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AN OVERVIEW : ANTI ULCER ACTIVITY WITH MEDICINAL PLANTS**Dr.S.Senthilkumar****Karur, Tamilnadu, India.****KEYWORDS:**

Anti ulcer, *Helicobacter Pylori*,
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ABSTRACT

Peptic ulcer disease is considered as one of the common disease in the world. Treatment of peptic ulcer with synthetic drugs such as proton pump inhibitors, H₂ receptor antagonists and other non-steroidal anti-inflammatory drugs has shown adverse effects, relapses, drug interactions. Medicinal plants containing active chemical constituents are useful in prevention and treatment of various diseases. An ulcer is disease of epithelial cells of body or organ which represents an open sore in the lining of epithelial cells or deep lesion in the specific region resulting into its degradation thus disrupting the normal physiology of the organ affected. The damage can be to the extent of bleeding, which is commonly known as bleeding sore. Traditional medicines or to point out clearly, the medicinal plants have wider range of therapeutic advantages in the proper management of diseases, as they have better pharmacological activity along with low incidence of side effects or the adverse effects. Medicinal plants also thus have huge market for themselves in our ultimate goal of providing healing touch to the disease person.

INTRODUCTION:

Peptic ulcer is a gastro intestinal disorder due to an imbalance between the aggressive factors like acid, pepsin, *Helicobacter pylori* and defensive factors like bicarbonate secretion, prostaglandins, gastric mucus, innate resistance of the mucosal cell factors. Normally peptic ulcer develops when aggressive factors overcome the defensive. The major factors that disrupt the equilibrium between aggressive factors and defensive factors are *Helicobacter pyloro*, acid-pepsin hyper secretion, non-steroidal anti-inflammatory drugs, sometimes idiopathic due to usage of cobacco, psychological stress, rapid gastric emptying and Zollinger-Ellisson syndrome where there is a high and uncontrollable production of acid also leads to ulcer formation. Sunthetic drugs such as proton pump inhibitors, H₂ receptors, cytoprotectants, demulcents, anti cholinergics, antacids and prostaglandin analogues are used for the treatment of ulceration but these drugs produce several side effects.

So herbal medicines are considered as better alternatives for the treatment of peptic ulcer for example, proton pump inhibitors (omeprazole, Lansoprazole) may cause gynaecomastia, loss of libido. Due to the cocurrence of many side effects by use of synthetic drugs as they have less or no side effects. As herbal medicines are considered as safe for the treatment of ulcers with lesser adverse effects, economical, effective, relatively less toxic, extensive research is carried out in search for potent antiulcer agents of plant origin.

Peptic ulcer disease represents a serious medical problem. Approximately 500,000 new cases are reported each year, with 5 million people affected in the United States alone. Interestingly, those at the highest risk of contracting peptic ulcer disease are those generations born around the middle of the 20th century. Ulcer disease has become a disease predominantly affecting the older population, with the peak incidence occurring between 55 and 65 years of age. In men, duodenal ulcers were more common than gastric ulcers; in women, the converse was found to be true. Thirty-five percent of patients diagnosed with gastric ulcers will suffer serious complications. Although mortality rates form peptic ulcer disease are low, the high prevalence and the result in pain, suffering, and expense are very costly.

Ulcer can develop in the esophagus, stomach or duodenum, at the margin of a gastroenterostomy, in the jejunum, in Zollinger-Ellison syndrome, and in association with a meckel's diverticulum containing ectopic gastric mucosa. Peptic ulcers disease is one several disorders of the upper gastrointestinal tract that is caused, at least partially, by gastric acid. Patients with peptic ulcer dis

may present with a range of symptoms, from mild abdominal discomfort to catastrophic perforation and bleeding.

SYMPTOMS:

More patients with peptic ulcer disease present with abdominal discomfort, pain or nausea. The pain is located in the epigastrium and usually does not radiate. However, these symptoms are neither sensitive nor specific. Pain radiating to the back may suggest that an ulcer has penetrated posteriorly, or the pain may be pancreatic in origin. Pain radiating to the right upper quadrant may suggest disease of the gallbladder or bile ducts.

Patients may describe the pain of peptic ulcer as burning or gnawing, or as hunger pains slowly building up for 1-2 hours, then gradually decreasing. Use of antacids may provide temporary relief. Classically, gastric ulcer pain is aggravated by meals, whereas the pain of duodenal ulcers is relieved by meals. Hence, patients with gastric ulcers tend to avoid food and present with weight loss, while those with duodenal ulcers do not lose weight. It is important to remember that although these patterns are typical, they are not pathognomonic. The nature of the presenting symptoms alone does not permit a clear differentiation between benign ulcers and gastric neoplasm.

TYPES OF ULCERS:**PEPTIC ULCER:**

Any ulcer that is exposed to pepsin is referred to as peptic ulcers. Peptic ulcers are found in the lining of stomach or duodenum. Pepsin is normally present along with hydrochloric acid in the stomach lining.

DUODENAL ULCER:

When a peptic ulcer is in duodenum, it is called a duodenal ulcer. This type of peptic ulcer develops in the first part of the small intestine. Some of the symptoms of a duodenal ulcer are interestingly quite opposite to those of gastric ulcers. Duodenal ulcers are the most common ulcers found in the western world.

GASTRIC ULCER:

When a peptic ulcer is in stomach, it is called a gastric ulcer. The symptoms of gastric ulcers are more specific than peptic ulcer symptoms.

ESOPHAGEAL ULCER:

This type of ulcer occurs in the lower end of your esophagus. Esophageal ulcers are often associated with a bad case of acid reflux, or GERD as it is commonly called (short for Gastro Esophageal Reflux Disease).

BLEEDING ULCER:

Internal bleeding is caused by a peptic ulcer which has been left untreated. When this happens, it is now referred to as a bleeding ulcer-this is the most dangerous type of ulcer. See your doctor immediately if you are showing symptoms.

REFRACTORY ULCER:

Refractory ulcers, are simply peptic ulcers that have not healed after at least 3 months of treatment.

STRESS ULCER:

Stress ulcers are a group of lesions found in the esophagus, stomach or duodenum. These are normally only found in critically ill or severely stressed patients.

DIAGNOSIS OF PEPTIC ULCER DISEASE:

Symptoms depend on ulcer location and patient age. Many, patients particularly elderly patients, have few or no symptoms. Pain is however the most common symptom, often localized to the epigastrium or mid-epigastrium and relieved by food or antacids. The pain is described as burning, gnawing, constant or annoying, or sometimes a sensation of hunger. The course is usually chronic and recurrent. Only about 50% of patients present with the characteristic pattern of symptoms.

Gastric ulcer symptoms often do not follow a consistent pattern (for example, eating sometimes exacerbates rather than relieves, pain). This is especially true for pyloric channel ulcers, which are often associated with symptoms of obstruction (for example bloating nausea and vomiting) caused by oedema and scarring. In general, however, in gastric ulcers, pain typically starts whenever the stomach is empty (usually approximately an hour after eating), and is generally relieved by antacids or food but aggravated by alcohol and caffeine. Weight loss and gastrointestinal bleeding occur more frequently with gastric ulcers. Patients can experience weight loss of 5kg to 10kg and although this could indicate carcinoma, especially in people over 40 years, on investigation a benign gastric ulcer is found most of the time.

Duodenal ulcers tend to produce more consistent pain. Pain is absent when the patient awakens but appears midmorning, is relieved by food, but recurs two to three hours after a meal. Pain that awakens a person at night, a few hours after falling asleep, is also common and is highly suggestive of duodenal ulcer. The pain then usually subsides by morning and is often relieved after eating. This is not commonly noticed in gastric ulceration. In neonates, perforation or haemorrhage may be the first manifestation of duodenal ulcer. Haemorrhage may also be the first recognized sign in later infancy and early childhood, although repeated vomiting or evidence of

abdominal pain may be a clue. Diagnosis of peptic ulcer is by patient history, and confirmed by endoscopy and testing for H pylori.

Carbon-13 urea breath tests detect active H pylori infection by testing for the enzymatic activity of bacterial urease. In the presence of urease produced by H pylori, labeled carbon dioxide is produced in the stomach, absorbed into the bloodstream, diffused into the lungs and exhaled.

Stool or faecal antigen testing identifies active H pylori infection by detecting the presence of H pylori antigens in stools.

Serology, which is immunoglobulin G(IgG) based, can be measured in serum, plasma or whole blood. It will, however, not distinguish between a previous or a current infection.

Biopsy-based urease tests, which are invasive and can only be done at gastroscopy or in the acute hospital setting. There are two methods for this test. In the CLO test (“Campylobacter-like organisms” test, the rapid urease test) a fragment of mucosal membrane is placed into a special jelly which undergoes a colour change in 10 to 20 minutes, or the specimen is sent for histology which may take up to 24 hours to obtain the result.

ENDOSCOPY:

An endoscope is a thin, flexible tube with a tiny camera at the end. The patient is given a mild sedative, and then the tube is passed through mouth in to the lining of the stomach to diagnose a peptic ulcer. Tiny samples of the tissue will be taken (biopsy), which are examined under a microscope.

If a diagnostic imaging test reveals an ulcer, the patient will most likely have a test to see if H pylori bacteria are present.

CAUSES OF PEPTIC ULCER DISEASE:

PROTECTIVE VS.HOSTILE FACTORS:

“No gastric acid, no peptic ulcer” is a misconception. Excessive gastric acid secretion is only one factor in the pathogenesis of peptic ulcer disease. Decreased mucosal defense against acid is another cause. The integrity of the upper gastrointestinal tract is dependent upon the balance between “hostile” factors such as gastric acid, H. pylori, NSAIDs and pepsin, and “protective” factors such as prostaglandins, mucus, bicarbonate, and blood flow to mucosa affecting gastrointestinal mucosa injury to gastric and duodenal mucosa develops when deleterious effects of gastric acid overwhelm the defensive properties of the mucosa. Inhibition of endogenous prostaglandin synthesis leads to a decrease in epithelial mucus, bicarbonate secretion, mucosal blood flow, epithelial proliferation, and mucosal resistance to injury. Lower mucosal resistance

increases the incidence of injury by endogenous factors such as acid, pepsin, and bile salts as well as exogenous factors such as NSAIDs ethanol and other noxious agents.

