ex Roxb. *World J Microbiol Bioltechnol*. 24 (5): 717-24.
110. Subramaniam D, Giridharan P, Murmu N, Shankaranarayanan NP, May R, Houchen CW *et al* (2008). Activation of apoptosis by 1-Hydroxy-5,7-Dimethoxy-2-Naphthalene Carnoxaldehyde, a Novel Compound from *Aegle marmelos*. *Cancer Res*. 68(20): 8573-81.
111. Khan TH, sultana S (2011). Effect of *Aegle marmelos* on DEN initiated and 2-AAF promoted hepatocarcinogenesis: a chemopreventive study. *Toxicol Mech Methods*. 21(6): 453-62.
112. Islam MS, Kusumoto Y, Al-Mamun MA (2011). Cytotoxicity and Cancer (HeLa) Cell killing Efficacy of Aqueous Garlic (*Allium sativum*) Extract. *J Sci Res*. 3(2): 375-62.
113. Thomson M, Ali M (2003). [*Allium sativum*]: a review of its potential use as an anti-cancer agent. *Curr Cancer Drug Targets*. 3(1): 67-81.
114. Karmakar S, Choudhury SR, Banik NL, Swapan RK (2011). Molecular Mechanisms of Anti-cancer Action of Garlic Compounds in Neuroblastoma. *Anticancer Agents Med Chem* 11(4): 398-407.
115. Shukla Y, Kalra N (2007). Cancer chemoprevention with garlic and its constituents. *Cancer Lett*. 247(2): 167-81.
116. Guruvayoorappan C, Kuttan G (2007). Immunomodulatory and antitumor activity of *Biophytum sensitivum* extract. *Asian Pacific J Cancer Prev* 8(1): 27-32.
117. Bhaskar VH, Rajakshmi V, Anti-tumor activity of aqueous extract of *Biophytum sensitivum* Linn. *Annals Biol Res*. 1(3): 76-80.
118. Guruvayoorappan C (2007). Apoptotic Effect of *Biophytum sensitivum* on B16F-10 cells and its Regulatory Effects on Nitric Oxide and Cytokine Productionon Tunor-Associated Macrophages. *Integr Cancer Ther*. 6(4): 373:80.

119. Guruvayoorappan C (2008). *Biophytum sensitivum* (L.) DC inhibits Tumor Cell invasion and Metastasis Through a Mechanism Involving Regulation of MMPs, Prolylhydroxylase, Lysyl Oxidase, nm23, ERK-1, ERK-2, STAT-1, and proinflammatory Cytokine Gene Expression in Metastatic Lung Tissue. *Integr Cancer Ther.* 7(1): 42-50.
120. Cichwicz RH, Kouzi SA (2004). Chemistry, biological activity, and chemotherapeutic potential of betulinic acid for the prevention and treatment of cancer and HIV infection. *Med Res Rev* 24 (1): 90-114.
121. Chatterjee D, Sahu RK, Jha AK, Dwivedi J (2011). Evaluation of Antitumor Activity of *Cuscuta Reflexa* Roxb (Cuscutaceae) Against Ehrlich Ascites Carcinoma in Swiss Albino Mice. *Trop J Pharm Res.* 10(4): 447-54.
122. Suresh V, sruthi V, padmaa B, asha VV (2011). *In vitro* anti-inflammatory and anti-cancer activities of *Cuscuta reflexa* Roxb. *J Ethnopharmacol.* 134(3): 872-7.
123. Gupta M, Mazumder UK, Kumar RS, Sivakumar T, Vamsi ML (2004). Antitumor activity and antioxidant status of *Caesalpinia bonducella* against Ehrlich ascites carcinoma in Swiss albino mice. *J Pharmacol Sci* 94(2): 177-84.
124. Gupta M, Mazumder UK, Rath N, Mukhopadhyay DK (2000). Antitumor activity of methanolic extract of *Caesalpinia bonducella* against Ehrlich ascites carcinoma activities of *Carcinoma*. *J Ethnopharmacol* 72(1): 151-6.
125. Rejuya CS, Cibin TR, Annie A (2009). Leaves of *Cassia tora* as a novel cancer therapeutic –An *in vitro* study. *Toxicol in Vitro.* 23(6): 1034-8.
126. Bala A, Kar B, Pallab KH, Mazumdar UK, Bera S (2010). Evaluation of anticancer activity of *Cleome gynandra* Ehrlich Ascites Carcinoma treated Mice. *J Ethnopharmacol.* 129(1): 131-4.
127. Chul PB, Kefa BO, Eung-Seok L, Soo LY, Jung-Ae K (2005). Asiatic acid induces Apoptosis in SK-MEL-2 Human Melanoma cells. *Cancer Lett.* 218(1): 81-90.
128. Cho CW, Choi DS, Cardone MH, Kim CW, Sinskey AJ, Rha C (2006). Glioblastoma cell death induced by Asiatic acid. *Cell Biol Toxicol* 22(6):393-408.
129. Heidari M, Heidari-Vala H, Sadeghi MR, Akhondi MM (2012). The inductive effects of *Centella asiatica* on rat spermatogenic cell apoptosis *in vivo*. *J Nat Med.* 66(2): 271-8.
130. Rai N, Agrawal RC, Khan A (2011). Chemopreventive Potential of *Centella asiatica* on B6F10 Melanoma Cell Lines in Experimental Mice. *Pharmacologyonline* 1: 748-58.