HELICOBACTER PYLORI:

H. pylori is the etiologic factor in most patients with ulcer disease and may predispose individuals to the development of gastric carcinoma. H. pylori colonizes in the human stomach. The method of H. pylori transmission is unclear, but seems to be person-to-person spread via a fecal-oral route. The prevalence of H. pylori in adults appears to be inversely related to the socioeconomic status. It is also thought that water is a reservoir for transmission of H. pylori.

NONSTEROIDAL ANTI-INFLAMMATORY DRUGS(NSAIDS):

A small but important percentage of patients have adverse gastrointestinal events associated with NSAID use that results in substantial morbidity and mortality. Risk factors for the development of NSAID-associated gastric and duodenal ulcers include advanced age, history of previous ulcer disease, concomitant use of corticosteroids and anticoagulants, higher doses of NSAIDs, and serious systemic disorders. The concept of gastroduodenal mucosal injury has evolved from the notion of topical injury to concepts that involve multiple mechanisms.

NSAIDs initiate mucosal injury topically by their acidic properties. By diminishing the hydrophobicity of gastric mucus, endogenous gastric acid and pepsin may injure surface epithelium. Systemic effects of NSAIDs appear to play a predominant role through the decreased synthesis of mucosal prostaglandins. The precursor of prostaglandins, arachidonic acid, is catalyzed by the cyclo-oxygenase isoenzymes, cyclo-oxygenase-1 and cyclo-oxygenase-2. The gene for cyclo-oxygenase-1. The housekeeping enzyme, maintains the homeostasis of organs. Cyclo-oxygenase-2. the inflammatory enzyme, is inducible. Although NSAIDs can inhibit both pathways, only the gene for cyclo-oxygenase-2 contains corticosteroid-responsive repressor element. Literature suggests that the anti-inflammatory properties of NSAIDs are mediated through inhibition of cyclo-oxygenase-2, and adverse effects, such as gastric and duodenal ulceration, occur as a result of effects on the constitutively expressed cyclo-oxygenase-1. H. pylori is prevalent among 22-63% of patients taking NSAIDs. Most studies do not show a significant difference in H. pylori prevalence between NSAIDs users and nonusers. Gastritis in patients on NSAID therapy appears to be related to underlying H. pylori rather than drug use. The lower incidence of H. pylori among patients with gastric ulcers than those with duodenal ulcers is presumably the result of NSAID use. NSAIDs are more likely to cause gastric than duodenal ulcers. NSAIDs appear to

cause ulcers by a mechanism independent of H.pylori based on the inhibition of prostaglandin synthesis.

GASTRINOMA (ZOLLINGER-ELLISON SYNDROME):

The classic triad to Zollinger-Ellison syndrome involves peptic ulcers in unusual locations (i.e., the jejunum), massive gastric acid hypersecretion and a gastrin-producing islet cell tumor of the pancreas (gastrinoma). Gastrinoma in the pancreas appears in approximately 50% of patients. Another 20% patients have it in the duodenum and others have it in the stomach, peripancreatic lymph nodes, liver, ovary, or small-bowel mesentery.

Zollinger-Ellison syndrome accounts for only 0.1% of all duodenal ulcer disease. One fourth of patients have this syndrome as part of the multiple neoplasia syndrome type I (MEN I).

Patients with gastrinoma may have intractable ulcer disease. Because gastrin is trophic to the gastric mucosa, endoscopy or x-ray may demonstrate hypertrophy of the gastric rugae. Patients may also experience diarrhea (including steatorrhea from acid inactivation of lipase) and gastroesophageal reflux. These symptoms are episodic in 75% of patients.

HYPERCALCEMIA:

Hypercalcemia has a direct bearing on the gastric acid hypersecretory state found in patients with Zollinger-Ellison syndrome and MEN I. Intravenous calcium infusion in normal volunteers induces gastric acid hypersecretion. Additionally, calcium has been demonstrated in vivo and in vitro to stimulate gastrin release directly from gastrinomas. Resolution of hypercalcemia (by parathyroidectomy) reduces the basal acid output and serum gastrin concentration in fasting gastrinoma patients and those with MEN I, suggesting that resolution of hypercalcemia plays an important role in the therapy of this subgroup of patients.

GENETIC FACTORS:

Genetic factors play a role in the pathogenesis of ulcer disease. The lifetime prevalence of developing ulcer disease in first-degree relatives of ulcer patients is about three times greater than the general population. Approximately 20-50% of duodenal ulcer patients report a positive family history; gastric ulcer patients also report clusters of family members who are likewise affected.

SMOKING:

The literature reveals a strong positive correlation between cigarette smoking and the incidence of ulcer disease, mortality, complications, recurrences and delay in healing rates. Smokers are about two times more likely to develop ulcer disease than nonsmokers. Cigarette smoking and H.pylori are co-factors for the formation of peptic ulcer disease. There is a strong association between H.

pylori infection and cigarette smoking in patients with and without peptic ulcers. Cigarette smoking may increase susceptibility, diminish the gastric mucosal defensive factors, or may provide a more favorable milieu for H. pylori infection.

STRESS:

Numerous studies have revealed conflicting conclusions regarding the role of psychological factors in the pathogenesis and natural history of peptic ulcer disease. The role of psychological factors is far from established. Acute stress-results in increases in pulse rate, blood pressure and anxiety, but only in those patients with duodenal ulcers did acute stress actually result in significant increases in basal acid secretion. There is no clearly established “clear –type “ personality. Ulcer patients typically exhibit the same psychological makeup as the general population, but they appear to perceive greater degree of stress. In addition, there is no evidence that distinct occupational factors influence the incidence of ulcer disease.

ALCOHOL AND DIET:

Although alcohol has been shown to induce damage to the gastric mucosa in animals, it seems to be related to the absolute ethanol administered (200 proof). Pure ethanol is lipid soluble and results in frank, acute mucosal damage. Because most humans do not drink absolute ethanol, it is unlikely there is mucosal injury at ethanol concentrations of less than 10% (20 proof). Ethanol at low concentrations (5%) may modestly stimulate gastric acid secretions; higher concentrations diminish acid secretion. Though physiologically interesting, this has no direct link to ulcerogenesis or therapy.

Some types of food and beverages are reported to cause dyspepsia. There is no convincing evidence that indicates any specific diet causes ulcer disease. Epidemiologic studies have failed to reveal a correlation between caffeinated, decaffeinated, or cola-type beverages, beer, or milk with an increased risk of ulcer disease. Dietary alteration, other than avoidance of pain-causing foods, is unnecessary in ulcer patients.

THE CHANGING SPECTRUM OF THERAPY FOR ACTIVE PEPTIC ULCER DISEASE:

Conventional therapy for the prevention of peptic ulcers: Cimetidine and later ranitidine revolutionized the treatment of peptic ulcers, with H₂ receptor antagonists being the most widely used and effective novel drugs over the past decade. However, relapse ulceration following cessation of treatment with such agents is a frequent clinical observation. It is against this background that we have to deal with the current status and future advances in drug development for the therapy of gastroduodenal ulceration.

H₂ RECEPTOR ANTAGONISTS:

H₂ receptor antagonists are capable of reducing over 90% of basal, food stimulated, and nocturnal secretion of gastric acid stimulated by histamine, gastrin, cholinomimetic drugs and vagal stimulation. Histamine antagonists prevent occurrence of stress induced ulcers. However, their use in combination with antacids may be preferred. In addition, they are important in the medical management of Zollinger Ellison Syndrome and gastric hypersecretory states seen in systemic mastocytosis. As described earlier, recurrence of ulcer after healing is a frequent complicant of therapy with H₂-receptor antagonists, and therefore long term treatment is required. H₂-receptor antagonists are thus remarkable but not perfect drugs. These drugs include mainly cimetidine, ranitidine, famotidine, roxatidine and nizatidine. Saltidine, mifentidine, TZU-0460, CM-57755 etc. are also under investigation and have shown better antiulcer activity.

Ranitidine Bismuth Citrate: Ranitidine bismuth citrate (RBC) is the new anti-ulcer drug developed by Glaxo Laboratories (U.K.), which when combined with clarithromycin can eradicate *H.pylori* in 94% of patients. This has been confirmed in a clinical trial on 232 patients with duodenal ulcers. RBC was given 400 mg twice daily for 28 days and clarithromycin for times a day for first 14 days.

PROSTAGLANDINS:

In 1979, Robert recognised that PGs inhibits gastric acid secretion and protect against experimental ulcers caused by NSAIDs, diet and life styles(eg. alcohol, smoking and stress). Misoprostol (Cytotec) is a synthetic prostaglandin E analog with acid reducing and cytoprotective properties. Prostaglandins enhance mechanisms thought to be involved in mucosal defense of the chronic peptic ulcer (e.g., the secretion of mucus, output of bicarbonate, and blood flow). It is indicated for the prevention of NSAID-induced gastric ulceration. Short term co-administration of enprostil lowered the serum gastrin levels in patients on long term treatment with omeprazole. Misoprostol does not prevent duodenal ulcer. It is contraindicated in pregnancy because of its abortifacient property and requires special precautions if prescribed in pregnancy because of its abortifacient property and requires special precautions if prescribed to women of child bearing potential. The main side effect is diarrhoea in 6 to 30% of users. The synthetic PGs currently available in market are misoprostol, enprostil, rioprostil, arbaprostil and trimoprostil like compounds. Several other compounds like nocloprost, enisoprost, mexiprost, nileprost, rosaprostil etc. are undergoing clinical trials.

H⁺ K⁺-ATPASE INHIBITORS:

Blocked of the gastric proton pump constitutes a more direct mechanism for acid secretion inhibition compared to blockade of histamine and cholinergic receptors. Omeprazole is not the active inhibitor of H⁺K⁺ ATPase enzyme but is reversibly transformed in acidic media to the sulphenamide which can react with thiols to form disulfides, thus representing a model for the covalently linked enzyme-drug complex. Omeprazole has been shown to inhibit the growth of *H. pylori*. Recently, lansoprazole has been introduced in the markets of United States of America. NC-1300, RO 18-5362, B831-56 are series of fluorinated benzimidazoles and are potent and long acting inhibitors of acid secretion in animals and have shown mechanism similar to that of omeprazole.