131. Suboj B, Jose P, Priya PS, Vinod V, Karedath AAT, Priya S, Srinivas G (2009). Apoptosis induction of *Centella Asiatica* on Human Breast cancer Cells. Afr J trad Complement Altern Med. 6(1): 9-16.
132. Susi E, Jaksa S, Marsiati H, Fauziah O, Rahmat A (2011). Effect of cola nut (*Colanitida*) on the apoptotic cell of human breast carcinoma cells lines. J Med Plants Res. 5(11): 2393-7.
133. Liu S, Luo X, Li D, Zhang J, Qiu D, Liu W, She L, Yang Z (2006). Tumor inhibition and improved immunity in mice treated with flavones from *Cirsium japonicum* DC. Int Immunopharmacol. 6(9): 1387-93.
134. Liu S, Zhang J, Li D, Liu W, Luo X, Zhang R, Li L, Zhao J (2007). J Anticancer activity and quantitative analysis of flavones of *Cirsium japonicum* DC. Nat Prod Res. 21(10): 915-22.
135. Yin Y, Heo S, Wang MH (2008). Antioixdant and Anticancer activities of Methanol and water Extracts from Leaves of *Cirsium japonicum* J Appl Biol Chem. 51(4): 160-4.
136. Jin Y, Lu Z, Cao K, Zhu Y, Chen Q, Zhu F, Qian C, Pan J (2010). The antitumor activity of Homoharringtonine against Human Mast Cells Harboring the KIT D816V Mutation Mol Cancer ther. 9(1): 211-23.
137. Meng H, Yang C, Jin J, Zhou Y, Qian W (2008). Homoharringtonine inhibits the AKT pathway and induces *in vivo* cytotoxicity in human multiple myeloma cells. Leuk Lymphoma. 49(10):1954-62.
138. Ni D, Ho DH, Vijeswarapu M, Felix E, Rhea PR, Newman RA (2003). Metabolism of homoharringtonine, a cytotoxic component of the evergreenplant *Cephalotaxus harringtonia*. J Exp Ther Oncol. 3(1):47-52.
139. Bakshi HA, Sam S, Anna F, Zeinab R, Ahmad SG, Sharma M (2009). Crocin from Kashmiri Saffron (*Crocus sativus*) induces *in vitro* and *in Vivo* Xenograft Growth Inhibition of Dalton's Lymphoma (DLA) in Mice. Asian Pacific J Cancer Prev. 10 (5):887-90.
140. Fikrat AL (2002). Cancer Chemopreventive and Tumoricidal Properties of Saffron (*Crocus sativus* L). Exp Biol Med. 227(1): 20-5.
141. Kumar PK (2006). Protective effect of saffron (*Crocus sativus* L.) aqueous extract against genetic damage induced by anti-tumor agents in mice. Hum Exp Toxicol. 25(2): 79-84.

142. Gomez-Flores R, Martinez HH, Guerra PT, Guerra RT, Licea RQ, Enriqueta CM, Padila CR (2010). Antitumor and immunomodulating potential of *Coriandrum sativum*, *Piper nigrum* and *Cinnamomum zeylanicum*. J Nat Prod. 3:54-63.
143. Huang TC, FU HY, Ho Ct, Tan D, Huang YT, Pan MH (2007). Induction of apoptosis by cinnamaldehyde from indigenous cinnamon *Cinnamomum osmophloeum* Kaneh through reactive oxygen species production, glutathione depletion, and caspase activation in human leukemia K562 cells. Food Chem. 103 (2): 434-43.
144. Singh R, Koppikar SJ, Paul P, Gilda S, Paradkar AR, Ghanekar Rk (2009). Comparative analysis of cytotoxic effects of aqueous cinnamon extract from *Cinnamomum zeylanicum* bark with commercial cinnamaldehyde on various cell lines. Pharm. Biol 47(12):1174-9.
145. Koppikar SJ, Choudhari AS, Suryavanshi SA, Kumari S, Chattopadhyay S, Kaul-Ghanekar R (2010). Aqueous Cinnamon Extract (ACE-c) from the bark of *Cinnamomum cassia* Causes apoptosis in human cervical cancer cell line (SiHa) through loss of mitochondrial membrane potential. BMC Cancer. 10(1):210.
146. Csupor – Löffler B, Hajdu Z, Zupko I, Molnar J, Forgo P, Vasas A, Kele Z, Hohmann J (2011). Antiproliferative Constituents of the Roots of *Conyza Canadensis* Planta Medica. 77(11):1183-8.
147. Maneerat W, Rha-in S, Cheenpracha S, Prawat U, Laphookhieo S, New amides from the seeds of *Clausena lansium*. J Med Plants Res. 5:2812-5.
148. Prasad KN, Hao J, Yi C, Zhang D, Qiu S, Jiang Y, Zhang M, Chen F (2009). Antioxidant and Anticancer Activities of Wampee (*Clausena landium* (Lour). Skeels) Peel. J Biomed Biotech. Article ID 612805, 6 pages.
149. Motta FCM, Santos DYAC, Salatino MLF, Almeida JMD, Negri G, Carvalho JE, Ruiz ALTG, Ines C, Salatino A (2011). Constituents and antiproliferative activity of extract from leaves of *Croton macrobothrya*. Rev bras farmacogn. 21(6):972-7.
150. Wang X, Xu Y, Yang M, Hong Z (2011). Chloroform extract of Tibetan herbal medicine *Dracocephalum tanguticum* Maxim . inhibits proliferation of T98G glioblastomas cells by modulating Caspase-3 cleavage and expression of Bax and p21. J. Med Plants Res. 5(25): 6024-31.
151. Hu M, Xu L, Yin L, Qi Y, Li H, Xu Y, Han X, Peng J, Wan X (2011). Cytotoxicity of dioscinin human gastric carcinoma cells through death receptor and mitochondrial pathways. J. Appl Toxicol. 33(8):712-22.

152. Wang G, Chen H, Huang M, Wang N, Zhang J, Zhang Y, Bai G, Fong QF, Yang M, Yao X (2006). Methyl Protodioscin induces G2/M cell cycle arrest and apoptosis in HepG2 liver cancer cells. *Cancer Lett.* 241(1): 102-9.
153. Cibin TR, Srinivas G, Gayathri DD, Priya S, Lija Y, Annie A (2006). Antioxidant and Antiproliferative Effects of Flavonoids from *Emilia sonchifolia* Linn on Human Cancer Cells. *Int J Pharmacol.* 2(5):520-4.
154. Sidambaram RR, Dinesh MG, Jayalakshmi ET (2011). *An in vitro* Study of Cytotoxic Activity of *Euphorbia Hirta* on Hep2 Cells of Human Epithelioma of Larynx. *Int J Pharm Sci.* 3(Supple 3): 101-3.
155. Guo Z, Xu Y, Han L, Bo X, Huang C, Ni L (2011). Antioxidant and cytotoxic activity of the acetone extracts of root of *Euphorbia hulonoma* its ellagic acid derivatives, *J Med Plant Res.* 5(23): 5584-9.
156. Baliga MS, Dsouza JJ (2011). Amla (*Embllica officinalis* Gaertn), a wonder berry in the treatment and prevention of cancer. *Eur J Cancer Prev* 20(3):225-39.
157. Poojari R, Gupta S, Maru G, Khade B, Bhagwat S (2010). Chemopreventive and Hepatoprotective Effects of Embelin on N-Nitrosodiethylamine and Carbon Tetrachloride induced Preneoplasia and Toxicity in Rat Liver. *Asian Pac J Cancer Prev.* 11(4): 1015-20.
158. Yang CJ, Wang CS, Hung JY, Huang HW, Chia YC, Wang PH, Weng CF, Huang MS (2009). Pyrogallol induces G2-M arrest in human lung cancer cells and inhibits tumor growth in an animal model. *Lung Cnacer.* 66(2): 162-8.
159. Balijepalli MK, Tandra S, Pichika MR (2010). Antiproliferative activity and induction of apoptosis in estrogen receptor-positive and negative human breast carcinoma cell lines by *Gmelina asiatica* roots. *Pharmacog Res.* 2(2):113-9.
160. Noudeh GD, Shrififar F, Noodeh AD, Moshfi MH, Afzadi MA, Behravan E, *et al* (2010). Antitumor and antibacterial activity of four fractions from *Heracleum Persicum* Dest. And *Cinnamomum zeylanicum* Blume. *J Med Plants Res.* 4(21): 2176-80.
161. Lam SK, Ng TB (2009). Novel galactonic acid-binding hexameric lectin from *Hibiscus mutabilis* seeds with antiproliferative and potent HIV-1 reverse transcriptase inhibitory activities. *Acta Biochimica Plonica.* 56(4):649-54.
162. Lin CC, Kuo CL, Lee MH, Hsu SC, Huang AC, Tang NY, Lin JP, Yang JS, Lu cc, Chiang JH, Chueh FS, Chung JG (2011). Extract of *Hedyotis diffusa* Willd influences Murine

- Leukemia WEHI-3 Cells *in vivo* as well as Promoting T-and B-Cell Proliferation in Leukemic Mice. *In vivo* 25(4): 633-40.
163. Shao J, Gong G, Trombetta L (2011). An Evidence-based Perspective of *Hedyotis Diffusa* or *Oldenlandia Diffusa* (Spreading Hedyotis) for Cancer Patients. Evidencebased Anticancer Complementary and Alternative Medicine, Evidence-based Anticancer Materia Medica 179-92.
164. Wang JH, Shu LH, Yang LL, Meng Z, Ping H, 2-Hydroxy -3-methylantraquinone from *Hedyotis diffusa* Wild induces Apoptosis via Alteration of Fas/FasL and Activation of Caspase-8 in Human Leukemic THP-1 cells. *Archive Med Res.* 42(7): 577-83.
165. Kumar RS, Raj Kapoor B, Perumal P. Antitumor and cytotoxic Activities of Methanol Extract of *Indigofera linnaei* Ali, Asian Pacific J Cancer Prev. 2011;12(3):613-8.
166. Wong YH, Abdul KH, Ling SK (2012). Bioassay-Guided Isolation of Cytotoxic Cycloartene Triterpenoid Glycosides from the Traditionally Used Medicinal Plant *Leea indica*. *eCAM* 2012: Article ID 164689, 11 pages,
167. Raihan MO, Tareq SM, Brishti A, Alam MK, Haque A, Ali MS (2012). Evaluation of Antitumor Activity of *Leea indica* (Burm.f.) Merr. Extract against Ehrlich Ascites Carcinoma (EAC) Bearing Mice. *Am J Biomed Sci.* 4(4): 143-32.