MUSCARINIC RECEPTOR ANTAGONISTS:

Pirenzepine, a selective muscarinic M-1 receptor antagonists, reduces basal and stimulated acid secretion, in animals and man. Its efficacy in duodenal ulcer is equivalent to cimetidine. It has more cytoprotective effect than of histamine receptor antagonists against gastric mucosal lesions induced by ethanol, HCL, NaOH and taurocholate. Telenzepine is 4-10 times more potent than pirenzepine, as an inhibitor of acid secretion in rats and dogs.

MUCOSAL COATING AGENTS:

Sucralfate: Sucralfate is a sulfated disaccharide-basic aluminium sulfate complex. It forms an adherent coating with proteinaceous material at ulcerated mucosal sites. When pH is low, there is extensive polymerization and crosslinking of sucralfate. The coating provides barrier to hydrogen ion diffusion, reduces peptic activity and adsorbs bile salts. Further, sucralfate can bind to both epidermal growth factor (EGF), which also enhance ulcer healing. Recently, sucralfate has also been reported to suppress the associated *H. pylori* infection.

BISMUTH COMPOUNDS:

Bismuth subcitrate, formerly called tripotassium dicitrate bismuthate (TDB) is the most recent of the bismuth salts to be tested and found effective clinically. The substance is a colloidal suspension. When the pH is above 3.5 to 4.0; it forms a white precipitate in gastric acid. Bismuth subcitrate has a strong affinity for mucosal glycoproteins, especially in the necrotic tissue in ulcer craters. Ulcer craters become preferentially and visibly coated with a white layer of polymer glycoprotein complex, which is only slowly permeated by H₃O⁺, such that the layer constitutes a diffusion barrier to gastric acid. Bismuth salts also have some antimicrobial activity against

H.pylori infection. However, the chronic use of other bismuth salts has caused encephalopathy and osteodystrophy .

CARBENOXOLONE:

It is a synthetic derivative of glycyrrhizic acid (a constituent of liqyorice) which has been shown to be of value in promoting healing of peptic ulcers. The mechanism of action is not understood but, is believed to involve an effect on mucus, increasing its secretion and viscosity and thus protecting mucosa from attack by acid and pepsin. Its principal adverse effect is sodium retention which may lead to edema, hypertension and heart failure and this limits its use especially in the old people.

MISCELLANEOUS GROUP:

The anti-ulcer activity of calcium channel blockers namely verapamil, nifedipine and diltiazem against experimental ulcers has been established. Proglumide, a cholecystokinin and gastrin receptor antagonist is also found to possess antisecretory and antiulcer activity. The histidine decarboxylase inhibitors like(+)

PATHOPHYSIOLOGY OF ULCER:

Peptic ulcer is basically a lesion located at the level of the stomach, duodenum or esophagus. Ulcer tends to affect the entire gastrointestinal tract, starting from the lining of the mouth and ending with the rectal region. Peptic ulcer suggests the involvement of hydrochloric acid and pepsin in the development of the disorder. when gastric acid is produced in excess, the mucosal membrane that protects the stomach and internal organs from danger is damaged, enabling the bacteria *Helicobacter pylori* to penetrate the barrier and cause internal infections. Therefore, in the case of peptic ulcer, both gastric acid and bacteria are responsible for the development of the disorder. peptic ulcer located in the stomach is called gastric ulcer; peptic ulcer located at the level of the duodenum is called duodenal ulcer and peptic ulcer developed at the level of the esophagus is called esophagealulcer.

Despite extensive research, the etiology of peptic ulcer disease remains unclear. Given the multiple processes that control acid and pepsin secretion and defense and repair of the gastroduodenal mucosa, it is likely that the cause of ulceration differs between individuals. Acid and pepsin appear to be necessary but not sufficient ingredients in the ulcerative process. It is clear that the majority of gastric ulcers and a substantial number of duodenal ulcers do not have increased gastric acid secretion. Recent research has focused more on protection and repair of the stomach and duodenum.

Historically, our understanding of the pathophysiology of peptic ulcer disease focused on abnormalities in the secretion of gastric acid and pepsin, and on the suppression of acid as a treatment strategy. Today, gastric hypersecretion-associated with gastrinoma in Zollinger- Ellison syndrome, antral G-cell hyperplasia, an increase in parietal-cell mass, and a physiological imbalance between the antagonistic gastric hormones gastrin and somatostatin-is still an important issue in peptic ulcer disease. Moreover, it is known that cholinergic hypersensitivity and parasympathetic dominance are related to the stimulation not only of hydrochloric acid but also pepsin, which is often neglected as a cofactor in the development of erosive injury to the gastric mucosa. Psychologic stress, cigarette smoking, alcohol consumption, use of nonsteroidal anti-inflammatory drugs (NSAIDs) including aspirin, oral bisphosphonates, potassium chloride, immunosuppressive medications, and an age-related decline in prostaglandin levels have all been shown to contribute to peptic ulcer disease. It was, however, the isolation of *H.pylori* and its identification as the most important cause of peptic ulcer disease that led to exploration of the role of inflammation and its associated cytokine cascade in gastric acid secretion.

H. pylori evades attack by the host immune system and causes chronic, indolent inflammation by several mechanisms. *H.pylori* can damage the mucosal defense system by reducing the thickness of the mucus gel layer, diminishing mucosal blood flow, and interacting with the gastric epithelium throughout all stages of the infection. *H. pylori* infection can also increase gastric acid secretion; by producing various antigens, virulence factors, and soluble mediators, *H.pylori* induces inflammation, which increases parietal-cell mass and, therefore, the capacity to secrete acid. The *H. pylori* cytotoxin-associated gene CagA also has an important role: it interferes with gastric epithelial cell-signaling pathways, thereby regulating cellular responses and possibly contributing to apical junction barrier disruption, interleukin-8 secretion and phenotypic changes to gastric epithelial cells.

Understanding the pathophysiology of peptic ulcer disease is at something of a crossroads: mechanisms of injury differ distinctly between duodenal and gastric ulcers. Duodenal ulcer is essentially an *H.pylori* related disease and is caused mainly by an increase in acid and pepsin load, and gastric metaplasia in the duodenal cap²⁵. Gastric ulcer, least in Western countries, is most commonly associated with NSAID ingestion, although *H.pylori* infection might also be present²⁶. Chronic, superficial and atrophic gastritis predominate in patients with gastric ulcers, when even normal acid levels can be associated with mucosal ulceration²⁷. In both conditions, ulcer is

associated with an imbalance between protective and aggressive factors, with inflammation being a leading cause of this imbalance.

The isolation of *H.pylori* in the early 1980s was one of the most exciting advances in the history of peptic ulcer disease²⁸, and it has dramatically changed the management of peptic ulcer. Eradication of *H.pyori* infection is now the mainstay of treatment for peptic ulcer disease, and has resulted in very high ulcer healing rates and recurrence rates that have dropped dramatically, especially for individuals with a duodenal ulcer. The greater recognition of the role of NSAIDs and aspirin in gastrointestinal-tract injury has led to the development of therapeutic and preventive strategies that rely on the use of antisecretory drugs, the prostaglandin analog misoprostol, or selective cyclo-oxygenase (COX)-2 inhibitors(coxibs).

TESTS FOR DIAGNOSING HELICOBACTER PYLORI:

The following tests are used to diagnose *Helicobacter pylori* infection Testing may also be done after treatment to ensure the bacteria are fully eradicated.

1. **BREATH TEST:** A simple test called the carbon isotope-urea breath test (UBT) can identify up to 99% of people who harbor *Helicabacter pylori*. Up to 2 weeks before the test, the patient must descontiune taking any antibiotics, bismuth-containing agents such as Pepto-Bismol and proton-pump inhibitors (PPIs). As part of the test, the patient swallows a special substance containing *urea* (a compound in mammals metabolized from nitrogen) that has been treated with carbon atoms. *If Helicobacter pylori* are present, the bacteria convert the urea into carbon dioxide, which is detected and recorded in the patients's exhaled breath after 10 minutes. This test can also be used to confirm that *Helicobacter pylori* have been fully treated.
2. **BLOOD TESTS:** Blood tests are used ot measure antibodies to *Helicobacter pylori*, with results available in minutes. Diagnostic accuracy is reported at 80-90%. One such important test is called enzyme-linked immunosorbent assay (ELISA). An ELISA test of the urine is also showing promise in children.
3. **STOOL TEST:** A test to detect genetic fingerprints of *Hekucibacter pylori* in the feces appears to be as accurate as the breath test for initial detection of the bacteria and for detecting recurrences after antibiotic therapy. This test can also be used to confirm that the *Helicobacter pylori* infection has been fully treated.
4. **TISSUE BIOPSY:** The most accurate way to identify the presence of *Helicobacter pylori* is a tissue biopsy from the lining of the stomach. However, this is clearly an

invasive task and many patients are treated for *Helicobacter pylori* based on the above three noninvasive tests.

It should be noted that such tests are not as accurate as endoscopy, an invasive procedure, which is needed to confirm a diagnosis of *Helicobacter pylori*. The breath and stool tests, however, can be – are used most often. Other drugs that may be useful include H₂ blockers, such as Famotidine (Pepcid AC), Cimetidine (Tagamet), and Ranitidine (Zantac). Sucralfate is another drug used to heal ulcers and reduce the stomach upset caused by NSAIDs.

A number of alternative medications may be tried for people with chronic pain, to minimize the risk of ulcers associated with NSAIDs.