168. Wang L, Xu GF, Liu XX, Chang AX, Xu ML, Ghimeray AK, Piao JP, Cho DH (2012). *In vitro* antioxidant properties and induced G2/M arrest in HT-29 cells of dichloromethane fraction from *Liriodendron tulipifera*. *J Med Plants Res.* 6(3):424-32.
169. Moon MK, Oh HM, Kwon BM, Beak NI, Kim SH, Kim JS, Dae Keun Kim DK (2007). A Farnesyl Protein Transferase and Tumor Cell Growth Inhibitory Activities of Lipiferolide isolated from *Liriodendron tulipifera* *Arch Pharm Res.* 30(3):299-302.
170. Bhoopat L, Srichairatanakool S, Kanjanapothi D, Taesorikul T, Thananchai H, Bhoopat T (2011). Hepatoprotective effects of lychee (*Litchi chinensis* Sonn): A combination of antioxidant and anti-apoptotic activities. *J Ethnopharmacol.* 136(1): 55-56.
171. Roya S, Besra SE, Deb T, Banerjee B, Mukherjee J, Vedasiromoni JR (2008). Induction of Apoptosis in Human Leukemic Cell Lines U937, K562 and HL-60 by *Litchi chinensis* Leaf extract Via Activation of Mitochondria. Mediated Caspase Cascades. *Open Leukemia J.* 1(1):1-14.
172. Guevara AP, Vargas C (1996). Anti-inflammatory and antitumor activities of seed extracts of malunggay, *Moringa oleifera* L. (Moringaceae). *Philippine J Sci* 125(issue??): 175-84.

173. Khalafalla MM, Abdellatef E, Dafalla HM, Nassrallah AA, Aboul-Enein KM, Lightfoot DA, Ei-Deeb FE, Ei-Shemy HA (2010). Active principle form *Moringa oleifera* Lam leaves effective against wro leukemias and a hepatocarcinoma. *African J Biotech.* 9(49): 8467-71.
174. Khalafalla MM, Abdellatef E, Dafalla HM, Nassrallah AA, Aboul-Enein KM, Hanu A, El-Shemy HA, Abdellatef E (2011). Dedifferentiation of leaf explants and antileukemia activity of an ethanolic extract of cell cultures of *Moringa oleifera*. *African J Biotech.* 10(14):2746-50.
175. Lee SJ, ChoYH, Park K, Kim EJ, Kang BS, Jung KH, Kim CH, Kim Ch, Kim VVj, Moon SK (2009). Inhibitory effects of the aqueous extract of *Magnolia officinalis* on the responses of human urinary bladder cancer 5637 cells in vitro and mouse urinary bladder tumors induced by N-Butyl-N-(4-hydroxybutyl) nitrosamine in vivo. *Phytother Res.* 23(1):20-7.
176. Romeilah RM (2009). Anticancer and antioxidant activities of *Matricaria chamomilla L.* and *Marjorana hortensis* essential oils. *Res J Medicine Medical Sci.* 4(2):332-9.
177. Sharma V. A (2011). Polyphenolic compound rottlerin demonstrates significant in vitro cytotoxicity against human cancer cell lines: isolation and characterization from the fruits of *Mallotus philippinensis* *J Plant Biochem Biotechnol.* 20(2): 190-5.
178. Yanga MY, Changa YC, Chanb KC, Lee YJ, Wanga CJ (2011). Flavonoid-enriched extracts form *Nelumbo nucifera* Leaves inhibits proliferation of breast cancer *in vitro* from the fruits of *Mallotus Philippinensis*. *J Plant Biochem Biotechnol.* 20(2):190-5.
179. Zhang X, Liu Z, Xu B, Sun Z, Gong Y, Neferine CS (2012). An alkaloid ingredient in lotus seed embryo inhibits proliferation of human osteosarcoma cells by promoting p38 MAPK-mediated p21 stabilization. *Eur J Pharmacol.* 677(1): 47-54.
180. Kohli KR, Nipanikar SU, Kadhane KP (2010). A Comprehensive Review on *Trivrit [Operculina Turpethum Syn. Lpomoea Turpethum]*. *Int J Pharma Bio Sci.* 1(4): 443-52.
181. Fan HW, Min H, Li YU, Yan LI, Ying-bin LI (2009). Study on the Anti-tumor effect of *Oldenlandia diffusa* on HL60 and B16BL6 cell line *in vitro*. *Chinese J Hospital Pharm.* 20.
182. Wu HW, Chi Shing, Tai W, Linag ZT, Zhao ZZ, Hsiao ZZ, Hsiao WL (2009). Oleanolic acid isolated from *Oldenlandia diffusa* exhibits a unique growth inhibitory effect against ras-transformed fibroblasts. *Life Sci.* 85(3):113-21.
183. Islam MS, Akhtar MM, Rahman MM, Rahman MA, Sarker KK, Alam MF (2009). Anti-tumor and phytotoxic Activities of leaf methanol extract of *Oldenlandia diffusa* (Willd.) Roxb. *Global J Pahrmacol.* 3(2):99-106.

184. Raveendran VV, Vijayan FP, padikkala J (2012). Antitumor Activities of an ANthraquinone Fraction Isolated from in Vitro Cultures of *Ophiorrhiza rugosa* vsr decumbens. *Integr Cancer ther*, 12(2): 120-8.
185. Patanayak P, Behera P, Das D, Panda SK (2010). *Ocimum sanctum* Linn. A reservoir plant for therapeutic applications: An overview. *Pharmacogn Rev*. 4(7): 95-105.
186. Rastogi S, Shukla Y, Paul BN, Chowdhruai DK, Khanna SK, Das M (2007). Protective effect of *Ocimum sanctum* on 3-methylcholanthrene, 7,12-dimethylbenz(a) anthracene and aflatoxin B1 induced skin tumorigenesis in mice. *Toxicol Appl Pharmacol*. 224(3): 228-40.
187. Thompson MD, Thompson HJ, Brick MA, McGinley JN, Jiang W, Zhu Z, Wolfe P (2008). Mechanisms Associated with Dose-Dependent inhibition of Rat Mammary Carcinogenesis by Dry Bean (*Phaseolus vulgaris*, L.). *J Nutr*. 138(11): 2091-7.
188. Ogunwande IA, Walker TM, Bansal A, Setzer WN, Essien EE (2010). Essential oil constituents and biological activities of *Peristrophe bicalyculata* and *Borreria verticillata*. *Nat Prod Commun*. 5(11): 1815-8.
189. Sivajothi V, Leelaprakash G (2010). Cytotoxicity Screening of Alcoholic Extract of the Whole Plant of *Phyllanthus rheedii*. *Res J Chem Env*. 14(3): 81-2.
190. Wu HY, Lin TK, Kuo HM, Huang YL, Liou CW, Wang PW, Chuang JH, Huang ST (2012). *Phyllanthus ruinaria* induces Apoptosis in Human Osteosarcoma 143B cells via Activation of Fas/FasL- and Mitochondria-Mediated Pathways. *eCAM*. 2012: Article ID 925824, 13 pages.
191. Gao LL, Li FR, Jiao P, Yang MF, Zhou XJ, Si YH, Jiang WJ, Zheng TT (2011). *Parischinensis* Dioscin induces G2/M cell cycle arrest and apoptosis in human gastric cancer SGC-7901 cells. *World J Gastroenterol*. 17(39): 4389-95.
192. Sun J, Liu FR, Wei J, Qian XP, Yu LX, Guo RH, Shen H, Wang TS (2011). The extract of *Paris polyphylla* exerts apoptotic induction and synergic antiproliferative effect with anticancer drugs in SMMC-7721 human liver cancer cells. *Biomed Prev Nutr*. 1(3): 186-94.
193. Yun H, Lijan C, Wenhong Z, Yuhong D, yongli W, Qiang W, Ding Z (2007). Separation And identification of Steroidal Compounds with Cytotoxic Activity Against Human gastric Cancer Cell Lines In Vitro From The Rhizomes of *Paris polyphylla* VAR. *Chinensis*. *Chem Nat Comp*. 43(6): 672-7.

194. Lee KJ, Hwang SJ, Choi JH, Jeong HG (2008). Saponins derived from the roots of *Platycodon grandiflorum* inhibit HT-1080 cell invasion and MMPs activities: regulation of NF-KB activation via ROS signal pathway. *Cancer Lett.* 268(2): 233-43.
195. Shin DY, Kim GY, LiW, Choi BT, kim ND, Kang HS,choi YH. Implication of intracellular ROS formation , caspase-3 activation and Egr-1 induction in platycodon D-induced apoptosis of U937 human leukemia cells, *Biomed Pharmacother.* 63(2): 86-94.
196. Checker R, Shurma D, Snadur SK, Subrahmanyam G, Krishnan S, poduval TB, Sainis KB (2010). Plumbagin inhibits proliferative and inflammatory responses of T cells independent of ROS generation but by modulating intracellular thiols. *Cell Biochem.* 110(5): 1082-93.
197. Chen CA, Chang HH, Kao CY, Tsai TH, Chen YJ (2009). Plumbagin, Isolated from *Plumbago zeylanica*, induces Cell Death through Apoptosis in Human Pancreatic Cancer Cells. *Pancreatology* 9(6): 797-809.
198. Powolny AA, singh SV, Plumbagin-induced Apoptosis in Human Prostate Cancer Cells in Associated with Modulation of Cellular Redox Status and Generation of Reactive Oxygen Species, *Pharm Res*, 25(9): 2171-80.
199. Liu Y, Yadev VR, Aggarwall BB, Nair MG (2010). Inhibitory effects of black pepper (*Piper nigrum* ) extracts and compounds on human tumor cell proliferation, cyclooxygenase enzymes, lipid peroxidation and unclear transcription factor-kappa-B. *nat Prod Commun* 5(8): 1253-7.
200. Reshmi SK, Sathya E, Sugnay PD (2010). Lsolation of piperdine from *Piper nigrum* and ikjts antiproliferative activity. *J Pharm Res.* 3(10): 2502-7.
201. Lin ES. Chou JH, Kuo PL, Huang YC(2010). Antioxidant and antiproliferative activities of methanolic extracts of *Perilla frutescens*. *J Med Plants Res.* 4(6): 477-83.