1. **COX-2 INHIBITORS (COXIBS):** Coxibs block an inflammation-promoting enzyme called COX-2. This drug class was initially thought to work as well as NSAIDs, while causing less gastrointestinal distress. However, following numerous reports of cardiovascular events, the FDA banned Rofecoxib (Celebrex) is still available, but patients should discuss with their doctor whether this drug is appropriate and safe for them. The use of Cox-2 inhibitors may provide a decrease in uncomplicated ulcers, but more serious events do not seem to be reduced by the use of these medications
2. **ARTHROTEC:** Arthrotec is a combination of Misoprostal and the NSAID Diclofenac. It may reduce the risk for gastrointestinal bleeding. This drug can cause miscarriage (abortion) at any stage of pregnancy and therefore should not be used during pregnancy.
3. **ACETAMINOPHEN:** Acetaminophen (Tylenol, Anacin-3) is the most common alternative to NSAIDs. Acetaminophen is inexpensive and generally safe. It poses far less of a risk of gastrointestinal problems than NSAIDs. Its dose should not exceed 4 grams (4,000mg); some studies suggest that ulcer risk is increased even in doses exceeding 2 grams (2,000 mg) a day, if the drug is used on a long-term basis. Patients who take high doses of Acetaminophen for long periods are also at risk for liver damage, particularly if they drink alcohol. It may pose a small risk for serious kidney complication in people with preexisting kidney disease, although Acetaminophen remains the drug of choice for patients with impaired kidney function.
4. **TRAMADOL:** Tramadol (Ultram) is a pain reliever that has been used as an alternative to opioids. It has opioid-like properties, but is not as addictive. However, dependence and abuse have been reported. It can cause nausea, but does not cause severe gastrointestinal problems, as NSAIDs can. Some patients experience severe itching. A

combination of Tramadol and Acetaminophen (Ultracet) provides more rapid pain relief than Tramadol alone and more durable relief than Acetaminophen alone. Side effects are the same as for each of these agents.

5. If continuation of NSAIDs is necessary, the lowest possible dose should be used.

PROTON-PUMP INHIBITORS(PPIs):

ACTIONS AGAINST ULCERS:

PPIs are the drugs of choice for managing patients with peptic ulcers, regardless of the cause. They suppress the production of stomach acid by blocking the gastric acid pump --- the molecule in the stomach glands that is responsible for acid secretion³⁰.

PPIs can be used either as part of a multidrug regimen for *Helicobacter pylori* or alone for preventing and healing NSAID-caused ulcers. They are also useful in treating ulcers caused by Zollinger-Ellison syndrome. They are considered to be more effective than H2 blockers.

STANDARD BRANDS:

Most PPIs are available by prescription as oral drugs. There is no evidence that one brand of PPI works better than another. Brands approved for ulcer prevention and treatment include:

- 1). Omeprazole (generic, Prilosec OTC)
- 2). Esomeprazole (Nexium)
- 3). Lansoprazole (Prevacid)
- 4). Rabeprazole (Aciphex)

H₂ BLOCKERS:

H₂ blockers interfere with acid production by blocking histamine, a substance produced by the body that encourages acid secretion in the stomach. H₂ blockers were the standard treatment for peptic ulcers until proton pump inhibitor and antibiotic regimens against *H. pylori* were developed. These drugs cannot cure ulcers, but they are useful in certain cases. They are effective only for duodenal ulcers, however.

Four H₂ blockers are currently available over-the-counter in the U.S.: Famotidine (Pepcid AC), Cimetidine (Tagamet), Ranitidine (Zantac), and Nizatidine (Axid). All have good safety profiles and few side effects. There are some differences between these drugs:

1. FAMOTIDINE (PEPCID AC):

Famotidine is the most potent H₂ blocker. The most common side effect is headache, which occurs in 4.7 % of people who take it. Famotidine is virtually free of drug interactions, but it may have significant adverse effects in patients with kidney problems.

2. CIMETIDINE (TAGAMET):

Cimetidine has few side effects; about 1% of people taking Cimetidine experience mild temporary diarrhea, dizziness, rash, or headache. Cimetidine interacts with a number of commonly used medications, including phenytoin, theophylline, and warfarin. Long-term use of excessive doses (more than 3 grams a day) may cause impotence or breast enlargement in men. These problems resolve after the drug is discontinued.

3. RANITIDINE (ZANTAC):

Ranitidine interacts with very few drugs. In one study, Ranitidine provided more pain relief and healed ulcers more quickly than Cimetidine in people younger than age 60, but there was no difference in older patients. A common side effect of Ranitidine is headache, which occurs in about 3% of people who take it.

4. NIZATIDINE (AXID):

Nizatidine is nearly free of side effects and drug interactions.

TREATMENT OF H. PYLORI INFECTION:

Ultrastructural studies have shown that following an exposure to bismuth preparations *H. pylori* detaches from the gastric epithelium and is lysed within 30-90min. In spite of the clinical success of *H. pylori* with oral bismuth, recrudescence is extremely common following monotherapy with bismuth. Therefore, it seems obvious that short term treatment with bismuth alone is inadequate for *H. pylori* infection. Chronic administration of bismuth for the treatment of *H. pylori* is sensitive to a wide range of anti-microbials including ampicillin, erythromycin, tetracyclines, ciprofloxacin, ofloxacin, metronidazole, tinidazole and nitrofurantoin. But, monotherapy with antibiotics has been shown to lead to the rapid onset of resistant organisms in some circumstances. Acquired resistance to quinolones has been demonstrated in *H. pylori* after ofloxacin and ciprofloxacin. The inadequacy of single agent therapy has led some workers to propose that a combination of bismuth with antibiotics, may provide the optimum approach. Combination therapy has provided the best results to date. Eradication rates vary from 40% with TDB and amoxicillin to 80% with TDB and metronidazole. Triple therapy has been found still more effective and 96% of patients will eradicate the organism with TDB, metronidazole and

tetracycline (or amoxicillin) combination. Further, it is reported that the efficacy of many antibiotics is pH dependent. Hence , there is a rationale for combination of acid inhibitory drugs with antibiotics in the treatment of *H.pylori* infection. A controlled study of combination therapy with omeprazole and amoxicillin in patient with duodenal ulcer showed eradication of *H.pylori* in 82% of patients. Over the past five years, several workers have suggested that eradication of *H.pylori* significantly reduces the rate of duodenal ulcer relapse. Recently, National Institute of Health (NIH) Consensus Conference recommended H.pylori eradication regimen as the first line medical therapy for patients with peptic ulcer disease who are *H.pylori* positive .

THERAPY AND PREVENTION OF NSAID ASSOCIATED ULCERS:

Ulcers associated with NSAIDs usually heal spontaneously when the NSAIDs are withdrawn. Even with the continued use of NSAIDs, ulcers may heal spontaneously. Limited studies suggest that H₂ blockers, prostaglandins, and omeprazole accelerate healing as compared to placebo. Omeprazole is emerging as the most promising agent for troublesome ulcers, especially when NSAIDs cannot be discontinued. A few trials have tested regimens for the prophylactic treatment of NSAID-associated ulceration. A three month trial indicated that misoprostol prevented NSAID-associated gastric ulcer, but data is lacking on the efficacy of PGs in preventing NSAID-associated duodenal ulcer. One short term study proved higher efficacy of PGs than H₂ blockers in preventing NSAID-associated gastric damage whereas H₂ blockers were effective in preventing duodenal damage.

TREATMENT OF REFRACTORY ULCERS:

Regardless of the mode of therapy, the healing of ulcers is time dependent; 90 to 95 percent of all ulcers heal if therapy is continued for 12 weeks. After that, ulcers or symptoms (or both) can be considered refractory to therapy. Since persistent symptoms may not be due to persistent ulcer disease, endoscopy is essential to establish the diagnosis. If persistent ulceration is found on endoscopy, a few possibilities ulceration is found on endoscopy, a few possibilities warrant consideration. Poor patient compliance, use of NSAIDs, smoking and gastrinoma are common causes of refractory ulceration. The role of *H.pylori* infection in refractory ulceration also remains uncertain. A 40mg dose of omeprazole or a full dose antisecretory therapy will probably have to be continued to maintain the healing of refractory ulcers, since it does not alter the natural history of ulcer disease. Treatment with bismuth or the eradication of *H.pylori* (or both) deserves consideration for patients who do not respond to conventional therapy.

RECURRENCE OF DUODENAL ULCERS:

Duodenal ulcers recur in 70 to 90% of patients within 1 year of the cessation of drug therapy. The evidence suggests that the rate of recurrence differs depending on the initial therapy used.

The relapse rates are higher after healing with H₂ antagonists as compared to tri-potassium dicitrato bismuthate therapy. Maintenance therapy with an H₂-receptor antagonists reduces this rate to 40%. Patients with complications of recurrence or those taking NSAIDs, and those who have pulmonary, renal or heart diseases are considered to be at a higher risk and should be placed on maintenance therapy to avoid recurrence. A single night time administration of H₂ antagonist at a dose of one half of the normal therapeutic dose is appropriate. This should be continued for one year. Sucralfate, 1g at bedtime, is also an effective maintenance regimen. Patients should be then reassessed for further therapy. Patients who have a minimal risk and who experience a recurrence should be reevaluated and reassigned to maintenance therapy. Maintenance therapy is also appropriate for recurrent gastric ulcer, but the response rate may not be as high as that for duodenal ulcer.

MEDICINAL PLANTS FOR THE TREATMENT OF ANTIULCER:

In this modern era also 75-80% of the world populations still use herbal medicine mainly in developing countries, for primary health care because of better cultural acceptability, better compatibility with the human body and lesser side effects. The chemical constituents present in the herbal medicine or plant are a part of the physiological functions of living flora and hence they are believed to have better compatibility with human body.

Natural products from plants are a rich resource used for centuries to cure various ailments. The use of natural medicine in the treatment of various diseases like peptic ulcer is an absolute requirement of our time. Therefore, alternative approach in recent days is the research of medicaments from traditional medicine. The use of phyto constituents as drug therapy to treat major ailments has proved to be clinically effective and relatively less toxic than the existing drugs and also reduces the offensive factors serving as a tool in the prevention of peptic ulcer.