202. Vinod PV (2012). Guruvayoorappan C. Evaluation of immunostimulant activity and chemoprotective effect of mabgrove *Rhizophora apiculata* against cyclophosphamide induced toxicity in BALB/c mice. *Immunopharmacol immunotoxicol.* 34(4): 608-15.
203. Patel P, Nagar A, Patel R, Rathod D, Liu L, Yang L (2011). *In vitro* Anticancer Activity of *Rubia Cordifolia* aganist Hela and Hep2 Cell Lines. *Int J Pharm Pharm Sci.* 3(2): 70-1.
204. Long G, Wang G, Ye L, Lin B, Wei D, Liu L, Yang L (2009). Important Role of TNF- $\alpha$  in Inhibitroy Effects of Tadix Sophorae falvescentis Extract on Vascular Restenosis in a Rat Carotid Model of Ballon Dilatation injury. *Planta Medica.* 75(12): 1293-9.

205. Parmar J, Sharma P, Verma P, Sharma P, Goyal PK (2011). Anti-tumor and Anti-oxidative Activity of *Rosmarinus officinalis* in 7, 12 Dimethyl Benz (a) Anthracene induced Skin Carcinogenesis in mice. *Am J Biomed Sci.* 3(3): 199-209.
206. Tai J, Cheung S, Wu M, Hasman D (2012). Antiproliferation effect of Rosemary (*Rosmarinu officinalis*) on human ovarian cancer cells in vitro. *phytomedicine* 19(5):436-43.
207. Tsai CW, Lin CY, Lin HH, Chen JH (2011). Carnosic Acid, a Rosemary Phenolic compound, induces Apoptosis through Reactive Oxygen Species-Mediated p38 Activation in Human Neuroblastoma IMR-32 Cells. *Neurochem Res.* 36(12): 2442-51.
208. Kim EJ, Lim SS, park SY, Shin HK, Kim JS, Park JH (2008). Apoptosis of DU 145 human prostate cancer cells induced by dehydrocostus lactone isolated from the root of *Saussurea lappa*. *Food Chem Toxicol.* 46(12): 3651-8.
209. Robinson A, Kumar Tv, Sreedhar E, Naidu VG, Krishna SR, Babu KS, Srinivas PV, Rao JM (2008). A New sesquiterpene lactone from the roots of *Saussurea lappa*: Structure-anticancer activity study. *Bioorg Med Chem Lett.* 18(14): 4015-7.
210. Anand HK, Goyal D (2011). Extractoin of tylphorine from in vitro raised Manju plants of *Thlophora indica*. *J Med Plnats Res.* 5(5):729-34.
211. Sigstedt SC, Hooten CJ, Callewaert MC, Jenkins AR, Romero AE, Pullin MJ, Kornienko A, Lowery TK, Slambrouck SV, Steelant WF (2008). Evaluation of Aqueous Extracts of *Taraxacum Officinale* on Growth and incasion of Breast and Prostate Cancer Cells. *Int J Oncol.* 32(5): 1085-90.
212. Shinozaki Y, Fukamiya N, Fukushima M, Okano M, Nehira T, Tagahara K, Zhang SX, Zhang DC, Lee KH (2002). Dantaxusins C and D, Two Novel Taxoids from *Taxus yunnanensis*. *J Nat Prod.* 65(3):371-4.
213. Queiroz ML, Valadares MC, Torello CO, Ramos AL, Oliveira AB, Rocha FD, Arruda VA, Accorci WR (2008). Comparative studies of the effects of *Tabenuia avellanadae* bark extract and  $\beta$ -lapachone on the hematopoietic response of tumorbearing mice. *J Ethnopharmacol.* 117(2): 228-35.
214. Yamashita M, Kaneko M, Tokuda H, Nishimura K, Kumeda Y, lida A (2009). Synthesis and evaluation of bioactive naphthoquinones from the Brazilian medicinal plant, *Tabebuia avellanadae*. *Bioorg Med Chem.* 17(17): 6286-91.
215. He X, Liu RH(2006). Cranberry phytochemicals: Isolation, structure elucidation and their antiproliferative and and antioxidant activities. *J Agric Food chem..* 54(19):7069-74.

216. Kondo M, Mackinnon SL, Craft CC, Matchett MD, Hurta RA, Neto CC (2011). Ursolic acid and its esters: occurrence in cranberries and other *Vaccinium fruit* and effects on matrix metalloproteinase activity in DU145 prostate tumor cells. *J Sci Food Agric.* 91(5): 789-96.
217. Mueller- Harvey I (1999). Tannins: their nature and biological significance. In : Secondary plants products. Antinutritional and beneficial actions in animal feeding Cayill JC and Mueller-Harvey I, eds. Nottingham Univ Press (UK), 17-70.
218. Mueller – Harvey I, McAllan AB (1992). Tannins. Their biochemistry and nutritional properties. In : Advances in Plant cell biochemistry and biotechnology, Vol. 1 Morrison IM, Ed, JAI Press Ltd, London(UK), 151-217.
219. Rao RVk, Ali N, Reddy MN (1978). Occurrence of both saponins and alkaloid lycorine in *Curculigo orchoides*. *Indian Journal Pharma Science*, 40: 104-105.
220. Mishra SN (1989). Analytical methods for analysis of total alkaloids in root of *Withania* spp. Proc. All India Workshop on M&AP, Faizabad, 492-95.
221. Krishan R, Chandravadana MV, Ramachander PR, Bharath kumar H (1983). Inter-relationships between growth and alkaloid production in *Catharanthus roseus* G. Don . *Herba Hungarica*, 22: 47-54.
222. Molyneuz RJ, Nash RJ, Asano N (1996). Alkaloids: Chemical and Biological Perspectives, Vol. 11, Pelletier SW, Ed. Pergamon, Oxford, 303.
223. Wink m, Schmeller T, Latz-Briining B (1998). Modes of action of allelochemical alkaloids: Interaction with neuroreceptors , DNA and other molecular targets. *Journal of chemical Ecology*, 24: 1888-1937.
224. Elbein AD, Molyneux RJ (1999). *Comprehensive Natural Products Chemistry*, Vol. 3, Barton D and Nakanishi K, ed. Amsterdam, 129.
225. Harborne JB, Tomas-Barberan FA (1991). *Ecological Chemistry and Biochemistry of plant Terpenoids*, Clarendon, Oxford.
226. Langenheim JH (1994). Higher plant terpenoids: A phyto-centric overview of their ecological roles. *Journal of Chemical Ecology*, 20: 1223-1280.
227. McCaskill D, Croteau R (1998). Some caveats for bioengineering terpenoid metabolism in plants. *Trends Biotechnology*, 16: 349-355.
228. Degenhardt J, Gershenzon J, Baldwin IT, Kessler A (2003). Attracting friends to feast on foes: Engineering terpene emission to make crop plants more attractive to herbivore enemies. *Current Opinion biotechnology*, 14: 169-176.

229. Dudareva N, Pichersky E, Gershenzon J (2004). Biochemistry of plant volatiles. *Plant physiology* 135: 1893-1902.
230. Bohlmann J, Meyer-Gauen G, Croteau R (1998). Plant terpenoid synthases: Molecule biology and phylogenetic analysis. *Proc Natl Acad Sci USA*, 95: 4126-4133.
231. Lasztity R, Hidvegi M, Bata A (1998). Saponins in food. *Food review International*, 14: 371-390.
232. Lacaille-Dubois MA, Wagner H(2000). Bioactive saponins from plants:an update. In studies in Natural Products Chemistry; Atta-Ur- rahman, ed. Elsevier Science. Amsterdam. 21: 633-687.
233. Morrissey JP, Osbourn AE (1999). Fungal Resistance to plant antibiotics as a mechanism of pathogenesis. *Micro biological and Molecular Biological Reviews*, 63: 708-724.
234. Takechi M, Matsunami S, Nishizawa J, Uno C, Tanaka Y (1999). Haemolytic and antifungal activities of saponins or anti-ATPase and antivital activities of cardiac glycosides. *Planta Medica*, 65: 585-586.
235. Traore F, Faure R, Ollivier E, Gasquet M, Azas N, Debrauwer L, Keita A, Timon-David P, Balansard G (2000). Structure and antiprotozoal activitu of triterpenoid saponins from *Glinus oppositifolius*. *Planta Medica*, 66: 368-371.
236. George F, Zohar K, Harinder PS, Makkar, Klaus B (2002). The biological action of saponins in animal systems: a review. *British journal of Nutrition*, 88: 587-605.
237. CHEN, R.-Q. ET AL. (1982). Zhi Mu sapogenin is a powerful inhibitor of  $\text{Na}^+$ ,  $\text{K}^+$  ATPase. *Acta biochimica et biophysica Sinica*, 14: 159-164 .
238. DUGGAN, D. E.& NOLL, R. M(1965). Effects of ethacrynic acid and cardiac glycosides upon a membrane adenosinetriphosphatase of renal cortex. *Archives of biochemistry and biophysics*, 109:388-396.
239. PEZZUTO, J. M. ET AL (1978). Metabolism of benzo(a) pyrebe and (-)-trans -7, 8-dihydroxy-7, 8-diyhydrobenzo(a)pyrene by rat liver nuclei and microsomes. *Cancer research* 38: 1241-1245.
240. MERRIMAN , R. L. & BERTRAM, J. S. Reversible inhibition by retinoids of 3-methylcholanthrene-induced neoplastic transformation in C3H/10T ½ clone 8 cells. *Cancer research*, 39: 1661-1666.

241. NESLOW, S. & HEIDELBERGER, C (1976). The effect of modifiers of microsomal enzymes of chemical oncogenesis in cultures of C3H mouse cell lines. *Cancer research*, 36: 1801-1808.
242. GERAN, R. I. ET AL (1972). Test systems. *Cancer chemotherapy, reports, part 3*, 3: 1-102 .
243. SARRIFF, A. M. ET AL. (1979). Establishment of photo-affinity label derivatives of fluorine as probes in studies of chemical carcinogens in mammalian cell cultures. *Cancer research*, 39: 3903-3908.
244. TAPPEL, L.A. (1972). Vitamin E and free radical peroxidation of lipids. *Annals of the New York Academy of Sciences*, 12-28.
245. STACPOOLE, P. W. ET AL (1982). Stimulation of rat liver 3-hydroxy-3-methylglutaryl-coenzyme A reductase activity by *o,p'* DDD. *Biochemical Pharmacology*, 31: 857-860.