ANTI ULCER ACTIVITY OF THE SOME IMPORTANT MEDICINAL PLANTS:

Jasminum grandi florum, Anogeissus latifolia, Alchornea castaneaefolia, Uleria salicifolia, solanum nigrum, Oimum sanctum, Scoparia dulcis, Bry sonima crassa, Asparaagus racemosus, centaurs Solstitialis, Anacardium Occidentale, callophyllum brasiliense, Rhizophora mangle, Larrea diraricata, Hemidesmus indicus, Spaartium junceum, Amomum subulatum, Aloe vera, Butea frondosa, Capsicum annum, curcuma longa, Desmostachya bipinnata, Exocoecaria

agallocha, mangifera indica, morus alba, Ocimum Sanctum, Panax ginseng, Piper betel, Polyalthia longifolia, Rhizophora mangle, Sapindus trifoliatus, SOLLANUM NIGURM, Syzygium aromaticum, Terminalia chebula, Triticum aestivum, Vinca minor, Zingiber officinalis, Ficus religiosa, Hibiscus roasainensis, Indigofera tinctoria, Lawsonia alba, Myrtus Communis, Momordica charantia, carica papaya, Adansonia digitata, Allium sativum, Psidium gujava, Rhuscoriaria, Sesbania grandiflora, Shorea robusta.

CONCLUSION:

Preliminary phytochemical screening of this medicinal plants identified the presence of important secondary metabolites like flavonoids and tannins. There are several botanical products with potential therapeutic applications because of their high efficacy and low toxicity. These plants provide leads to find therapeutically useful compounds, thus more efforts should be made towards isolation and characterization of the active principles and their structure active relationship. From this study, it is clear that the medicinal plants play a vital role against various diseases.

REFERENCES:

- 1) Dashputre NL, Naikwade NS, (2011). Evaluation of Anti-Ulcer Activity of Methanolic Extract of *Abutilon indicum* Linn Leaves in Experimental Rats. International Journal of Pharmaceutical Sciences and Drug Research 3(2): 97-100.
- 2) Izzo A, Borrelli F. (2000). The Plant Kingdom as a Source of Anti-ulcer Remedies. *Phytother Res* 14:581-591.
- 3) Baron J, Calam J. (2001). ABC of the Upper Gastrointestinal Tract: Pathophysiology of Duodenal and Gastric Ulcer and Gastric Cancer. *B M J* 323:980-982.
- 4) Manan MJA, Khan M, Safwan AKM, Hussain SA, Zakaria ZA. (2011). Anti-ulcer activity of *Ficus religiosa* stem bark Ethanolic extract in rats. *Journal of Medicinal Plants Research* 5(3): 354-359.
- 5) Nayaka H, Nanjundaiah M, Siddaraju (2011). *et al.* Gastroprotective Effect of Ginger Rhizome (*Zingiber officinale*) Extract: Role of Gallic Acid and Cinnamic Acid in H⁺,K⁺ATPase/H. pylori Inhibition and Anti-Oxidative Mechanism. Hindawi Publishing Corporation Evidence-Based Complementary and Alternative Medicine 1(1):1-13.

- 6) Vanisree AJ, Shyamala DCS, Mahendran P. (2002). The Antiulcer Activity of *Garcinia cambogia* Extract against Indomethacin Induced Gastric Ulcer in Rats. *Phytotherapy Research* 16:80-83.
- 7) Srivastava DP, Rajani GP, Gupta N, Sharma RK, Mandal S.(2011). Antiulcer and Anti inflammatory activity of fresh Leaves Extracts of *Polyalthia Longifolia* In Rats. *International Journal of Drug Development & Research* 3(1): 531-359.
- 8) Vinay SC, Pushpesh KM, Rakesh M, Dhamani P, Gautam P.(2005). *Allophylus serratus*: a plant with potential anti-ulcerogenic activity. *Journal of Ethnopharmacology* 99:361-366.
- 9) Gaikwad DK, Chavan RB. (2013). Antibacterial Activity of Medicinally Important Two Species of *Allophylus-Allophylus cobbe* (L). Raesch. & *Allophylus serratus* (Roxb). Kurz. *Journal of Pharmacognosy and Phytochemistry* 2(1).
- 10) Naibedy C, Manmeet K, Devendra M, Rakesh M, Rashmi P, Preeti R *et al.*(2010). Anti-osteoporotic constituents from Indian medicinal plants, *Phytomedicine* 17(13):993-999.
- 11) Sadilova E, Stintzing FC, Carle R. (2006). Anthocyanins, colour and antioxidant properties of eggplant (*Solanum melongena* L.) and violet pepper (*Capsicum annuum* L). peel extracts. *Z Naturforsch C* 61:527-535.
- 12) Dumpierrez AG, Blanco C, Alvarez M, Ortega N, Castillo R, Carillo T. (1998). *Carica papaya*, Pollen allergy, *Ann Allergy. Asthma Immunol* 81(2):171-175.
- 13) Ghongane BB, Rahul K. (2011). Evaluation of anti-ulcer activity of *Curcuma longa* in rats. *Journal of Advances in Pharmacy and Healthcare Research* 1(2).
- 14) Chandra D, Gupta SS.(1972). Anti-inflammatory and anti-arthritis activity of volatile oil of *Curcuma longa* (Haldi). *India J Med Res* 60(1):138-142.
- 15) Awaad AS, Maitland DJ, Soliman GA, Mohamed NH. (2008). Anti-ulcerogenic activity of extract and some isolated flavonoids from *desmostachia bipinnata* (L). *Stapf Rec Nat Prod* 2(3):76-82.
- 16) Ashok KBS, Girija K, Lakshman K, Medha M, Hegde LPV. (2010). Assessment of anti-diarrhoeal activity of *Desmostachya bipinnata* L. (poaceae) root extracts. *Boletin Latino Americano y del Caribe de plantas Medicinales y Aromaticas* 9(4):312-318.

- 17) Ramkumar L, Thirunavukkarasu P, Ramanathan T. (2009). Anti-ulcer Activity of *Excoecaria agallocha* bark on NSAID-induced Gastric Ulcer in Albino Rats. *Global Journal of Pharmacology* 3(3):123-126.
- 18) Ashraful A, Firoj A, Istat ZS, Nustat S.(2008). Anti-nociceptive and gastoprotective effect of the crude ethanolic extracts of *Excoecaria agallocha* Linn. *Turk J Pharm Sci* 5(3):143-154.
- 19) Lakshmi BVS, Mrityunjaya BP, Neelapu N, Muvvala S. (2012). Anti-ulcer Activity and HPTLC Analysis of *Mangifera indica* L. leaves international journal of Pharmaceutical and Phytopharmacological Research 1(4):146-155.
- 20) Vagdevi HM, Latha KP, Latha MS, Virupaxappa SB. (2012). Anti-inflammatory activity of *Mangifera indica* L. Var Rasapuri root extracts. *J Chem Pharm Res* 4:333-336.
- 21) Ali HM, Ahmed KA, Abdulla MA, Ismail S, Noor SM.(2009). Evaluation of the anti-ulcer activities of *Morus alba* extracts in experimentally-induced gastric ulcer in rats. *Biomed Res* 20(1): 35-39.
- 22) Jamshid M, Prakash RN. (2012). The histopathologic effects of *Morus alba* leaf extract on the pancreas of diabetic rats. *Turk J Biol* 36:211-216.
- 23) Bandna D, Dinesh K, Kamal J, Neha S.(2013). *Morus alba* Linn: a phytopharmacological review. *International Journal of Pharmacy and Pharmaceutical Sciences* 5(2).
- 24) Jeong CS, Hym JE, Kim YS. (2003). Ginsenoside Rb1: the antiulcer constituent from the head of *Panax ginseng*. *Arch Pharm Res* 26:906-911.
- 25) Chan CP, Lei D, Wang YJ. (2003). Antioxidative and antiplatelet effects of aqueous inflorescence *Piper betle* extract. *J Agric Food Chem*.
- 26) Waris G, Ahsan H. (2006). Reactive oxygen species: role in the development of cancer and various chronic conditions. *J Carcinog* 5(14).
- 27) Murakami A, Ali AM, Ohigashi H, Koshimizu K, Mat-Salleh K. (2000). Screening for the *in vitro* anti-tumor-promoting activities of edible plants from Malaysia. *Biotech Biochem* 64:9-16.
- 28) Geetha G, Jessi KVK, Malai RP, Narasimhan S.(2008). Evaluation of Anti-ulcer activity of *Polyalthia longifolia* Sonn Thwaites in experimental animals. *India J pharmacol* June 40(3).

- 29) Dharmaraj S, Lachimanan YL, Subramaniam D, Subramanion LJ, Sreenivasan S, Soundararajan V.(2013). Polyalthia longifolia Agents Remedy to Explore for Novel Therapeutic Agents Research. Journal of Pharmaceutical Biological and Chemical Sciences 4(1).
- 30) Antonia RM, Arturo E, Betty M, Caden S, Luz MSP.(2010). Pharmacological and toxicological evaluation of *Rhizophora mangle* L., as a potential antiulcerogenic drug: Chemical composition of active extract. Journal of Pharmacognosy and Phytotherapy June 2(4): 56-63.
- 31) Quilez A, Berenguer B, Sanchez LM. (2006). Protective and antioxidant effects of *Rhizophora mangle* L. against NSAID induced gastric ulcers. Journal of Ethno pharmacology, 103(2):194-200.
- 32) Branford-white C, de Armas E, Marrero E, Fern Andez O, sarracent Y.(2005). Efficacy of *Rhizophora mangle* aqueous bark extract (RMABE) in the treatment of aphthous ulcers: a pilot study. Current Medical Research and Opinion 21(11): 1711-1715.
- 33) Kishore DV, Pinto J, Mini KV. (2011). Anti-ulcer activity of methanolic and aqueous extracts of leaves of *Sapindus trifoliatius* L. international Journal of Pharmaceutical Sciences 6(1):25-27.
- 34) Chaitanya SK, Sarvani M, Sri lakshmi S.(2011). Ant helminitic activity of *Sapindus trifoliatius* seed extract. International Journal of Pharmacy and Technology 3(1):1603-1608.
- 35) Aastha S, Jaya D, Ruby K, Rajani C. (2012). *Solanum nigurm* with dynamic therapeutic role: a review. International Journal of Pharmaceutical Sciences Review and Research Rev Res 15(1):65-71.
- 36) Okasha MAM, Abubakar MS, Fatihu MY, Magaji RA. (2008). Antiulcerogenic and anti-secretory activity of the n-butanol portion.
- 37) Olsen PS, Poulsen SS, Krikegaard P, Nexo E. (1984). Role of submandibular saliva and epidermal growth factor in gastric cytoprotection. *Gastroenterology* 87:103-108.
- 38) Whittle BJR, Oren-Wolman N, Guth PH. (1985). Gastric vasoconstrictor actions of leukotrienes C, PGF and thromboxane mimetic U-46619 on rat submucosal microcirculation in vivo. *Am J Physiol* 248:G560-586.

- 39) Schwartz K. (1910). Uber penetrierende Magen-und jejuna Geschwung. *Beitr Klin Chir* 76:96-128.
- 40) Hills BA, Buttler BD, Lichtenberger LM,(1983) Gastric Mucosal barrier: hydrophobic lining to the lumen of the stomach. *Am J Physiol* 244:G 561- G 568.
- 41) Blaser MJ.(1987) Gastric Campylobacter-like organisms, gastritis and peptic ulcer disease. *Gastroenterology* 93:371-383.
- 42) Dooley CP, Cohen H. (1988). The clinical significance of Campylobacter pylori. *Ann Intern Med* 108:70-79.
- 43) Parmar NS, Deasi JK(1994). Helicobacter pylori and gastroduodenal disease. *Indian Drugs* 31:175-182.
- 44) Rauws EA, Langenberg W, Houthoff HJ, Zanen HC, Tytgat GN. (1988). Campylobacter pyloridis-associated chronic active antral gastritis: a prospective study of its prevalence and the effects of antibacterial and antiulcer treatment. *Gastroenterology* 94: 33-40.
- 45) Wyatt JI, Rattbone GI, Dixon MF, Heatley RV. (1987). Campylobacter pyloridis and acid induced gastric metaplasin in the pathogenesis of duodenitis. *J Clin Pathol* 40:841-848.
- 46) Hazell SL, LeeA. (1986). Campulbacter pyloridis, urease, hydrogen ion back diffusion and gastric ulcers. *Lancet ii*: 15.
- 47) Rainford KD, Watkins J, Smith MJH.(1968). Aspirin and mucus. *J Pharm Pharmacol* 20: 941-948.
- 48) Danenport HW. (1966). Fluid produced by the gastric mucosa during damage by acetic and salicyclic acids. *Gastronenterology* 50:487.
- 49) Pggogrt CJ, Lewandowski LG,(1972). Wirkung Von Prostaglandin und aspirin auf die Magen ssekretion des laborfrettches. *Leber Magen Darm* 2: 142.
- 50) Whittle BJR, Higgs GA, Eakins KE, Moncada S, Vane JR. (1980). Selective inhibition of prostaglandin production in inflammatory exudates and gastric mucosa. *Nature* 284:271-273.
- 51) Siegel ML, McConnel RT, Cuatrecasas P,(1979). Aspirin like drugs interfere with arachidonate metabolism by inhibition of the 12-hydroperoxy 5,8,10,14-eicosatertracine acid peroxidase activity of the lip-oxygenase pathway. *Proc Natl Acad Sci, USA* 76:3774.

- 52) Mangla JC, Kim Ym, Rubulis AA. (1974). Adenyl cyclase stimulation by aspirin in rat gastric mucosa. *Nature* 250:61-64.
- 53) Bennett JR. (1972). Smoking and the gastrointestinal tract. *Gut* 13:658-665.
- 54) World Health Organization Technical Report (1975). Smoking and its effect on health. *Report of a WHO Expert Committee*. Series No.568, Geneva, WHO.
- 55) Doll R, Jones FA, Pygott F. (1958). Effect of smoking on the production and maintenance of gastric and duodenal ulcers. *Lancet* 1:657-552.
- 56) Tatiq M, Parmar NS, Ageel AM. (1986). Effect of nicotine and alcohol pretreatment on the gastric mucosal damage induced by aspirin, phenylbutazone and reserpine in rats. *Alcoholism : Clinical and Experimental Research* 10:213-215.
- 57) Eastwood GL.(1988). The role of smoking in peptic ulcer disease. *J Clin Gastroenterol* 10:S19-S23.
- 58) Muller-Lissner SA.(1986). Bile reflux is increased in cigarette smokers. *Gastroenterology* 90: 1205-1209.
- 59) Tovey FI, Tunstall M. (1975). Progress report-duodenal ulcer in black populations in Africa South of the Sahara. *Gut* 16:564-576.
- 60) Tovey FI. (1979). Peptic ulcer in India and Bangladesh. *Gut* 20:329-347.
- 61) Shay H, Gruenstein H, Siple H, Kwmarov SA. (1948). Protection of gastric mucosa of the rat against ulceration by prefeeding with protein hydrolysate. *Proc Soc Exp Biol Med* 69:369-372.
- 62) Tovey FI.(1972). A trial of rice bran as a supplement to polished rice in the treatment of duodenal ulcers. *J Christ Med Ass India* 47:312-313.
- 63) Malhotra SL. (1978). A comparison of unrefined wheat and rice diets in the management of duodenal ulcer. *Postgrad Med J* 54:6-9.
- 64) Jauaraj AP, Rees KR, Tovey FI, White JS. (1986). A molecular basis of peptic ulceration due to diet. *Br J Exp Path* 67:149-155.
- 65) Henke PG, Ray A.(1992). The limbic brain, emotions and stress ulcers. *Exp Clin Gastroenterol* 1(4): 287-292.
- 66) Ray A, Henke PG.(1991). The basolateral amygdale, dopamine and gastric stress ulcer formation in rats. *Bruin Research* 558:335-338.

- 67) Soll AH. (1990). Pathogenesis of peptic ulcer and implications for therapy. *N Engl J Med* 322:909-916.
- 68) Parmar NS. (1989). Anti-ulcer drugs: present status and new targets. *Indian Drugs* 26:381-387.
- 69) Burkhalter A, Frick OL.(1989). Drugs with important actions on smooth muscle: Histamine, Serotonin and the ergot alkaloids. In Basic and Clinical Pharmacology. Karzung, B.G. eds., Prentice Hall International Inc., p.207.
- 70) Robert A, Nezamis JE, Lancaster C, Hanchar AJ.(1979). Cytoprotection by prostaglandins in hypertonic NaCl and thermal injury. *Gastroenterology* 77:433-443.
- 71) Walt RP.(1992). Misoprotosal for the treatment of peptic ulcer and anti-inflammatory durg induced gastroduodenal ulceration. *N Engl J Med* ;327;1575;1580.
- 72) Meijer JL,Jansen JB,Biemon I,Kuijpers IJ,Lamers CB. (1994). Effect of enrostil on serum gastrin and pepsinogen A and C levels in patients on long term treatment with omeprazole. *Aliment Pharmacol Ther* 8(12): 221-227.
- 73) Lam SK Ching CK. (1994). Sucralfate in clinical practice *J Gastroenterol Hepatol* . 9 : 401-411.
- 74) Lam SK , Hu WHC, Ching CK.(1995). Sucralfate in Helicobacter pylori eradication strategies, scand *J Gastroenterology* ;(in press).
- 75) Duncan WAM, Parsons ME.(1980). Reminiscenes of the development of cimetidine . *Gastroenterology* 78; 620-625.
- 76) Jain SM.(1992). Modifications of experimentally induced gastric and duodenal ulcers by calcium channel blockers . *Ph.D.Thesis, Gujarat University, India*.
- 77) Tariq M, Parmar NS , Ageel AM.(1987). Gastric anti-secretory, gastric and duodenal anti-ulcer and cyto-protective properties of proglumide in rats . *J pharmacol Exp Therap* 241:602-607
- 78) Glavin GB, Szabo S.(1990). Dopamine in gastrointestinal disease.*Dig Dis Sci* 35:1153-1161.
- 79) Glavin GB. (1989). Activity of selective dopamine DA₁ and DA₂ agonists and antagonists on experimental gastric lesions and gastric acid secretion. *J Pharmacol Exp Therap* 251: 726-730.

- 80) Desai JK, Parmar NS. (1994). Gastric and duodenal anti-ulcer activity of sulpiride, a dopamine D₂-receptor antagonist in rats. *Agents and Actions* 42:149-153.
- 81) Lagenberg W, Rauws EAJ, Wijojokusumo A, Tytgat GHJ, Zanen HC. (1986). Identification of *Campylobacter pyloridis* by restriction endonuclease DNA analysis. *J Clin Microbiol* 24: 414-417.
- 82) Glupzynski Y, Labbe M, Burette A, Delmee M, Avesani V, Bruck C. (1987). Treatment failure of ofloxacin in *Campylobacter pylori* infection. *Lancet* i: 1096-1098.
- 83) George LL, Borody TJ, Andrews P, Devine M, Moore Jones D, Walton M, Brandl S.(1990). Cure of duodenal ulcer after eradication of *Helicobacter pylori*. *Med J Aust* 153: 145-149.
- 84) Rauws EAJ, Lagenberg W, Houthoff HI, Zanen HC, Tytgat GNG: (1988). *Campylobacter pyloridis*-associated chronic active antral gastritis; a prospective study of its prevention and the effects of antibacterial and anti-ulcer treatment. *Gastroenterology* 94: 33-40.
- 85) Westblom TU, Duriex DE.(1991). Enhancement of antibiotic concentrations in gastric mucosa by H₂-receptor antagonist. Implication for treatment of *Helicobacter pylori* infection. *Dig Dis Sci* 36: 25-28.
- 86) Rauws EAJ, TytgatGNG. (1990). Cure of duodenal ulcer associated with eradication of *Helicobacter pylori*. *Lancet* i 1233-1235.
- 87) Coghlan JG, Gilligan D, Humphries H, Mckenna D, Dookey C, Sweeney E, Keane C, O Morain C.(1987). *Campylobacter pylori* and recurrence of duodenal ulcers –a 12 months follow-up study. *Lancet* ii:1109-1111.
- 88) NIH Consensus Development Panel.(1994). *Helicobacter pylori* in peptic ulcer disease. *Jama*, 272: 65-75.
- 89) Walan A, Bader JP, Classen M. (1989). Effect of Omeprazole and ranitidine on ulcer healing and relapse rates in patients with benign gastric ulcer. *N Eng J Med* 320: 69-75.
- 90) McCarty Dm. (1989); Nonsteroidal anti-inflammatory drug induced ulcers: management by traditional therapies. *Gastroenterology* 96:662-674.