246. SALMON, S. E. ET AL (1978). Quantitation of differential sensitivity of human-tumor stem cells to anticancer drugs. *New England journal of medicine*, 298: 1321-1327.
247. VON HOFF, D. D. ET AL. (1981). Association between human tumor colony-forming assay results and response of an individual patient's tumor to chemotherapy. *American journal of medicine*, 70:1027-1032.
248. MEISNER, J. ET AL (1981). Phagodererreny induced by leaves and leaf extracts of *Catharanthus rosesu* in the larva of *Spodoptera littoralis*. *Journal of economic entomology*, 74: 131-135.
249. MUNAKATA, K. & WADA, K(1981). Insect antifeedants in plants. In: S. Natori *et al* . ed., *Advances in natural products chemistry- extraction and isolation of biologically active compounds* ,New York Wiley, pp. 240-248.
250. SU, H.C. F. ET AL (1982). Isolation , purification and characterization of insect repellants from *Curcuma longa* L. *journal of agricultural and food chemistry*. 30: 290-292.
251. JACOBSON, M *et al* (1950). Survey of plants for insecticide activity, *Lloydia*. 13:89-162.
252. Molluscicide screening and evaluation. *Bulletin of the world Health Organization*, 33: 567-581.
253. ROBINSON, D.S. ET AL (1968). Effects of drugs on human blood platelet and plasma amine oxidase acticity *in vitro and in vivo*. *Biochemical pharmacology*. 17 109-119.

254. AMES, B. N. ET AL (1975). Methods for detecting carcinogens and mutagens with the *Salmonella / mammalian* –microsome mutagenicity test. *Mutation research* 31: 347-364.
255. O'NEILL, J. P. ET AL (1977). A quantitative assay of mutation induction at the hypoxanthine-guanine phosphoribosyl transferase locus i Chinese hamster ovary cells (CHO/HGPRT system): development and definition of the system. *Mutation research*, 45: 91-101.
256. UNGSURUNSIE, M, et al (1982). Mutagenicity screening of popular thai spices. *Food and cosmetic toxicology*, 20:527-530.
257. PEZZUTO, J . M. ET AL (1981). Metabolic activation of 3-amino-1-methyl-5H-pyrido (4,3-b) indole and several structurally related mutagens. *Biochemistry*, 20: 298-305.
258. KUCHLER, R. J. Biochemical methods in cell culture and virology. Stroudsburg, PA, Dowden, Hutchinson & Ross.
259. SEDWICK, W. D. ET AL (1974). The DNA polymerases of KB cells. *Methods in enzymology*, 29: 89-102.'
260. NIKAIDO, T. ET AL (inhibitors of cyclic AMP phosphodiesterase in medicinal plants, *Planta medica*, 43: 18-23.
261. MAKHEJA, A. N. ET AL (1979). Effects of onions (*Allium cepa*) on platelet aggregation and thromboxane synthesis. *Prostaglandins and medicine*. 2: 413-424.
262. OHUCHI, K. ET AL (1981). Glycyrrhizin inhibits prostaglandin- E<sub>2</sub> production by activated peritoneal macrophages from rats. *Prostaglandins and medicine*. 7: 457-463.
263. SAEED, S. A. ET AL (1981). Inhibitor (s) of prostaglandin biosynthesis in extracts of oat (*avena sativa* ) seeds. *Biochemical Society transactions*, 9: 444.
264. LEWASZ, J. ET AL (1981). Electrophoretic method for the determination of molecular forms of trypsin inhibitors of potato tubers. *Analytical biochemistry*, 115: 27-29.
265. NORIOKA, S. ET AL (1982). Purification and characterization of protease inhibitors frompeanuts ( *Arachis hupogaea*). *Journal of biochemistry*. 91: 1427-1434.
266. OHKOSHI, M. (1981). Inhibition of growth of 3-methylcholanthrene-induced mouse skin tumor by protease inhibitor (*N, N-dimethylcarbamoylmethyl-4-(4-guanidinobenzoyloxy)-phenylacetate*) methanesulfate. *Gann*, 72: 959-965.
267. PEZZUTO, J. M. & HECHT , S. M. Amino acid substitutions in protein biosynthesis. Poly (A)- directed polyphenylalanine synthesis. *Journal of biological chemistry*, 255: 865-869.

268. WOODWARD, W. R. ET AL (1974). Protein synthesis with rabbit reticulocyte preparations. *Methods in enzymology*, 30: 724-731.
269. PERRY, P. & EVANS, H. J. (1975). Cytological detection of mutagen-carcinogen exposure by dopa formed from 1—<sup>14</sup>C-L-tyrosine. *Analytical biochemistry*, 43: 588-600.
270. WAYMIRE, J. C. & SAN, R. H. C. (1971). Use of unscheduled DNA synthesis in freshly isolated human intestinal mucosal cells for carcinogen detection. *Cancer research*, 40: 3155-3157.
271. FREEDMAN, H. J. & SAN, R. H. C. (1980). Use fo unschedulaed DNA synthesis in freshly isolated human intestinal mucosal cells for carcinogen detection. *Cancer research*, 40: 3155-3157.
272. SIRICA, A. E. ETAL (1980). Use of primary cultures of adult rat hepatocytes on collagen gel-nylon mesh to evaluate carcinogen-induced unscheduled DNA synthesis. *Cancer research*, 40: 3259-3267.