- 91) Ehsanullah RSB, Page MC, Tildesley G, Wood JR. (1988). Prevention of gastroduodenal damage induced by non-steroidal anti-inflammatory drugs: Controlled trial of ranitidine. *Br Med J* 297: 1017-1021.
- 92) Lanza FL, Aspinall RL, Swabb EA, Davis RE, Rack MF, Rubin A. (1988). Double-blind placebo controlled endoscopic comparison of the mucosal protective effects of misoprostol versus cimetidine on tolmetin-induced mucosal injury to the stomach and duodenum. *Gastroenterology* 95: 289-294.
- 93) Miller JP, Gfaraghar EB. (1987). Relapse of duodenal ulcer. Does it matter which drug is used in initial treatment? *Br Med J* 2: 779-780.
- 94) Sonowski RA. (1990). The changing spectrum of therapy for active peptic ulcer disease. *Modern Medicine* 58: 50-58.
- 95) Greer D. (2006). Peptic ulcer Disease-Pharmacological Treatment. *Hospital Pharmacist*, 13: 245-250.
- 96) Rutter P.(2005). Symptoms, Diagnosis and treatment: A Guide for Pharmacists and Nurses. Edinburgh: Elsevier Churchill Livingstone.
- 97) Le TH & Fantry GT. (2008). Peptic ulcer Disease. *eMedicine*, 17 July. Available on the web: http://www.emedicine.com/MED/topic_1776.htm(date accessed: 3 November 2008).
- 98) Boreham H. (2008). Peptic ulcer Treatment. *Pharmacy Update*, August/September, 1(4): 29-32.
- 99) Aebi, H. (1983). Catalase. In : Bergmeyer, H. ed. *Methods in enzymatic Analysis*. New York, Academic Press, 3:276-286.
- 100) Amar, A., and Sanyal, A.K.(1981). immobilization stress in rats: effects on rectal temperature and possible role of brain monoaminase in hypothermia. *Psychopharmacology*, (73: 157-160).
- 101) Ghosh, M.N. *Fundamental of Experimental Pharmacology*.(1984). 2nd ed. Calcutta, Scientific Book Agency, 153-159.
- 102) Nadakarni, A.K.(1973). *Indian Materia Medica*. Mumbai, Popular Prakashan, 1(1): 72.
- 103) Kumar, R.S., Sivakumar, T., Sundram, R.S., Sivakumar, P., Nethaji, R., Senthil, V. (2005). Anti-inflammatory and analgesic effects of *Careya arborea* stem bark in experimental models. 43-61.

- 104) Tipnis, H., Komarov, S.A., Fels, S.S., Meranze, D., Gruenstein, M., Sipler, H. (1945). A simple method for uniform production of gastric ulceration in the rat. *Gastroenterology*, 5: 43-61.
- 105) Walker, R., Edwards, C.(2003). *Clinical pharmacy and therapeutics*.3rd edn. Edinburgh: Churchill Livingstone International edition, 55.
- 106) Wallace, J., M. Miller and C. Keenan, (2000). Nitric oxide in mucosal defense:a little goes a long way. *Gastroenterology* , 119: 512-520.
- 107) Akah P.A., Orisakwe O.E., Gamanies K.S., Shittu A., (1998). Evaluation of Nigerian traditional medicines: 11. Effects of some Nigerian folk remedies on peptic ulcer, *J Ethnopharmacol*, 62(2): 123-127.
- 108) Bortolotti, M., Coccia, G., Grossi, G. & Miglioli, M., (2000-a). “The treatment of functional dyspepsia with red pepper”, *Aliment Pharmacol Ther.*, 16(6): 1075-1082.
- 109) Akhtar AH and Ahmad KU.(1995). Anti-ulcerogenic evaluation of the methanol extract of some indigenous medicinal plants of Pakistan in aspirin ulcerated rats. *J Ethnopharmacol*, 46:16.
- 110) Desai W.G. *Aushadhisangrah*, Shree Gajanan. (1975). Book Depot Prakashan, 1:223.
- 111) Sahani Y.P, Shrivastava D.N. and Gaidani S.N):(1990) Inhibition of gastric Ulcer by Indigenous medicine *Withania somnifera*. *Indian Veterinary medical Journal* 18:42-43.
- 112) Alan B, Thomson R, Mahachai V, Martin H, S., Fenton K, et al. (1985). Medical management of un complicated peptic ulcer disease”; In: William editors,
- 113) Lewis DA, Fields WN, Shaw GP. (1978). Factors in the etiology of chronic duodenal ulcer in Ibadan. *Trop Geogr Med.*, 30: 75-9.
- 114) Khan Kh. (2009). Roles of *Emblica officinalis* in Medicine-A Review. *Botany Research International* 2(4): 218-28.
- 115) Mohan Kumar M, Joshi MC, Prabha T, Dorababu, M, Goel RK. (2006). Effect of palantain banana on gastric ulceration in NIDDM rats: Role of gastric mucosal glycoproteins, cell proliferation, antioxidants and free radicals. *Indian J Exp Biol.*, 44: 292-99.

- 116) Maury PK, Jain SK, Nand Lal and Shashi Alok,(2012). A review on antiulcer activity. *Int J Pharma Sci & Res.*, 3:2487=93.
- 117) Martin MJ, La-Casa, C, Alarcon DeLa Lastra C, Cabeza J. Villegas I, Motilva, V. (1998). Antioxidant mechanism involved in gastriprotective effects of quercetin. *Z. Narutforsch. C.J. Biosci*, 53: 82-88.
- 118) Sairam K, Tao CV, Goel RK. (2001). Effect of *Centella asiatica* linn a physical and chemical factores induced gastric ulceration and secretion. *Indian J Exp. Biol.*, 39: 137-142.
- 119) Kirtikar KR, Basu BD. (1961). *Indian Medicinal Plants*. Edn 2, Lalit Mohan Basu, Allahabad, India, 314-315.
- 120) Davenport HW. (1968). Destruction of the gastric mucosal barrier by detergents and urea. *Gastroenterol*, 54: 175-180.
- 121) Garg GP, Nigam SK, Ogle CW. (1993). The gastric antiulcer effects of the leaves of the neem tree. *Planta Medica*, 59:215-217.
- 122) Alkofahi, A. and A.H. Atta,(1999). Pharmacological experimental ulcer models: Possible mechanism forscreening of the antiulcerogenic effects of some the inhibition of acid formation. *J. Ethnopharmacol., Jordanian Mecicinal Plants in rats. J.*, 104: 156-163.
- 123) Chakraborty, G.S., R.S. Badujarand and Antiulcerogenic potential of *Strychros potatorum* C.R. Pardeshi. (2009). Analgesic activity of chloroform Linn. Seed on aspirin plus pyloric ligation extract of *Caesalpinia pulcherrima*. *J. Pharm. Induce ulcers in experimental rats. Phytomedicine, Res.*, 2: 1199-1200. 14:360-365.
- 124) Chiang, L.C., W. Chiang, M.C. Liu and C.C. Lin,(2003). indomethacin induced gastric ulcer. *J. In vitro antiviral activities of Caesalpinia Ethnopharmacol.*, 1;70: 171-176.
- 125) Alkofahi A, Atta AH.,(1999). Pharmacological screening of the antiulcerogenic effects of some Jordanian Mecicinal Plants in rats, *J Ethnopharmacol*, 65:341-5.
- 126) Borelli F, Izzo AA., (2000). The plant Kingdom as a source of anti-ulcer remedies, *Phytother Res.*, 14:581-91.
- 127) Chattopadhyay RR., Bhattacharyya SK. (2007). Plant Review Terminalia chebula: An update *Pharmacog Rev.*, 1(1): 151-6.

- 128) Olsen PS et al.(1984). Role of submandibular saliva and epidermal growth factor in gastric cytoprotectio. *Gastroenterology* 87;103-108.
- 129) Zaghlool SS, et al. (2015). Comparison between the Protective Effects of Gamotidine, Ginger and Marshmallow on pyloric ligation-Induced peptic Ulcer in Rats. *J Bioequiv Availab* 7:170-178.
- 130) Rainsford KD.(1972). Prostaglandins and the development of gastric mucosla damage by anti-inflammatory drugs. In : prostaglandins and inflammation, by Rainsford KD and Hutchinson AW, Birkhauser, Basel:1.
- 131) Bennett JR.(1972). Smoking and the gastrointestinal tract.*Gut* 13:658-665.
- 132) Rosenblum J and Papamichael M.(2016). Combined Ultrasound and Electric Field Stimulation Aids the Healing of Chronic Pressure Ulcers. *J Gerontol Geriatr Res* 5:319.
- 133) Vilela LHR, et al.(2016). Pain Assessment in Patients with Venous Leg Ulcer Treated by Compression therapy with Unna’s Boot. *J Tissue Sci Eng* 7:71.
- 134) Connelly TM, et al.(2016). Genetic and Demographic Correlates of Quality of Life after Ileal Pouch Anal Anastomosis for Ulcerative Colitis. *J Inflamm Bowel Dis & Disord* 1:107.
- 135) Wyatt JJ, et al. (1987). *Campylobacter pyloridis* and acid induced gastric metaplasia in the pathogenesis of duodenitis. *J Clin Pathol* 40:841-848.
- 136) Mary SJ and Merina AJ. (2015). Gastroprotective effect of *Guttarda speciosa* against Ethanol induced Gastric Ulcer in Rats. *Med Aromat Plants* 5:224.
- 137) Chowdhury ATMM, et al.(2015). An Unusual Presentation of Ulcerative Colitis with Numerous Colon Polyps and Formation of Multiple Band and Septum Like Structures in the Colonic lumen. *J Hepatol Gastroint Dis* 2015;1:i102
- 138) Ismail AE, et al.(2015). role of autologous Bone marrow stem cell transplantation in the treatment of active Ulceration Colities . *J Stem Cell res Ther* 5:313
- 139) Sundlass NK, et al.(2015). Infliximab –induced linear Iga Bullous Disease in a patient with ulcerative colitis . *J clin case Rep* 5:238
- 140) Shivakumar singh P,et al. (2015). Documentation of Floklori Knowledge on Medicinal Plants Used in the Treatment of Mouth Ulcer in Kodangal Mandalam, Mahabubnagar District, Telangana, india. *J Bioanal Biomed* 7:174-179.

- 141) Banala N, et al.(2015). Design and Evaluation of Floating Multi Unit Mini Tablets MUMTS Muco Adhesive Drug Delivery System of Famotidine to Treat Upper Gastro Intestinal Ulcers. *J Pharmacovigil*. 3:179.
- 142) Goldring M, et al.(2015). The Help seeking Behaviours of Patients with Ulcerative Skin Lesions Before Consultation in Yurimaguas, Peru. *J Anc Dis Prev Rem* ;3:127.
- 143) Frimpong M, et al.(2015). Microscopy for Acid Fast Bacilli: A Useful but Neglected Tool in Routine Laboratory Diagnosis of Baruli Ulcer .*J Trop Dis* ;3:158.
- 144) Kumar A, et al.(2015). The Foot Care Process of Diabetic Patients With and Without Foot Ulcer Attending ATertiary Care Hospital in India. *J Stem Cell Res Ther* ;5:280.
- 145) Mani P, et al.(2015). Treatment and Replenishment of G.I. Tract with Combined Regimen Therapy CRT of Allopathic PPIs and Ayurvedic Aloe Vera Medicine in Peptic Ulcer Disease to counteract Relapse. *J Gastrointest Dig Syst* 5: 272.
- 146) Kadhim G, et al.(2015). Risk factors Associated with Peptic Ulcer Disease. *J Bioengineer & Biomedical Sci* 5:10.42.
- 147) Kryczka T and Grieb P. (2015). Supportive Treatment of Pressure Ulcers with Dietary Supplementation. *Clin Pharmacol Biopharm* 3:130.
- 148) Erdogan EI .(2014). Gastric Ulcers in a Patient with Percutaneous Endoscopic Gastrostomy. *J Gastroint Dig Syst* 4:213.
- 149) Van Vliet EP, et al. (2008). Inappropriate prescription of proton pump inhibitors on two pulmonary medicine wards. *Eur J Gastroenterol Hepatol* 20:608-612.
- 150) Shafi S, et al.(2011). Proton pump inhibitors-over-prescribed in a rural community? *Pak J Med Sci* 27:300-302.
- 151) Fock Km, et al. (2008). Proton pump inhibitors: do differences in pharmacokinetics translate into differences in clinical outcomes? *Clin Pharmacokinet* 47:1-6.
- 152) Hung OY, et al. (2011). Hypergastrinemia, type 1 Gastric Carcinoid tumors; Diagnosis and Management, *J Clin Oncol* 25:e713-5.
- 153) Bertleff MJ and Lange JF. (2010). Perforated peptic ulcer disease: a review of history and treatment. *Dig surg* 27: 161-169.

- 154) Lau JY, et al.(2011). Systematic review of the epidemiology of complicated peptic ulcer disease: incidence, recurrence, risk factors and mortality. *Digestion* 84: 102-113.
- 155) Thorsen K, et al. (2011). Trends in diagnosis and surgical management of patients with perforated peptic ulcer. *J Gastrointest Surg* 15: 1329-1335.
- 156) Gisbert JP, et al.(2004). Helicobacter pylori and perforated peptic ulcer prevalence of the infection and role of non-steroidal anti-inflammatory drugs. *Dig Liver Dis* 36: 116-120.
- 157) Manfredini R, et al. (2010). Seasonal pattern of peptic ulcer hospitalizations: analysis of the hospital discharge data of the Emilia-Romagna region of Italy. *BMC Gastroenterol* 10:37.
- 158) Svanes C, et al.(1998). Rhythmic patterns in incidence of peptic ulcer perforation over 5.5 decades in Norway. *Chronobiol Int* 15: 241-246.
- 159) Watts DD and Fakhry SM. (2003). Incidence of hollow viscus injury in blunt trauma: an analysis from 275,557 trauma admissions from the East multi-institutional trial. *J Trauma* 54:289-294.
- 160) Goh BK, et al.(2006). Perforation of the gastrointestinal tract secondary to ingestion of foreign bodies. *World J Surg* 30: 372-377.
- 161) Bianchini AU, et al. (1997). Duodenal perforation by a grenfield filter: endoscopic diagnosis. *Am J Gastroenterol* 92: 686-687.
- 162) Mao Z, et al. (2008). Duodenal perforations after endoscopic retrograde cholangiopancreatography: experience and management. *J Laparoendosc Adv Surg Tech A* 18: 691-695.
- 163) Zeb F, et al. (2009). Duodenal impaction/perforation of a biliary stent-a rare complication in the management of choledocholithiasis. *J Gastrointest Liver Dis* 18: 391-392.
- 164) FY Le, et al. (2001). Predicting mortality and morbidity of patients operated on for perforated peptic ulcers. *Arch Surg* 136: 90-94.
- 165) Kocer B, et al.(2001). Factors affecting mortality and morbidity in patients with peptic ulcer perforation. *J Gastroenterol Hepatol* 22: 565-570.

- 166) Bucher P, et al. (2007). Results of conservative treatment for perforated gastroduodenal ulcer in patients not eligible for surgical repair. *Swiss Med Wkly* 137:337-340.
- 167) Siu W, leong H, Law B. Chau Ch, Li AC, et al. (2002). Laparoscopic repair for perforated peptic ulcer: a randomized controlled trial. *Ann Surg* 235: 313-319.
- 168) Uccheddu A, et al.(2003). Surgery for perforated peptic ulcer in the elderly. Evaluation of factors influencing prognosis. *Hepatogastroenterology* 50: 1959-1958.
- 169) Tsugawa K, et al. (2001). The therapeutic strategies in performing emergency surgery for gastroduodenal ulcer perforation in 130 patients over 70 years of age. *Hepatogastroenterology* 48: 156-162.
- 170) Linder MM, et al. (2010). The Mannheim Peritonitis Index. An instrument for the intraoperative prognosis of peritonitis, *Chirurg* 58: 84-92.
- 171) Moller MH, et al. (2012). The peptic Ulcer perforation (PULP) score: a predictor of mortality following peptic ulcer perforation. A cohort study. *Acta Anaesthesiol Scand* 56: 655-662.
- 172) So JB, et al. (2000). Risk factors related to operative mortality and morbidity in patients undergoing emergency gastrectomy. *Br J Surg* 87:1702-1707.
- 173) SC Le, et al. (2002). Candida peritonitis due to peptic ulcer perforation: incidence rate, risk factors, prognosis and susceptibility to fluconazole and amphotericin B. *Diagn Micro Infect Dis* 44:23-27.
- 174) Boey J, et al. (1982). Bacteria and septic complications in patients with perforated duodenal ulcers. *Am J Surg* 143: 635-639.
- 175) Nomani AZ, et al.(2014). A new prognostic scoring system for perforation peritonitis secondary to duodenal ulcers. *J Pak Med Assoc* 64 : 50-56.
- 176) Malhorta Ak, et al. (2000). Blunt bowel and mesenteric injuries: the role of screening computed tomography. *J Trauma* 48: 991-1000.
- 177) Jacobs DG, et al. (1990). Peritoneal lavage white count: a reassessment. *J Trauma* 30:607.
- 178) Rozycki GS, et al. (1998). Surgeon-performed ultrasound for the assessment of truncal injuries. *Ann Surg* 228:557.

- 179) Halley F.M.(1991). Self-regulation of the immune system through biobehavioral strategies. *Bio-feedback Self Regul* 16: 55-74.
- 180) Murakami, N., Nakai, Y., Fukunaga, M. et al.:(1999). Psychosomatic study on factors associated with development and recurrence of peptic ulcer. Multivariate analysis. *Shin Shin I* 39: 421-428.
- 181) Nakai, Y., Murakami, N., Fukunaga, M. et al.:(1998). Preparation of diagnostic criteria for psycho-somatic disease and their application. Gastric and duodenal ulcers. *Nippon Shinryo-naika Gakkai-shi (Journal of the Japanese Society of Psychosomatic internal Medicine)* 2: 119-121. (in Japanese).
- 182) Nakai, Y.: (1998). Miyagi, H., Karibe, M. et al.: Peptic ulcer and stress. *Sangyo Sutoresu Kenkyu (Industrial Stress Study)* 6: 189-195.
- 183) Nakai, Y.: (1998). Discussion on the results of a questionnaire study on "Health and Stress". *Nippon Iji Shinpo (Japan Medical Journal)* 3895: 43-49.
- 184) Nakai, Y.: (2000). The role of psychosomatic medicine in the 21st century with respect to life-style diseases. *Nippon shinryo-naika Gakkai-shi (Journal of the Japanese Society Of Psychosomatic Internal Medicine)* 4: 113-120.
- 185) Khanna D, Parle M. (2011). Clove: a champion spice. *International journal of research in ayurveda and pharmacy* 2(1).
- 186) Lumar KJ. (2006). Effect of geographical variation on contents of tannic acid, gallic acid, chenulinic acid and ethyl gallate in *Terminalia chebula*. *Natural Products* 2(3-4):170-75.
- 187) Raju D, Ashisj K, Ilango K, Chitra V. (2009). Evaluation of antiulcer activity of methanolic extract of *Terminalia chebula* fruits in experimental rats. *J Pharm Sci Res* 1(3):101-107.
- 188) Prakash CG. (2012). Biological and pharmacological properties of *Terminalia chebularetz*. (Haritaki)-an overview. *International Journal of Pharmacy and Pharmaceutical Sciences* 4(3).
- 189) Verlagsanstalt. (1974). *PDR for Herbal Medicines*, Edn 3, LaGow B, Austria, 1-11.
- 190) Banulova A, Machova J, Nosalova V. (1993). Protective action of vinpocetine against experimentally induced gastric damage in rats. *Arzneimittel forschung* 43:981-985.

191) Ghosh AK, Mullick HI, Banerjee J., Banerjee S. (2011). *Zingiber officinale*: a natural gold. International Journal of Pharma and Bio Sciences 2(1).